

ROLE OF THE PROSTAGLANDIN E₂ RECEPTOR EP1 IN
HYPERTENSIVE END-ORGAN DAMAGE

By

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LIST OF ABBREVIATIONS

AA	Arachidonic acid
AAALAC	Association for accreditation and assessment of laboratory animal care
ACEi	Angiotensin converting enzyme inhibitor
ACR	Albumin:creatinine ratio
ALT	Alanine transaminase
Ang II	Angiotensin II
ARB	Angiotensin receptor blocker
AT	Angiotensin II receptor
BP	Blood pressure
BUN	Blood urea nitrogen
[cAMP] _i	Intracellular cyclic adenosine monophosphate
CHF	Congestive heart failure
CNS	Central nervous system
COX	Cyclooxygenase
cPLA ₂	Cytosolic phospholipase A ₂
CVD	Cardiovascular disease
DGLA	Dihomo-gamma-linolenic acid
DN	Diabetic nephropathy
DOCA	Deoxycorticosterone acetate
EF	Ejection fraction
EGFR	Epidermal growth factor receptor

eNOS	Endothelial nitric oxide synthase
EP	E-prostanoid receptor
EPA	Eicosapentaenoic acid
FS	Fractional shortening
ESRD	End-stage renal disease
GBM	Glomerular basement membrane
GFR	Glomerular filtration rate
GPCR	G protein-coupled receptor
H/E	Hematoxylin and eosin
ICV	Intracerebroventricular
IVS	Intraventricular septum
JNC	Joint National Committee
Kim-1	Kidney injury molecule-1
LVID	Left ventricular interior diameter
LVPW	Left ventricular posterior wall
LTs	Leukotrienes
MAP	Mean arterial pressure
mPGES	Microsomal prostaglandin E synthase
mRNA	Messenger ribonucleic acid
Ngal	Neutrophil gelatinase-associated lipocalin
Nphx	Uninephrectomy
NSAIDs	Non-steroidal anti-inflammatory drugs
PGs	Prostaglandins

PGE ₂	Prostaglandin E ₂
PGIs	Prostacyclins
PTX	Pertussis toxin
RAAS	Renin-angiotensin-aldosterone system
SBP	Systolic blood pressure
STZ	Streptozotocin
TX	Thromboxanes

CHAPTER I

INTRODUCTION

History of Hypertension

In 1733, the idea of blood pressure (BP) was first credited to British veterinarian Stephen Hales who observed the pulsation of blood when inserting a pipe into the artery of a horse. Hales concluded that pressure must be pushing the blood. While blood pressure was an accepted idea, it was not until 1847 when the first recording was made using a kymograph by Carl Ludwig. Many years later, Samuel Karl Ritter von Basch invented the sphygmomanometer, which was further improved to a mercury filled sphygmomanometer with an inflatable cuff in 1896 by Scipione Riva-Rocci. Massachusetts General Hospital was among the first to incorporate its use into routine checkups, courtesy of neurosurgeon Harvey Cushing. Although the risks of high blood pressure were acknowledged by insurance companies, it was not until the 1960s that the medical community had wide acceptance of these dangers. Until this time many experienced and prominent cardiologists considered high blood pressure to be an important compensatory phenomenon that should be left alone.

The greatest danger to a man with high blood pressure lies in its discovery, because then some fool is certain to try and reduce it. – J.H. Hay, 1931

Hypertension may be an important compensatory mechanism which should not be tampered with, even were it certain that we could control it.

– Paul Dudley White, 1937

President Franklin Delano Roosevelt suffered and eventually died as a result of high blood pressure, and the progression of his disease is an eloquent example of the natural history of untreated hypertension (1). In 1937, 54 year old Roosevelt had a BP of 162/98 mm Hg, though received no treatment to reduce his hypertension. Within three years, his blood pressure had risen to 180/88 mm Hg. In 1941, his physician prescribed phenobarbital and massages, as his blood pressure had risen to 188/105 mm Hg. Over the next three years, President Roosevelt suffered shortness of breath, drowsiness, and lethargy. At the request of Roosevelt's daughter, cardiologist Howard G. Bruenn examined President Roosevelt finding strong evidence of congestive heart failure (CHF) including pulmonary edema, an enlarged heart, left ventricular hypertrophy and proteinuria. He treated Roosevelt by reducing his alcohol and cigarette use, limiting his activity, administering digitalis and implementing a low salt diet which helped to improve his symptoms. That year Roosevelt's blood pressure was as high as 230/140 mm Hg. In 1945, his blood pressure had risen to 260/150 and on April 12, 1945 while sitting for a portrait President Roosevelt lost consciousness. His blood pressure had risen to > 300/190 mm Hg and he died of a cerebral hemorrhage.

At this time there were very few options with regards to the treatment of hypertension and all had severe side effects. The low salt diet had been suggested as a treatment in 1922 by Allen and Sherrill, though this received very little attention in the United States (2). The Kempner rice diet published in 1944 was demonstrated to significantly impact BP although this diet had very low compliance (3). Other more radical therapies included inoculation with fever producing agents such as typhoid bacilli, or surgical procedures like sympathectomy and adrenalectomy (4). The herb *veratrum viride* showed great promise for lowering blood pressure but had a very narrow therapeutic window. Dr. Edward Freis, a leader in antihypertensive treatment, began using the antimalarial drug pentaquine as a hypotensive drug (1).

The 1950s were a time of great struggle with regards to acceptance of the dangers and need to treat hypertension in the medical community. In 1955, Perera of Columbia University published conclusions from a study of 300 patients suggesting hypertension is relatively benign and the label overused (5). Meanwhile, clinical evidence began accumulating demonstrating the benefit of lowering blood pressure, and the search for novel therapeutic agents boomed. Alternatives to the dangerous surgical procedures were discovered including sympathetic nerve blockers (phenoxybenzamine), ganglionic blockers (hexamethonium, pentolinium), peripheral adrenergic inhibitors (guanethidine), catecholamine reduction (reserpine) and vasodilators (hydralazine). In 1957, a clinical trial began for a major antihypertensive agent still used today, chlorothiazide (6).

In the 1960s, the Framingham Heart Study showed a correlation between blood pressure and heart attacks, congestive heart failure, stroke and kidney damage (7). The first Veterans Administration study showed antihypertensive therapy in patients with diastolic BP > 105 mm Hg was able to reduce the incidence of stroke, CHF, and kidney damage (8). Propranolol began its clinical use in the early 1960s as the first of the beta blocker family. The National High Blood Pressure Education Program was started as a way of informing the medical community and public of the dangers of high blood pressure and methods to treat it. In 1977, the first report of the Joint National Committee on Prevention, Detection, Evaluation, and Treatment of High Blood Pressure (JNC) was published, setting the standards for blood pressure diagnosis and treatment (9). These standards are updated regularly and JNC 8 is currently in development.

Pathophysiology and Consequences of Hypertension

Still today, hypertension is a prevalent disease which causes substantial morbidity and mortality in humans, and affects approximately 33 % of adults in the United States (10). Roughly one-third of afflicted individuals are receiving treatment for hypertension, though only one-half of treated patients have their blood pressure under control. Thus, prevention and treatment of hypertension is an important public health goal.

Hypertension can be classified into several categories based on the degree of blood pressure elevation (Table 1.1) ranging from prehypertension (systolic 120-139 mm Hg/ diastolic 80-90 mm Hg) to Stage 2 hypertension (systolic \geq 160 mm Hg/ diastolic \geq 100 mm Hg) (11). Alternatively, hypertension can be designated either essential or secondary, based on the root cause of the high blood pressure. About 95 % of hypertensive adults have essential hypertension, meaning no secondary cause was discovered. Secondary causes of hypertension can include renal, vascular, endocrine, neurogenic and drug or toxin induced hypertension.

Following chronic elevated blood pressure, target end-organ damage to the aorta and small arteries, heart, kidneys, and retina can be observed. Blood pressure is the product of cardiac output (stroke volume x heart rate) and systemic vascular resistance. Thus, it is reasonable to conclude that patients

Table 1.1 Classification of Hypertension

	Systolic BP (mm Hg)	Diastolic BP (mm Hg)
Normal	< 120	< 80
Prehypertension	120-139	80-90
Stage 1	140-159	90-99
Stage 2	\geq 160	\geq 100

For adults 18 years or older, based on recommendation of the JNC VII.

with hypertension may have increased cardiac output, increased vascular resistance, or both. Early in the course of disease, cardiac output is often elevated. Over time, vascular stiffness and increased systemic vascular resistance plays a greater role (12). This increases the afterload on the left ventricle, inducing left ventricular hypertrophy and dysfunction, which is an independent risk factor for sudden death and stroke. Coronary artery disease is another cardiac consequence of hypertension. At higher MAP the heart requires more oxygen to constrict, and lipid plaques in the arteries decrease coronary oxygen supply. This combination leads to myocardial ischemia and infarction. Congestive heart failure also occurs with chronic pressure overload. Often patients present with diastolic dysfunction, which eventually leads to systolic failure and cardiac congestion.

Renal disease is an additional complication of hypertension. Hypertension-induced vascular lesions can also result in ischemia to the nephrons, or hyperperfusion of the glomerulus can damage the glomerular capillaries themselves. Patients initially present with microalbuminuria, and pathology reveals glomerulosclerosis, which eventually leads to ischemic renal tubules. The risk of end-stage renal disease (ESRD) is greater in hypertensive black patients than white patients, even when blood pressure is properly regulated. Hypertension also increases the risk of ESRD in patients with diabetic nephropathy.

Hypertension-induced damage to the vasculature is a common feature observed in many complications of the disease. In addition to coronary artery

disease, stroke and renal disease, the damage to the vasculature of the eye or aorta can also have serious consequences. Increased pressure in the aorta can cause a weakening of the vessel wall leading to the formation of an aneurysm. Most patients with aortic aneurysms are asymptomatic until the aneurysm ruptures. This is a life-threatening event, and only 10-25 % of patients survive. The vasculature of the retina of the eye is also uniquely sensitive to damage. Chronic elevated pressure can result in narrowing of the arterioles with presentation of microaneurysm or macroaneurysm formation. These hypertensive retinopathies can result in loss of vision.

The regulation of blood pressure is vastly intricate and multifactorial. Many risk factors - both environmental and genetic - have been identified to increase BP including obesity, insulin resistance, high salt diet, aging, sedentary lifestyle, stress, and low potassium and calcium intake (13),(14). On a whole body-level, many more factors can influence blood pressure regulation including neurogenic control, vascular reactivity, the renin-angiotensin system, sodium and water retention, nitric oxide, adrenal steroids and eicosanoids – including prostaglandins (PGs).

Prostaglandins: Biosynthesis and Catabolism

Prostaglandins are biologically active lipid-derived autacoids generated by prostaglandin H synthase, commonly referred to as cyclooxygenase (COX). The COX enzymes are the target of non-steroidal anti-inflammatory drugs (NSAIDs).

PGs are part of the family of eicosanoids which includes prostaglandins, prostacyclins (PGIs), thromboxanes (TXs), and leukotrienes (LTs). Eicosanoids are derived from the 20-carbon essential fatty acids arachidonic acid (AA), eicosapentaenoic acid (EPA), or dihomo-gamma-linolenic acid (DGLA). Specifically, PGs are derived from AA which is cleaved from the plasma membrane by phospholipase A₂, the rate limiting step in PG biosynthesis. COX catalyzes the oxidation of AA to PGG₂ and reduction to PGH₂. The tissue-specific prostaglandin synthase enzymes then convert PGH₂ into one of the five principal prostanoids – PGD₂, PGE₂, PGF_{2α}, PGI₂ and TXA₂ (Figure 1.1).

Prostaglandins are ubiquitously produced and act in an autocrine or juxtacrine manner. Their actions can be halted by efficient catabolic pathways. When PGE₂ is administered intravenously, 95 % of PGE₂ is inactivated within the first pass through the lungs. This is achieved by β-oxidation, ω-oxidation by cytochrome P-450s, and PG-15-keto-reductase (15). PGE₂ is primarily inactivated by the conversion of the C-15 hydroxyl group into a ketone by NAD⁺ dependent 15-hydroxyprostaglandin dehydrogenase (16).

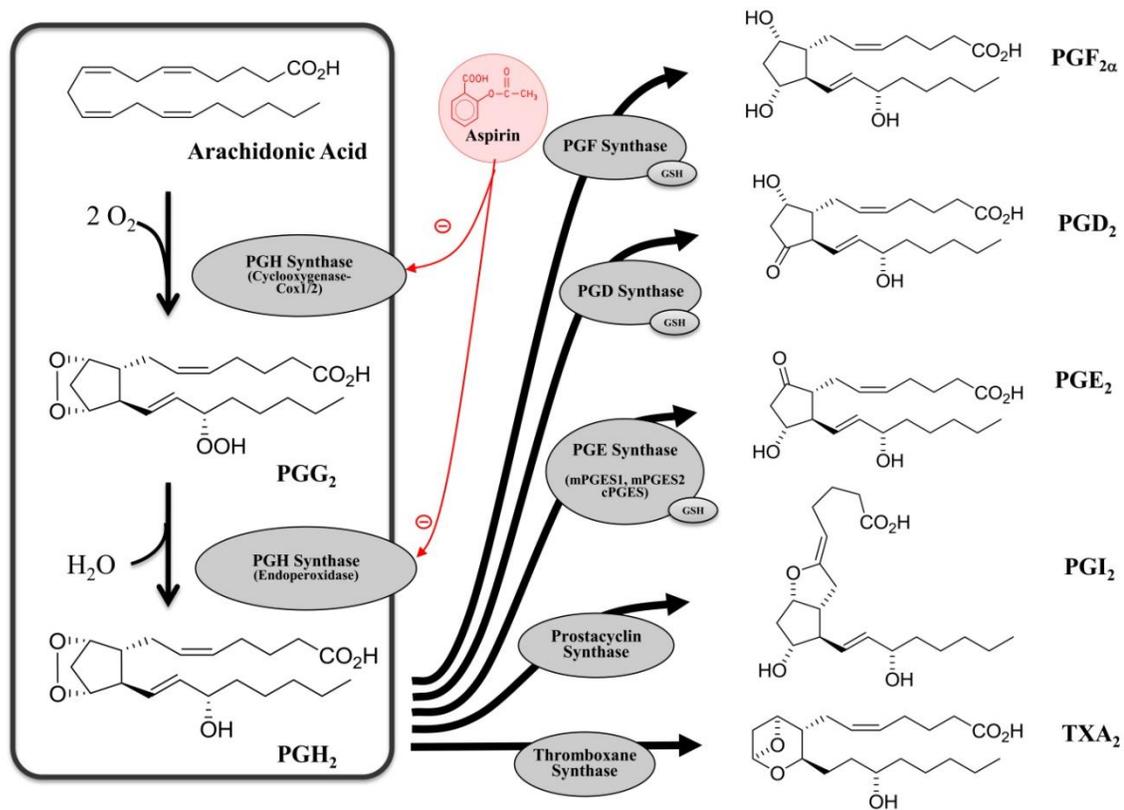


Figure 1.1 Biosynthesis of prostaglandins. This figure illustrates the formation of the five principal prostanoids from arachidonic acid.

PGE₂ Receptors: Pharmacological properties and signaling

PGE₂ mediates diverse and at times physiologically opposing effects, as is the case with regard to blood pressure regulation (17-19). This unique feature can be explained in part by the existence of four PGE₂ receptors, designated the E-Prostanoid (EP) receptors EP1 through EP4 which were cloned in the early 1990s (20-23). EP1-EP4 are G-protein coupled Family A receptors with seven transmembrane domains. The EP1 and EP3 receptors are unique among PGE₂ receptors in that splice isoforms exist (24-27). The splice isoforms for the EP3 receptor alter the C-terminal tail and each isoform has been shown to have similar binding properties but activate different signal transduction pathways. PGE₂ binds with highest affinity to the EP3 and EP4 receptors ($K_D < 1$ nM), and with lower affinity to the EP1 and EP2 receptors ($K_D > 10$ nM)(28). Even though each EP receptor binds the same endogenous ligand, the amino acid identity conserved across receptors is only 28-33 % (29). Thus, selective compounds for the EP subtypes have been highly pursued.

EP1 receptors

The EP1 receptor was cloned in 1993 from a mouse lung cDNA library and later from human erythroleukemia cells (21,30). EP1 was first characterized based on its ability to induce smooth muscle constriction. Signaling by the EP1 receptor is primarily transduced by increasing intracellular calcium, most likely mediated by G_q family G-proteins (31). However, it was reported that the calcium

flux weakly induces phosphoinositide turnover and generation of IP₃ and the calcium flux is partially dependent on extracellular calcium, suggesting a non-G_q dependent pathway may also exist (24). Additionally, the EP1 receptor has been demonstrated to couple to G_{i/o}-type G proteins, inducing the up-regulation of HIF-1 α (32).

An EP1 splice variant has been identified in rats (27). The rEP1-variant mRNA was found to be highly expressed in the rat kidney, in a proportion roughly equal to that of rEP1. This variant lacks the intracellular C-terminal tail of the receptor. It is able to bind ligand similar to rEP1, but unable to transduce signals. When co-expressed in CHO-cells with rEP1 or rEP4, rEP1-variant significantly suppresses signaling through the rEP1 and rEP4 receptors, suggesting it may be physiologically relevant for attenuating the actions of PGE₂ (27). However, no splice variants have been identified in other species to date.

Northern blot analysis of mouse EP1 mRNA demonstrated that the kidney has the highest expression of EP1, followed by lung, spleen and skeletal muscle, and low expression in the testes (21,30). More recently, utilizing quantitative PCR analysis Regard et al. showed ubiquitous expression of EP1, present in all 41 tissues tested, suggesting that EP1 is likely expressed in the vasculature (33). Unfortunately, due to the lack of well-characterized antibodies for EP receptors and the lower affinity of EP1 for PGE₂, the only radiolabeled ligand available, protein distribution remains unknown.

Selective agonists and antagonists have been essential for characterization of the individual EP receptor subtypes (Table 1.2). 17-phenyl- ω -

trilor-PGE₂ is the frequently utilized agonist with high affinity for mouse EP1 ($pK_i = 7.9$) (34,35). Sulprostone and partial agonist iloprost have the next highest affinities for mouse EP1 ($pK_i \approx 7.7$), but are less selective (34,36). ONO-DI-004 is by far the most selective agonist for EP1, but has lower affinity than other agonists ($pK_i = 6.8$)(37). AH6809 was frequently used in the past as an EP1 antagonist, although its affinity for the mouse EP1 receptor is very poor, and much lower than its affinity for EP2. ONO-8713, ONO-8711 and SC-51322 are currently used antagonists for EP1. ONO-8713 is the most selective antagonist with a pK_i of 9.5 at the mouse EP1 receptor (38). ONO-8711 also has high affinity ($pK_i = 8.8$) for mouse EP1, although it blocks EP3 receptor with reasonable affinity ($pK_i = 7.2$) (39). SC-51322 is by far the most commonly used antagonist due to its commercial availability, however, its pharmacological properties have only been reported in humans ($pK_i = 7.9$) and not in mice (28). In humans, SC51322 is also reported to block the EP3 receptor ($pK_i = 6.2$).

EP2 receptors

EP receptors were initially classified based on their function on smooth muscle (40). The EP2 and EP4 receptors induce smooth muscle relaxation. Initially, there were two cloned receptor subtypes called EP2 (22,23). Based on pharmacological selectivity, it was later shown that the first cloned EP2 receptor was actually EP4; the latter representing EP2.

Table 1.2 Pharmacological agents targeting mouse PGE₂ receptors

Receptor	Signaling	Selective Drugs
EP1	G _q , G _{i/o} , ?	Agonists: 17-phenyl- ω -trilor-PGE ₂ > sulprostone \approx iloprost > M&B 28767 \approx ONO-DI-004 Antagonists: ONO-8713 > ONO-8711 >> SC-51322 > AH6809
EP2	G _s , Arrestin	Agonists: ONO-AE1-259 >> butaprost \approx AH13205 \approx 11-deoxy-PGE ₁ Antagonists: PF-04418948 >>> AH6809
EP3	G _{i/o} , G _q , G _s , G ₁₂	Agonists: M&B 28767 \approx sulprostone \approx SC-46275 > GR63799 \approx ONO-AE-248 Antagonists: ONO-AE3-240 \approx L-798,106 \approx DG-041 >> ONO-8711 > SC-51322
EP4	G _s , G _i , Arrestin	Agonists: ONO-4819 > ONO-AE-329 \approx 11-deoxy-PGE ₁ > 1-OH-PGE ₁ Antagonists: ONO-AE3-208 \approx MF-498 >> GW 627368 >> AH23848

Both EP2 and EP4 receptors couple to G_s -type G proteins which activate adenylate cyclase and increase intracellular cAMP ($[cAMP]_i$) (22,23,41,42), however EP2 appears to increase $[cAMP]_i$ with greater efficacy than EP4 (43). Additionally, EP2 has also been demonstrated to couple to arrestin-mediated signaling pathways (44-46). The EP2-beta-arrestin1-p-Src complex plays a significant role in the development and progression of skin cancer by activating EGFR (46). Although EP2 has the least abundant mRNA expression of the EP receptors, EP2 mRNA is expressed in the lung, bone marrow and ovary (33,47), however its expression is known to be altered in response to stimuli. Therefore, mRNA quantitation in healthy mice may not portray an accurate picture of EP2 expression in a disease setting.

The EP2 receptor is classified due to its ability to be activated by the agonist butaprost. In the mouse, butaprost, AH13205 and 11-deoxy-PGE₁ all have similar affinity for the receptor ($pK_i = 6.6-7.3$), although 11-deoxy-PGE₁ also binds to the EP3 and EP4 receptors (34). ONO-AE1-259 has improved potency over the previously named agonists ($pK_i = 8.5$) and is highly selective for the EP2 receptor in mice (37). However, there is no reported data regarding its selectivity or affinity against human receptors. Until recently, a quality EP2 antagonist did not exist. AH6809 was shown to have moderate affinity for the EP2 receptor (mouse $pK_i = 6.5$), however it also blocked the EP1 and DP receptors with high affinity in human tissue (28,34,48). Recently, Pfizer reported development of a highly selective, potent EP2 receptor antagonist, PF-04418948 (49-51), which

should prove very useful for the study and validation of EP2 as a therapeutic target.

EP3 receptors

The EP3 receptor is known as the “inhibitory” PGE₂ receptor and induces smooth muscle contraction. EP3 primarily couples to a G_i-type family G proteins and inhibits adenylate cyclase and decreases [cAMP]_i (20,41,52). The EP3 receptor has been demonstrated to increase intracellular calcium (52,53), and also to couple to G₁₂-type G proteins, inducing the activation of RhoA (54). EP3 receptors are unique among the EP receptors in that several splice variants have been identified: at least 7 variants in humans, 5 in rabbits, 4 in rats, and 3 in mice (55-57). The splice variants of EP3 have been demonstrated to alter constitutive activity, agonist-induced desensitization, trafficking, and signal transduction (26,58-60). For example, of the three splice variants found in mice, alpha and beta isoforms couple to G_i G-protein, while the gamma isoform can couple to both G_i and G_s G-proteins (26).

EP3 receptor expression is widespread, and it is found in the kidney, uterus, pancreas, stomach, thymus, spleen, smooth muscle of the gastrointestinal tract, vasculature, and CNS (33,61). In the brain, EP3 receptor mRNA has been observed in the hippocampus, preoptic area, hypothalamus, locus coeruleus and raphe nuclei (62).

Several selective agonists and antagonists exist for the EP3 receptor. The most commonly used agonist for EP3 is sulprostone, which has a pK_i at mouse EP3 of 9.2 (34). However, sulprostone is not entirely selective for EP3,

and has a pK_i at mouse EP1 of 7.7. Therefore interpretation of studies utilizing high concentrations must be carefully considered. Sulprostone has a similar affinity for EP3 as M&B 28767 and SC-46275. GR-63799 and ONO-AE-248 are more selective for mouse EP3 and have pK_i 's of 8.7 and 8.1 respectively (34,37,63). Multiple antagonists exist for the EP3 receptor as well, although not all have been characterized with mouse receptors. ONO-AE3-240, L-798,106 and DG041 have the highest affinities for the EP3 receptor and are selective for the human EP3 receptor (64-67). ONO-8711 is also a selective antagonist with a pK_i of 7.6. Some of the effects of SC-51322, a commonly used antagonist for EP1, may be explained by the blockade of EP3 ($pK_i = 6.2$), especially when high concentrations are used (28).

EP4 receptors

The EP4 receptor is an inducer of smooth muscle relaxation. Initially, it was identified as the EP2 receptor but later determined to be the fourth subtype of EP receptors (22). EP4 mRNA is predominantly localized the uterus, thymus, ileum, lung, spleen, adrenal gland, and kidney (33). EP4 signals through G_s -type G protein, which activates adenylate cyclase and increases $[cAMP]_i$. The EP4 receptor can also activate PI 3-kinase signaling pathways, leading to activation of extracellular signal-regulated kinases 1 and 2 (68). The activation of PI-3K was demonstrated to be pertussis toxin (PTX) sensitive, and therefore mediated by G_i G-protein (69). PTX reduced PGE_2 stimulated $[cAMP]_i$ in cells expressing EP4 receptors, suggesting EP4- G_i signaling both inhibits adenylate cyclase and

activates PI-3K. EP4 was also demonstrated to couple to arrestin-mediated signaling pathways (45,46).

EP4 was distinguished from EP2 due to its lack of activation in response to butaprost. The EP4 receptor is preferentially activated by ONO-4819 ($pK_i = 9.2$) and ONO-AE-329 ($pK_i = 8.0$), whereas these compounds' affinities for the EP2 receptor are 100-1000 times lower (37,63,70). In humans, 11-deoxy-PGE₁ is selective for EP4, however in mouse and rat 11-deoxy-PGE₁ is able to bind EP2 and EP3 receptors in addition to EP4 (34,71,72). In mice, 1-OH-PGE₁ is a selective agonist but has lower affinity for EP4 ($pK_i = 6.7$) (34). In addition to agonists there are several antagonists available for blockade of the EP4 receptor. AH23848 was the most commonly used antagonist, however it has low affinity for EP4 ($pK_i = 5.3$), and although selective against mouse receptors, has higher affinity for the human TXA₂ receptor than human EP4 (28,71,73). Among the newer antagonists, ONO-AE3-208 has the highest affinity for the mouse EP4 receptor, although it also blocks the EP3 receptor at slightly higher concentrations despite being selective for the human EP4 receptor (74). Fortunately Merck compound MF-498 has both high affinity and selectivity for the EP4 receptor (75,76). Additionally, GW627368 is selective and has reasonable affinity for EP4 ($pK_i = 7.1$) (77). These EP4 antagonists have proven to be particularly useful for the study of EP4 in disease given 95 % of EP4^{-/-} mice die as a result of patent ductus arteriosus shortly after birth (78).

PGE₂ receptors as therapeutic targets

PGE₂ mediates a wide range of physiological conditions including modulation of salt and water transport in the kidney, maintenance of blood pressure, preservation of the gastric mucosa, tumor growth and angiogenesis, and modulation of immune responses. The COX/PGE₂ pathway has long been a target of therapeutic benefit. Some of the most commonly used over-the-counter medications - NSAIDs - block this pathway by inhibiting the COX enzymes. These drugs are very valuable for the treatment of pain, inflammation, and fever; however chronic use can cause serious side effects including ulcers and gastrointestinal bleeding, hypertension, heart attack or stroke. In an effort to eliminate the side effects, subtype selective inhibition of COX-2, while maintaining the function of the “housekeeping” isoform COX-1, was pursued. The drugs produced include rofecoxib (Vioxx) and celecoxib (Celebrex). Unfortunately, rofecoxib was withdrawn from the market due to an increased risk of myocardial infarction and stroke, despite showing efficacy against cancer progression and reduced incidence of gastrointestinal bleeding. These unexpected side effects were a result of an imbalance in thromboxane and prostacyclin action. Celecoxib has remained on the market and is currently prescribed for rheumatoid and osteoarthritis, acute pain and familial adenomatous polyposis.

Attention has since focused on alternate ways to modulate the COX/PGE₂ pathway. These include targeting the catabolic enzyme for PGE₂, prostaglandin dehydrogenase, targeting the synthases responsible for production of the PGE₂,

targeting the transporter which moves the intracellular PGE₂ across the plasma membrane, or targeting the PGE₂ receptors themselves. Therefore, selective agents and knockout mice have been used to investigate the subtype specific roles of the EP receptors in health and disease.

EP1 receptors

Although EP1^{-/-} mice have no gross dysfunctions, they have been used to support the role of EP1 in impulse activity, pain, hypertension, urine concentration and cancer. Most notably, the EP1 receptor is actively being pursued as a therapeutic for pain. In two acute pain models, EP1^{-/-} mice have been shown to have reduced nociceptive responses (79). The reduction in pain phenocopied the effect observed by treatment with a COX-2 inhibitor, suggesting the EP1 receptor is the major mediator of COX/PGE₂ pain (79). PGE₂-dependent mechanical allodynia, or enhanced sensitivity to a stimulus which normally does not cause pain, is prevented in EP1^{-/-} mice (80). Furthermore, anti-nociceptive and anti-inflammatory responses have been observed as a result of administration of an EP1 antagonist in several studies (67,81-84).

EP1^{-/-} mice exhibit modestly reduced systolic blood pressure (79). This effect was also observed in a setting of hypertension. EP1^{-/-} mice have a blunted response to acute and chronic angiotensin II-dependent hypertension (85). Additionally, EP1 antagonists significantly reduced hypertension in diabetic mice and spontaneously hypertensive rats (85,86). EP1 antagonists have also

been used for the prevention of hypertensive renal damage and diabetic nephropathy (87,88).

In the lung, EP1 receptors constrict the airway of mice and mediate surfactant secretion from rat alveolar cells (89,90). In the stomach, EP1 plays both beneficial and harmful roles. EP1 enhances histamine-induced gastric injury while also conferring cytoprotection against hemorrhagic lesions and is essential for HCO₃⁻ secretion (91-94). EP1 receptors have also been shown to play a critical role in carcinogenesis. EP1^{-/-} mice have reduced aberrant crypt foci and tumor formation in the azoxymethane colon cancer model (39,95). EP1 antagonist ONO-8711 was demonstrated to reduced tumor burden in a model of breast cancer (96). Therefore, EP1 antagonists may be of therapeutic importance for pain, hypertension, and cancer.

There are few indications when EP1 agonism would be beneficial. EP1^{-/-} mice exhibit behavioral deficits including impaired social interaction, aggression, diminished cliff avoidance and exaggerated acoustic startle response (97). Therefore, agonism of EP1 receptors could prove to be useful for control of impulsive behavior, although major unwanted side effects would be expected.

EP2 receptors

EP2 has been demonstrated to be involved in asthma, blood pressure homeostasis, uterine contraction, fertility, maintenance of bone, and cancer. EP2 is generally considered to have anti-inflammatory roles (98). EP2^{-/-} mice have reduced fertility (17,78,99). Although female EP2^{-/-} mice are able to become

pregnant, there is a reduction in litter size. This is a result of reduced fertilization of the oocyte due to incomplete expansion of the cumulus cells required for oocyte maturation (78,99,100). The EP2 receptor mediates the vasodilator responses of PGE₂ (101). During high salt intake, deletion of EP2 results in salt-sensitive hypertension (17,99). One mechanism by which EP2 can modulate blood pressure is through PGE₂-induced renin release. EP2 and EP4 activate renin exocytosis at the juxtaglomerular cell, eventually resulting in vasoconstriction of the afferent arteriole (102-104). EP2 also mediates sodium excretion, suggesting that in a high salt setting EP2 stimulates natriuresis and maintenance of normal blood pressure (105).

The EP2 receptor mediates the PGE₂ bronchodilator response, suggesting agonism of EP2 may be beneficial for the treatment of asthma. This has been demonstrated in both human and mouse (89,106-108). Agonism of EP2 receptors could prove to be more beneficial than commonly used treatments which relieve bronchospasm, because EP2 receptors also inhibit lung mast cell degranulation (109). AH13205 was tested clinically with no success, however it should be noted that the potency to this EP2 agonist is weak, and there may be great benefit from a more potent compound if it becomes available (110).

The EP2 receptor can positively and negatively contribute to cancer. In skin cancer, overexpression of EP2 enhances chemically induced skin cancer and EP2^{-/-} mice are resistant to the cancer (111,112). In intestinal and prostate cancers, EP2 receptors contribute to endothelial cell motility and mediate angiogenesis (113). In mammary cancer, EP2 deficiency reduces tumor growth,

angiogenesis, and metastasis, while over expression of EP2 enhances tumor burden (114). This supports a pro-tumor role of EP2. In contrast, loss of EP2 receptors in keratinocytes has been suggested as a mechanism for neoplastic progression by increasing invasiveness. Furthermore, EP2 receptors are protective in UV-induced carcinogenesis (115,116), supporting an anti-tumor role of EP2. It is essential to appreciate the complexity of the EP2 receptor in cancer progression. Therapeutic manipulation of EP2 for the treatment of cancer would require determining whether EP2 is acting in a pro- or anti-tumorigenic fashion prior to treatment.

EP3 receptors

EP3 receptors have been implicated in many physiologic events including fever, pain, platelet aggregation, myocardial infarction, bladder hyperactivity, cancer, obesity, and Alzheimer's disease. Upon gross observation, EP3^{-/-} mice appear to be normal (117,118). However, EP3^{-/-} mice have been shown to have an impaired febrile response (118), suggesting EP3 antagonists could be effective antipyretic agents. Analgesic effects are also produced by EP3 antagonists, or the use of EP3^{-/-} mice, in a model of acute herpetic pain (119). EP3 antagonism has been clinically pursued for treatment of thrombotic disorders. PGE₂ inhibits platelet aggregation at high dose, but also can potentiate other proaggregatory agents (120). EP3 receptors have been demonstrated to mediate this proaggregatory effect (64,121-124). Furthermore,

EP3^{-/-} mice have increased bleeding and reduced susceptibility to thromboembolism (123).

With the use of EP3 antagonists, the receptor has been identified as a target for treatment of bladder hyperactivity disorders. Treatment with DG041 inhibited responses to bladder distension and reduced bladder motility (125). Furthermore, EP3^{-/-} mice have been reported to have increased bladder capacity, while infusion of an EP3 agonist reduces bladder capacity in wildtype mice (126).

EP3 has also been implicated in metabolic regulation. EP3^{-/-} mice had increased frequency of feeding during the light cycle and became obese on a normal fat diet (127). Increased leptin and insulin levels were also observed with the obesity (127). In a model of subacute neuroinflammation, deletion of EP3 reduced pro-inflammatory gene expression, cytokine production, oxidative stress, and reversed the decline in presynaptic proteins, suggesting a possible role for EP3 in Alzheimer's disease (128,129).

Similar to the EP2 receptor, in cancer EP3 can have both pro- and anti-tumorigenic effects. In human colon cancer EP3 mRNA expression is reduced. Agonism of EP3 in a colon cancer cell line reduced cell viability, supporting the theory that downregulation of EP3 allows for enhanced colon carcinogenesis (130). Overexpression of individual splice variants decreased tumorigenic potential in cell lines and reduced tumor burden in vivo (54). EP3^{-/-} mice also exhibited reduced tumor burden and angiogenesis in an implanted tumor model (65). Similar reductions in tumor burden were observed upon treatment with an

EP3 antagonist (65). In contrast, EP3 decreases aromatase activity, suggesting a possible inhibitory role in breast cancer (131).

EP4 receptors

EP4 receptors have been proposed to mediate vasodepression, closure of the ductus arteriosus, renin secretion, bone homeostasis and intestinal homeostasis. EP4^{-/-} mice have been demonstrated to have persistent patent ductus arteriosus, and most mice die within 3 days of birth due to pulmonary congestion and congestive heart failure (132,133). The ductus arteriosus is the fetal shunt between the pulmonary artery and aorta, and its closure at birth is essential. Indomethacin can be given late in pregnancy to induce closure of the ductus in wildtype, but not EP4^{-/-} mice (132).

EP2 and EP4 receptors have also been shown to mediate PGE₂ stimulated renin secretion. PGE₂ is a well established stimulator of renin (19,134). It is also known that PGE₂ is produced locally at the macula densa in response to relevant concentrations of NaCl (20-40 mM) (135). Using isolated, perfused kidneys for EP1, EP2, EP3 and EP4 knockout mice, it was demonstrated that EP4 stimulates renin secretion at low PGE₂ concentrations, regulating basal activity, while both EP4 and EP2 stimulate renin at higher concentrations of PGE₂ (102). However, EP receptors do not appear to mediate COX-2 dependent production of renin at the macula densa in low-NaCl settings, since no differences in renin secretion were observed between genotypes when

loop diuretics, which impair the ability of the macula densa to sense NaCl, were administered (102).

EP4 is the primary PGE₂ receptor supporting bone maintenance, and is able to mediate both bone formation and bone reabsorption. EP4^{-/-} mice have been shown to have reduced bone mass and an inability to properly repair fracture damage (136). Despite having similarly sized bones, EP4^{-/-} mice had reduced structural and apparent material strength in the femur and vertebrae compared to wildtype mice (137). Administration of an EP4 agonist has been shown to restore bone mass and strength in ovariectomized or immobilized rats, suggesting activation of EP4 could be of great therapeutic benefit for conditions with reduced bone mass (70).

It has been established that treatment with NSAIDs can trigger or worsen inflammatory bowel diseases. This phenomenon is due to reduced activation of EP4. In a dextran sodium sulfate–induced colitis, disruption of EP4 and not EP1, EP2, EP3, FP, TP or IP resulted in enhanced colitis in wildtype mice (74). Administration of an EP4 antagonist to wildtype mice was able to mimic this effect, while treatment with an EP4 agonist ameliorated severe colitis (74). The anti-inflammatory EP4 receptor was critical to mucosal barrier function and maintenance of epithelial cells, and suggests EP4 agonism may be of therapeutic value for treatment of inflammatory bowel diseases such as Crohn disease and ulcerative colitis.

PGE₂ receptors in blood pressure regulation

PGE₂ is a major prostanoid contributing to the regulation of blood pressure, where it can exert either vasopressor or vasodepressor effects depending upon the setting (17-19). Previous studies have determined that the EP1 and EP3 receptors primarily mediate the pressor response, while the EP2 and EP4 receptors mediate the depressor response (17,85,99,134,138-140)(Figure 1.2).

The role of PGs in the regulation of blood pressure is highlighted by the prohypertensive action of non-steroidal anti-inflammatory drugs (NSAIDs) which inhibit cyclooxygenase mediated prostanoid production, suggesting that prostaglandins have an anti-hypertensive role (141,142). However, under certain conditions NSAIDs can also have hypotensive effects supporting a prohypertensive role for PGs (143). Of note, PGE₂ is a well established stimulator of renin (19,102,135,144). Experiments in conscious dogs demonstrated that intrarenal infusion of PGE₂ results in a sustained rise in mean arterial pressure, which highly correlates with plasma renin activity (19). This suggests the rise in blood pressure was related to the increased renin activity. Furthermore, 20 – 40 mM NaCl concentrations have to shown to induce local PGE₂ production in the macula densa (135). Overall, the balance of functionally antagonistic prostaglandin action fine-tunes blood pressure homeostasis.

Isomerization of PGH₂ to PGE₂ by microsomal PGE synthase (mPGES) plays a key role in the maintenance of blood pressure homeostasis by regulating

vascular tone, sodium balance and/or renin release (145). Inhibitors of mPGES are currently being pursued for treatment of cancer, pain and inflammation; however, unwanted pro-hypertensive effects may result from this strategy. In rodents, deletion of mPGES-1 has been shown to further increase blood pressure in several models of hypertension, including deoxycorticosterone-salt water-induced hypertension and acute or chronic treatment with angiotensin II (146-148). In humans, where there is a great deal of phenotypic heterogeneity, the contribution of PGE₂ towards hypertension is more controversial. In some cases, patients with essential hypertension have been shown to have low urinary excretion of PGE₂; however, in other cases patients have been shown to have high urinary PGE₂ excretion (149). This inter-patient variability may result in a wide range of untoward side effects for mPGES inhibitors. In order to gain a better understanding of the effects of PGE₂ on blood pressure homeostasis, attention has been focused on the actions of its receptors using knockout mouse models.

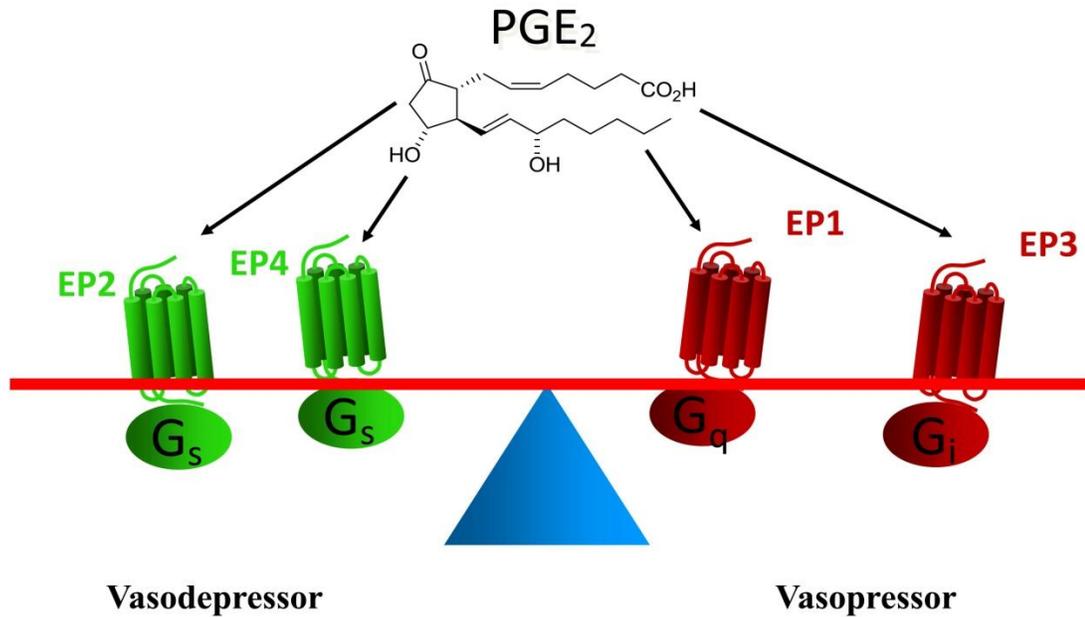


Figure 1.2 PGE₂ receptors in blood pressure regulation. Upon PGE₂-induced activation EP2 and EP4 function as vasodepressor receptors and EP3 and EP1 function as vasopressor receptors

Vasodepressor Receptors

Upon acute infusion, PGE₂ is a vasodepressor in both humans and mice (17,138,150). This observation underscores the pro-hypertensive effects of blockade of all prostaglandin production and subsequent receptor activation by NSAIDs (141,151-154), which is consistent with the loss of a tonic vasodepressor PG effect. It has been shown that this depressor effect is primarily due to the activation of the EP2 receptor; however, when the EP2 receptor is deleted, the loss of the depressor response unmasks a PGE₂ pressor response (17). Moreover, EP2^{-/-} mice fed a high-salt diet experienced an increase in blood pressure consistent with a protective role for EP2 activation in salt-sensitive hypertension (17). Intravenous infusion of the EP2 agonist ONO-AE1-259 into

Wistar rats increases retinal arteriolar and venous diameter and substantially reduces mean arterial pressure (155). EP2 and EP4 receptors evoke an increase in $[cAMP]_i$ through a G_s coupled pathway, a classical mechanism for smooth muscle relaxation. Hristovska et al. demonstrated a dose-dependent relaxation in response to PGE_2 in aortic rings which was lost in tissue from EP4 $^{-/-}$ mice but remained intact in EP2 $^{-/-}$ tissue. The EP4 dilator effect was dependent upon endothelium-derived nitric oxide production via eNOS (156). Acute blood pressure studies are challenging in EP4 $^{-/-}$ mice because the mice exhibit near complete perinatal lethality in inbred strains as a result of persistent patent ductus arteriosus (132). Analysis of studies performed with surviving EP4 $^{-/-}$ animals on a mixed-strain background may not be straightforward as their survival may be dependent on modifier genes. Nonetheless, deletion of EP4 resulted in a diminished vasodepressor response to PGE_2 (138). In rats, infusion of the EP4 selective agonist ONO-AE1-329 significantly reduces blood pressure; it does not alter retinal vessel diameter (155). Taken together is consistent with EP4 vasodilator action in a subset of vascular beds.

Vasopressor Receptors

As described above, the pressor effects of systemic PGE_2 infusion are only observed in the absence of the predominant depressor receptor EP2 (17). In contrast, infusion of EP3 receptor selective agonists such as sulprostone, M&B28767 or SC46275 in wildtype mice results in an acute and substantial rise in mean arterial pressure (139). The EP3 mediated pressor effect undergoes

desensitization with repeated administration of EP3 agonists. In EP2^{-/-} mice after desensitization of EP3 responses, the depressor action of EP4 in response to PGE₂ infusion is then apparent (139). Thus, upon systemic infusion of PGE₂ in mice the depressor action of EP2 predominates, followed by the pressor action of EP3, and then the depressor action of EP4. Importantly, the order of expression of EP receptor mRNA does not mirror the phenotypic effects of the EP receptors. RNA levels determined by RNase protection identified expression levels of EP3>>EP4>EP1≥EP2 in both renal resistance vessels and the aorta (139). It is unclear whether changes in EP receptor density underlie changes in vascular tone in the hypertensive state. Because messenger RNA levels do not correlate with receptor function, and anti-receptor antibodies are of questionable value, this remains an important unanswered question.

In contrast to the depressor effects of systemic administration, when PGE₂ is administered intracerebroventricularly (ICV) a rise in mean arterial pressure occurs, accompanied with tachycardia and enhanced renal sympathetic nerve activity (157). These effects were ascribed to the EP3 receptor using ICV infusion of subtype selective EP receptor agonists (157). Taken together, these data demonstrate that the centrally mediated pressor actions of PGE₂ are EP3-mediated.

Although the EP1 receptor does not appear to play a significant role in the blood pressure effects of systemically administered PGE₂, it has been shown to be a significant contributor to hypertension, particularly in cases with enhanced renin-angiotensin system activity. Genetic deletion of the EP1 receptor in mice

has been shown to significantly decrease systolic blood pressure, an effect amplified when mice are fed a low sodium diet (79). Importantly, EP1^{-/-} mice have blunted pressor responses to both acute and chronic angiotensin II administration (85). In isolated vascular preparations of preglomerular arterioles and mesenteric arteries, pre-treatment with SC51322, an EP1/3 antagonist, was able to abolish any angiotensin II-mediated vasoconstriction (85). Furthermore, treatment of spontaneously hypertensive rats, a multifactorial model of essential hypertension, with SC51322 significantly reduces blood pressure (85), indicating the EP1 receptor and/or EP3 receptor may be novel targets for the treatment of hypertension.

PGE₂ and the consequences of hypertension

Hypertension is an established risk factor for cardiovascular diseases including stroke, myocardial infarction, heart failure, arterial aneurysm and is the leading cause of chronic kidney failure. Current anti-hypertensive therapies reduce the risk of the related cardiovascular sequelae, though not to baseline risk observed in normotensive subjects. There is an unmet need for treatment which will reduce blood pressure and maximize target organ protection (158). In considering whether PGE₂ and its receptors make viable drug targets for hypertension, determination of their ability to reduce end-organ damage will be important. It has been shown that genetic deletion of mPGES-1 in mice results in increased blood pressure (146-148). Despite this, deletion of mPGES-1 has also

been shown to protect against aortic aneurysm formation and vascular injury (159,160). Angiotensin II infusion into hyperlipidemic mice produced fewer and less severe aneurysms, and reduced oxidative stress on a mPGES-1-/- background compared to wildtype mice (160). However, these results were complicated by the observed increase in PGI₂ and PGD₂ production accompanying the reduction in PGE₂ (160). It is yet to be determined whether potentially beneficial substrate diversion is a consequence specific to genetic mPGES-1 deletion, or would be recapitulated with chronic use of an mPGES inhibitor.

Blockade of individual PGE₂ receptors might result in a reduction in end-organ damage while being less likely to produce unwanted side effects. In addition, GPCRs are demonstrably “druggable” and are one of the most common targets of currently developed therapeutic agents. Antagonism of EP1 receptors has been shown to preserve renal function, and reduce tubulointerstitial damage, proliferative lesions, fibrotic area and proteinuria in stroke-prone spontaneously hypertensive rats (87), as well as cerebrovascular dysfunction induced by angiotensin II (161). In the study performed in stroke-prone hypertensive rats, tail cuff blood pressure was modestly reduced two weeks post-treatment with an EP1 antagonist, but this reduction was not maintained past five weeks of treatment. Nonetheless treatment with the EP1 antagonist provided end-organ protection. In contrast to the deleterious actions of the EP1 receptor, EP2 and EP4 receptors have been shown to be cardioprotective; it would seem important to maintain function of these receptors. For example, deletion of the EP4

receptor in a mouse model of ischemia reperfusion of the heart significantly increased infarct size, while treatment of wildtype mice with an EP4 agonist, 4819-CD, reduced infarct size (162). Therefore, EP4 agonists could be useful for reducing blood pressure and afford cardioprotective benefits. Selective blockade of EP1 and/or EP3 receptors, while EP2 and EP4 signaling remains intact may be preferable to the loss of signaling in all four receptors resulting from inhibition of PGE₂ ligand production.

PGE₂ plays a dynamic role in regulation of blood pressure homeostasis. The existence of multiple receptors with diverse signaling abilities allows for modulation both positively and negatively. The development and availability of additional highly selective agonists and antagonists for EP receptors is fundamental to the advancement of the field. Unwanted side effects resulting from inhibition of the cyclooxygenase enzymes upstream of prostanoid production demonstrated the value of selective targeting as proximal to the pathophysiological action as possible. Development of new therapeutics targeting specific PGE₂ receptors could reduce blood pressure and provide end-organ protection, while minimizing side effects.

Objective

As summarized above, PGE₂ and its receptors mediate a wide range of physiologic processes. With regards to blood pressure regulation, PGE₂ can act in a pro-hypertensive or anti-hypertensive manner. While it has been demonstrated that EP2 and EP4 receptors mediate the vasodepressor actions of PGE₂ and the EP1 and EP3 receptors mediate the vasopressor actions, the role of the EP receptors in modulating downstream end-organ damage is incompletely characterized. Current therapies for cardiovascular and renal complications associated with hypertension and diabetes are largely focused on reduction of mean arterial pressure. However, pharmacologic reduction of blood pressure in patients with hypertension – even to the same blood pressure as normotensive individuals – does not reduce the risk of CVD and renal events to an incidence similar to that of normotensive individuals. This highlights the need for development of novel therapeutic agents which will not only lower blood pressure but also reduce end-organ damage.

Drugs which block the renin-angiotensin-aldosterone pathway are considered superior to other anti-hypertensive treatments due to their beneficial actions directly on the kidney. Furthermore, PGE₂ and the EP1 receptor have been demonstrated to mediate at least part of the actions of angiotensin II. Therefore, I sought to determine the role of the vasopressor PGE₂ receptors in the development of hypertensive end-organ damage. Specifically, I sought to address the following questions: What are the consequences of genetic

disruption of EP1 or EP3 receptors in development of Ang II- mediated hypertension? Do EP1 or EP3 receptors contribute to end-organ damage in a setting of hypertension or diabetes? Are beneficial effects on end-organ damage dependent of blood pressure reduction? Does the EP1 receptor contribute to Ang II independent hypertension? This thesis describes studies utilizing EP1 and EP3 receptor knockout mice which are designed to answer these questions and advance our knowledge of the role of EP receptors in settings of disease.

CHAPTER II

DISRUPTION OF EP1 ATTENUATES END-ORGAN DAMAGE IN A MOUSE MODEL OF HYPERTENSION

Introduction

Hypertension is a major risk factor for cardiovascular diseases (CVD), increasing the risk of stroke, myocardial infarction, arterial aneurysms, and heart failure. Approximately 30 % of adults in the US have hypertension and the incidence of CVD remains greater in hypertensive patients than normotensive patients, highlighting the need for novel therapeutic agents (158).

PGE₂ is a major prostanoid found in the mouse kidney and vasculature contributing to the regulation of blood pressure, where it can exert either vasopressor or vasodepressor effects (17-19). Four PGE₂ receptors (EP1 through EP4) mediate these effects with the EP1 and EP3 receptors primarily mediating the pressor response of PGE₂, while the EP2 and EP4 receptors mediate the depressor response (17,85,99,134,138-140).

Each PGE₂ receptor has distinct tissue localization and elicits characteristic signal transduction pathways (61). EP1 couples to G_q-proteins, resulting in mobilization of intracellular calcium, and stimulation of phosphoinositide turnover activating protein kinase C. The EP2 and EP4 receptors couple to G_s-proteins, which increase intracellular cAMP. The EP3

receptor couples to G_i-proteins, decreasing intracellular cAMP (For reviews, see (29,61)). Receptors couple to alternative signal transduction pathways as well, including arrestin-mediated signaling pathways (45,46).

Systemic infusion of PGE₂ results in a vasodepressor response (17,138,150), primarily through EP2 activation. In the absence of EP2, a PGE₂ pressor response is unmasked (17), mediated by the EP3 receptor (139). Agonist induced EP3 tachyphylaxis in the background of EP2 -/- mice uncovers a depressor action of EP4 (139). EP1 does not appear to play a significant role in the blood pressure effects of systemically administered PGE₂, however, it does contribute to hypertension. Genetic disruption of the EP1 receptor in mice has been shown to decrease blood pressure, particularly when mice are fed a low sodium diet (79). Furthermore, EP1-/- mice have blunted pressor responses to both acute and chronic angiotensin II (Ang II) administration (85). In isolated vessel preparations, pre-treatment with the EP1 selective antagonist SC51322 reduced Ang II mediated vasoconstriction (85). Treatment of spontaneously hypertensive rats with SC51322 significantly reduces blood pressure (85), indicating blockade of the EP1 receptor may be a target for the treatment of hypertension.

EP1 blockade has been shown to positively affect renal function in stroke-prone spontaneously hypertensive rats (87), as well as cerebrovascular dysfunction induced by Ang II (161), implicating the EP1 receptor in hypertension and resultant end-organ damage. This has yet to be investigated in detail and in

the context of other organ damage. Therefore, EP1^{+/+} and EP1^{-/-} mice were studied in a model of severe hypertension.

Experimental Procedures

Animal procedures

The hypertension model was carried out as described by Kirchhoff and co-workers (163). Ten-to-16 week old male C57BL/6J (EP1^{+/+}, Jackson Labs, USA) and EP1^{-/-} mice (85) backcrossed at least ten generations onto C57BL/6J were uninephrectomized (Nphx) under ketamine/xylazine (100 mg/kg and 15 mg/kg) anesthesia. Two weeks later, a subcutaneous 50 mg deoxycorticosterone acetate (DOCA) pellet (Innovative Research of America, USA) was implanted and drinking water supplemented with 1 % NaCl. After an additional week, a subcutaneous osmotic mini-pump was implanted (Alzet model 1002; Durect Corporation, USA) delivering 1.5 ng angiotensin II (Calbiochem, USA) per minute per gram body weight. Mice were followed for two more weeks when tissues were collected for histology. Additional studies eliminating one of the three elements from the protocol were performed, while keeping the time frame between surgeries consistent. Animals were maintained in an AAALAC accredited rodent facility in individually ventilated microisolator cages on a 12:12 light dark cycle. All procedures were done in accordance with the policies of the Institutional Animal Care and Use Committee at Vanderbilt University.

Examination of aortic aneurysm and dissection

Upon necropsy aortic aneurysms and dissections were observed in both the thoracic and abdominal aorta, and severity was scored visually using the following scale adapted from Manning et al., 2002 (164). Type 0: no aneurysm, Type 1: dilated aorta with no thrombus, Type 2: remodeled tissue that frequently contains thrombus, Type 3: a pronounced bulbous form of type 2, Type 4: multiple overlapping aneurysms, or a dissection extending the length of the aorta, Type 5: ruptured aorta. Hematoxylin and eosin (H/E) and Masson's Trichrome stains was used on formalin-fixed, paraffin-embedded aorta sections. Immunohistochemistry was performed for macrophage (CD11b and F4/80, Novus Biologicals cat# NB600-1327 and NB600-404), neutrophils (myeloperoxidase, Dako cat# A0398), T-cells (CD3, Santa Cruz cat# sc-1127), and B-cells (CD45R/B220, BD Pharmingen cat# 553084). Inflammation, collagen organization and all immunohistochemistry sections were scored at the site of the lesion in a blinded fashion by a comparative veterinary pathologist.

Fraction water weight determination

Fractional water weight was determined as previously described (165). Eight EP1+/+ and seven EP1-/- mice were sacrificed 5 days post-Ang II administration, their carcasses weighed (wet weight) and incubated at 60°C. Body weight was measured daily until it remained unchanged for 3 days, indicating dry body weight. Fractional water weight was determined using the equation: Fractional water weight = 1 - (dry weight/wet weight).

Determination of urinary albumin/creatinine ratios

Albumin/Creatinine ratios (ACR; expressed as mg albumin/mg creatinine) were measured from 20-200 μ L volumes of spot urine using Albuwell M ELISA kit, and urinary creatinine was measured using the Creatinine Companion (Exocell, Philadelphia, USA).

Determination of Blood Urea Nitrogen levels

To assess the renal function, blood urea nitrogen (BUN) was determined using an iSTAT-1 analyzer (Abbott Point of Care Inc., New Jersey, USA). Whole blood was obtained from saphenous vein and immediately assayed utilizing Chem8+ cartridges.

Assessment of Renal Histopathology

Tissues were fixed overnight in 10 % neutral buffered formalin. Kidneys were processed routinely, embedded in paraffin, sectioned at 5 microns, stained with H/E or Masson's Trichrome and evaluated by light microscopy.

Quantitative PCR

Total RNA from kidneys and aortae was isolated using the TRIzol reagent followed by RNA cleanup with a Qiagen RNeasy kit. cDNA was made using the high-capacity cDNA archive kit (Applied Biosystems). Quantitative PCR was performed using Applied Biosystems 7900HT Fast Real-time PCR system with Taqman gene expression assays for Ngal (Mm01324470_m1), Kim-1

(Mm00506686_m1), Cyclooxygenase-2 (COX-2) (Mm00478374_m1), and 18S rRNA (4319413E). Fold difference in mRNA expression is plotted relative to a normal EP1+/+ kidney sample.

Echocardiography

Transthoracic echocardiography was performed on lightly-anesthetized (isoflurane) mice using the VisualSonics VEVO2100 system (30 MHz transducer). Left ventricular posterior wall dimensions (LVPW), intraventricular septum dimensions (IVS), left ventricular interior diameter (LVID), fractional shortening (FS) and ejection fraction (EF) were measured for analysis of cardiac structure and function at baseline, prior to uninephrectomy, and five days post-Ang II administration.

Intracarotid blood pressure measurement

Intracarotid blood pressure was measured under ketamine (25 mg/kg) and inactin (100 mg/kg) anesthesia delivered intraperitoneally. Mice were placed on a thermal pad and a PE-10 catheter was inserted into the left carotid artery. The catheter was connected to a TXD-310 transducer and blood pressure was measured using a Digi-Med BPA 400 (Micromed). Mice were equilibrated 30-60 minutes until stable values were attained. Ten minute blood pressure measurements were collected and average mean arterial pressure (MAP) is plotted.

Antihypertensive Treatment

Blood pressure reduction was achieved by adding hydralazine (200 mg/L) to the 1 % NaCl drinking water beginning three days prior to angiotensin II mini-pump implantation. Treatment was continued for the duration of the experiment, or a total of 17 days.

Statistical Analysis

Data are means \pm SEM, using GraphPad Prism software (GraphPad Software Inc., USA). Analysis utilized Student's t test and Fisher's exact test. Kaplan Meier survival curves were evaluated with the Log-rank (Mantel-COX) test. $P < 0.05$ was considered statistically significant for all studies.

Results

EP1^{-/-} mice were protected against Nphx/DOCA-NaCl/Ang II mortality

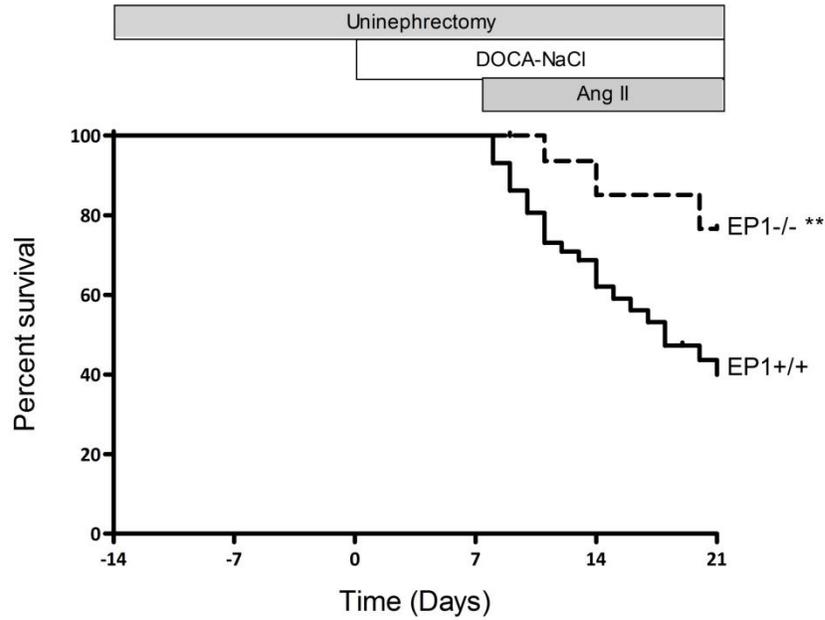
We employed a model of severe hypertension to investigate the contribution of the EP1 receptor in hypertensive end-organ damage (163). Unexpected mortality was observed following implantation of the Ang II osmotic pump (Figure 2.1A). Of 58 EP1^{+/+} mice, 60 % died within 14 days. EP1^{-/-} mice were significantly protected against mortality; of 35 EP1^{-/-} mice only 24 % died ($P = 0.0044$). Modified protocols omitting one of the three components (Nphx, DOCA-NaCl, or Ang II) demonstrated all components of the model played an essential role in causing mortality (Figure 2.1B, Nphx/DOCA-NaCl/Ang II vs. Nphx/DOCA-

NaCl, Nphx/Ang II or DOCA-NaCl/Ang II, N = 10 per group, P = 0.011, < 0.005, or < 0.005 respectively).

EP1^{-/-} mice had reduced aortic aneurysm rupture, but comparable aortic histopathology

Post-mortem analysis of EP1^{+/+} and EP1^{-/-} mice in the full model indicated a significant portion of the mice died as a result of rupture of the aorta. Aneurysms and dissections were observed in both the thoracic and abdominal aorta. 37 % of EP1^{+/+} and 13 % of EP1^{-/-} mice died due to aortic rupture. Aortic aneurysms were present in 67 % of EP1^{+/+} mice and 40 % of EP1^{-/-} mice. Aneurysm severity was scored at death on a scale of 0-5 (Figure 2.2A). A reduction in aneurysm severity was observed in EP1^{-/-} mice compared to EP1^{+/+} mice (Figure 2.2B, P = 0.049). H/E staining revealed aneurysms and dissections in the wall of the thoracic and abdominal aorta were accompanied by inflammation in both EP1^{+/+} and EP1^{-/-} mice (Figure 2.3A-E, P = 0.3975). Analysis of aortic sections from both thoracic and abdominal lesions of each genotype demonstrated macrophage and neutrophils were most abundant, with no differences observed between the genotypes in any immune cell component (Figure 2.3F). COX-2 has been shown to play a significant role in aortic aneurysm formation and macrophage infiltration (166,167). COX-2 mRNA was elevated in abdominal aortae 2-5 days post Ang II administration, though differences between genotypes were not observed (Figure 2.3G, P = 0.774). Aortic sections stained with Masson's Trichrome showed that regardless of genotype there was less fibrillar collagen present in vessels that ruptured, and

A.



B.

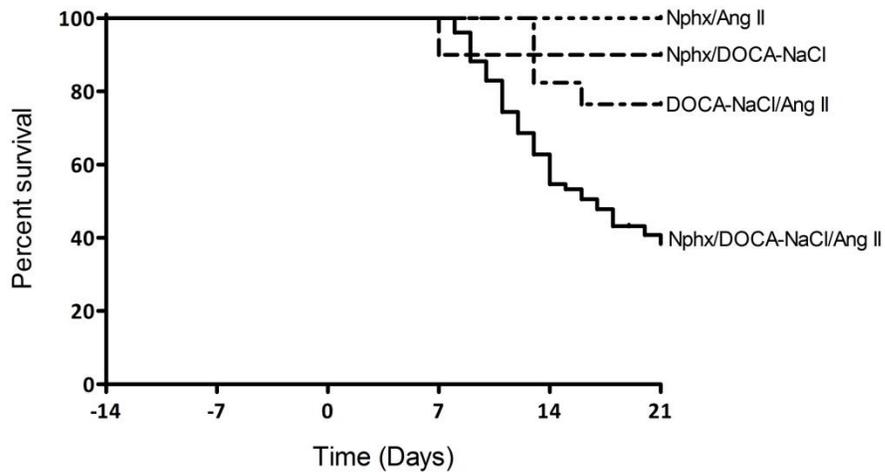


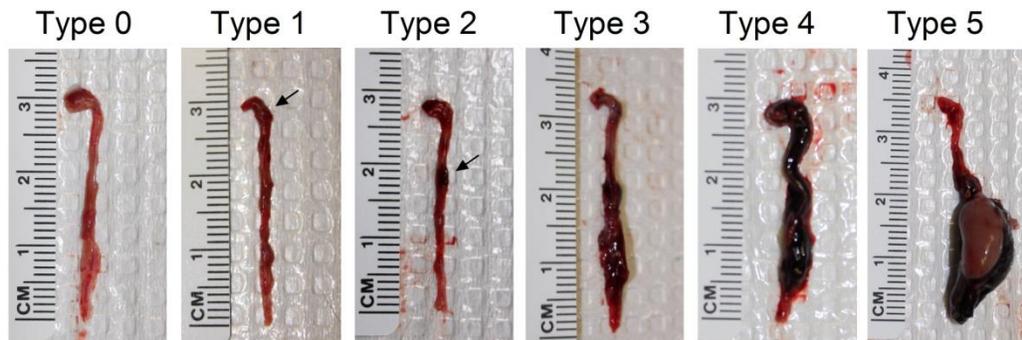
Figure 2.1. Nphx/DOCA-NaCl/Ang II induced substantial mortality in EP1+/+ mice. A. Survival of EP1+/+ and EP1-/- mice. EP1+/+ mice experience a high mortality rate after implantation of the Ang II minipump (60 %, N=58) which is significantly reduced in EP1-/- mice (25 %, N=35). Kaplan Meier survival curves for each genotype are plotted, **P = 0.004. B. Survival of EP1+/+ modified protocol groups. Survival curves were plotted indicating reduced mortality in all modified protocol groups. Nphx/DOCA-NaCl/Ang II (data replotted from Figure 1A) vs. Nphx/DOCA-NaCl, Nphx/Ang II or DOCA-NaCl/Ang II, P = 0.011, <0.005, or < 0.005.

the amount and organization of collagen surrounding the lesion was not significantly different in EP1+/+ and EP1-/- intact aneurysms (Figure 2.4, P = 0.1925).

Anasarca was observed in EP1+/+ mice

A subset of EP1+/+ mice appeared to have substantial edema and displayed an increase in body weight, peaking approximately five days post-Ang II administration (Figure 2.5A). Average body weight in EP1 +/+ cohort subsequently decreased due to mortality in the animals with the largest weight gain. At baseline, EP1+/+ mice weighed more than EP1-/- mice though this difference was modest (EP1+/+ 26.6 ± 0.39 grams, EP1-/- 24.9 ± 0.68 grams, P = 0.024). Body weight of EP1-/- mice was unchanged over the course of the study. EP1+/+ mice had a significantly greater fraction water weight as compared to EP1-/- mice (Figure 2.5B, P = 0.0138), and aneurysm incidence was lower in mice which developed anasarca compared to mice without anasarca (33 % vs 76 %). Anasarca is commonly a result of liver failure, nephrotic syndrome or heart failure (168,169). Plasma alanine transaminase (ALT) activity was analyzed as a marker of liver function in Nphx/DOCA-NaCl/Ang II treated EP1+/+ mice. ALT activity was not elevated above baseline values and no correlation with body weight was observed (Figure 2.6).

A.



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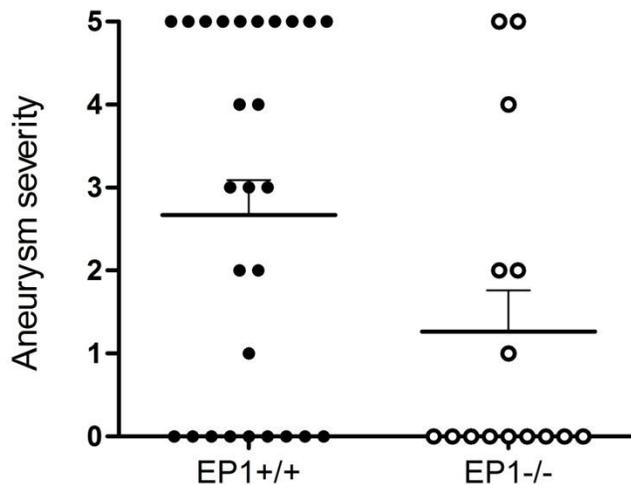


Figure 2.2. Nphx/DOCA-NaCl/Ang II model induces aortic aneurysm formation. A. Nphx/DOCA-NaCl/Ang II treatment resulted in formation of aneurysms and dissections in the thoracic and abdominal aorta. Aortae were scored visually upon necropsy and representative images are shown. B. Post-mortem examination revealed 18 of 27 EP1+/+ mice and 6 of 15 EP1-/- mice developed aneurysms as a result of the Nphx/DOCA-NaCl/Ang II model. Aneurysm severity was significantly reduced in EP1-/- mice compared to EP1+/+ mice. Fisher's exact test, Severity < 3 vs. ≥ 3 , $P = 0.0344$.

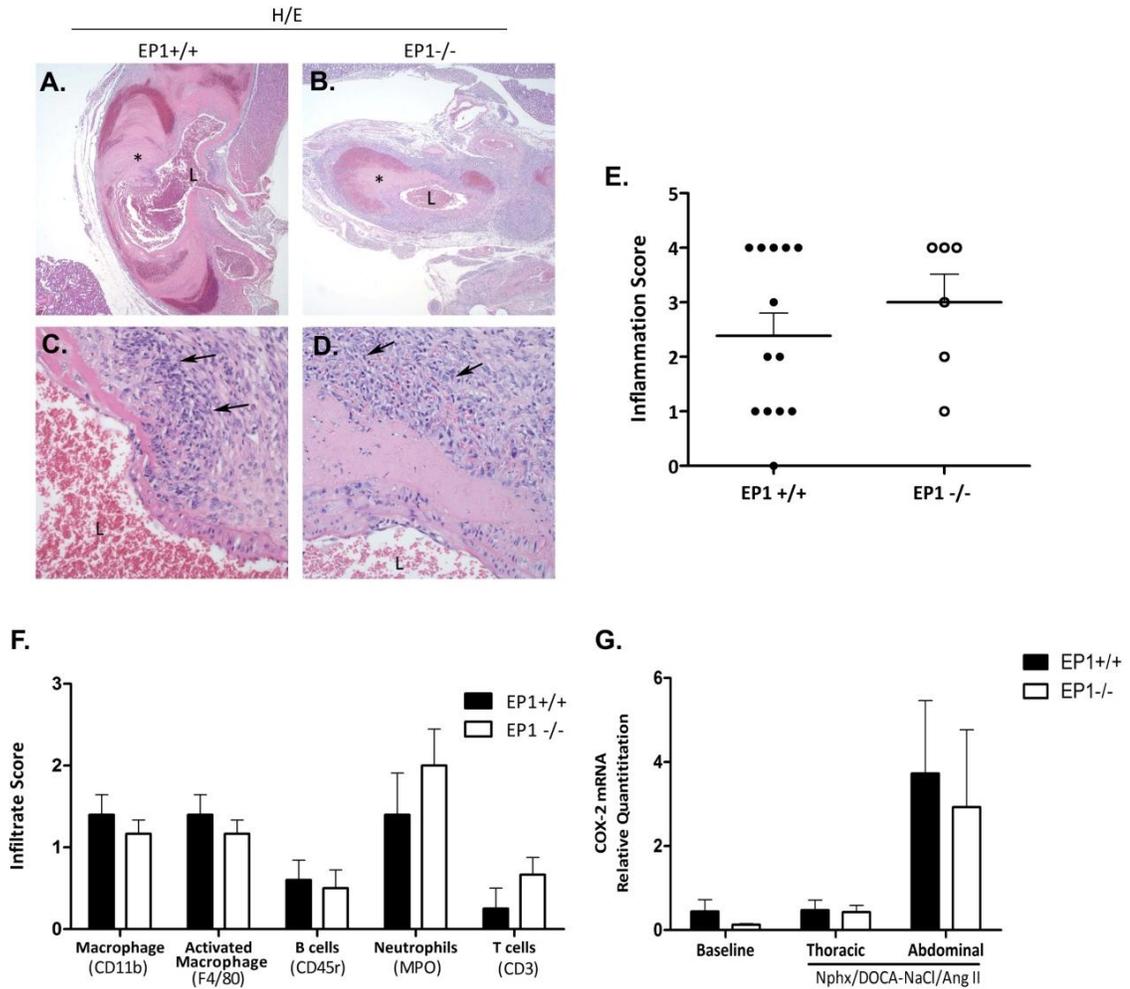


Figure 2.3. Aortic inflammation is observed in EP1+/+ and EP1-/- mice. A-D. Hematoxylin and eosin stained aortae (40X and 400X magnification). A ruptured aneurysm in an EP1+/+ mouse (A, C) and a large aneurysm without rupture in an EP1-/- mouse (B, D). Vessel necrosis and perivascular inflammatory infiltration of macrophages and neutrophils are observed under high magnification (arrows, C and D). L = vessel lumen, * = aneurysm. E. Overall inflammation was scored based on H/E stained aortic sections. No differences in inflammation were observed between EP1+/+ and EP1-/- aortas ($P = 0.3975$). F. Immunohistochemistry for detection of macrophage, B cells, T cells and neutrophils in aortae was performed and scored in a blinded fashion by a comparative veterinary pathologist. No differences in infiltrate were observed ($P > 0.05$). G. COX-2 mRNA in the thoracic and abdominal aortae 2-5 days post Ang II administration revealed a trend in increased COX-2 expression in the abdominal aorta of both EP1+/+ and EP1-/- mice post treatment. No significant differences were observed by treatment or between genotypes ($P > 0.05$).

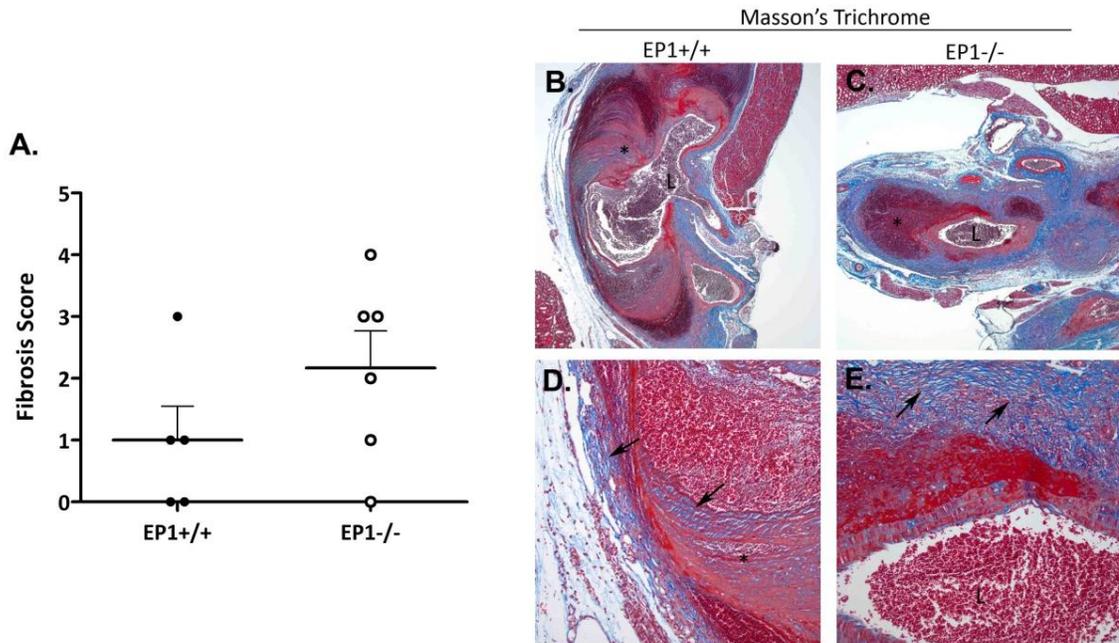
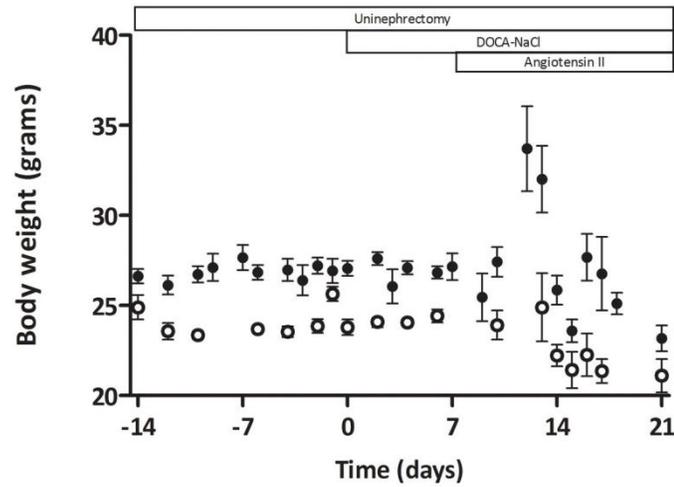


Figure 2.4. Aortic fibrosis is observed in EP1+/+ and EP1-/- mice. A. Extracellular matrix deposition was determined using Masson's Trichrome stain ($P = 0.1925$). B-E. Masson's trichrome stained aortae. EP1+/+ aorta (B, D) with rupture displays less collagen organization. EP1-/- aorta (C, E) with an intact aneurysm shows well developed and organized collagen (arrows) surrounding the aneurysm. L = vessel lumen, * = aneurysm.

A.



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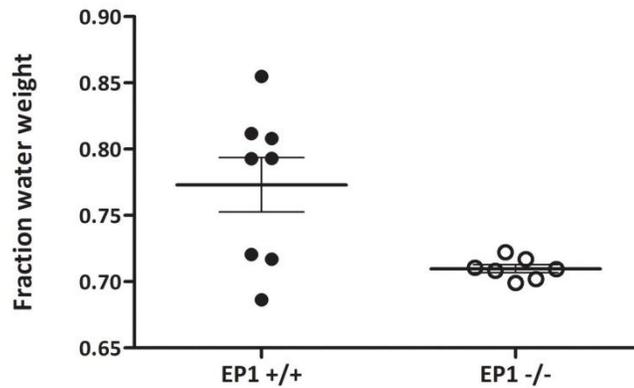


Figure 2.5. Body weight increases during Nphx/DOCA-NaCl/Ang II treatment in EP1+/+ mice. A. Average body weight of EP1+/+ (N=50) and EP1-/- mice (N=22) throughout the duration of the experiment. Body weight increased in EP1+/+ mice, but not EP1-/- mice, following Ang II administration. Average body weight declined at approximately five days post-Ang II due to mortality in the subset of mice which gained the most weight (open symbols, EP1-/-; closed symbols, EP1+/+). B. Fraction water weight 5 days post-angiotensin II administration. The fraction water weight was significantly greater in EP1+/+ mice, as compared to EP1-/- mice, $P = 0.0138$.

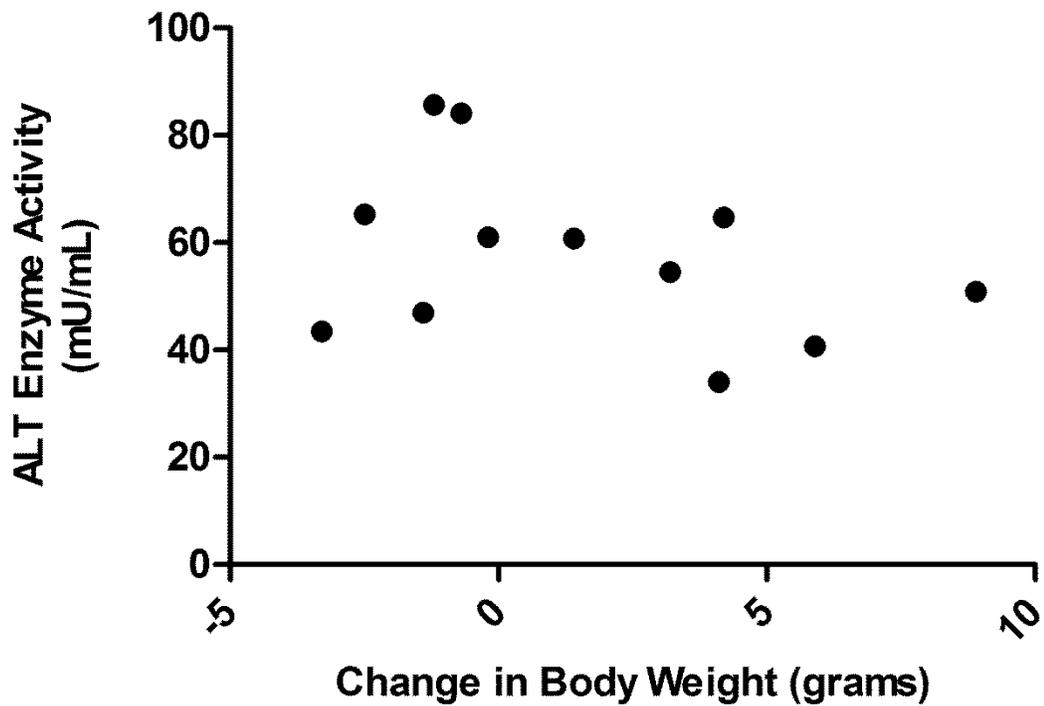
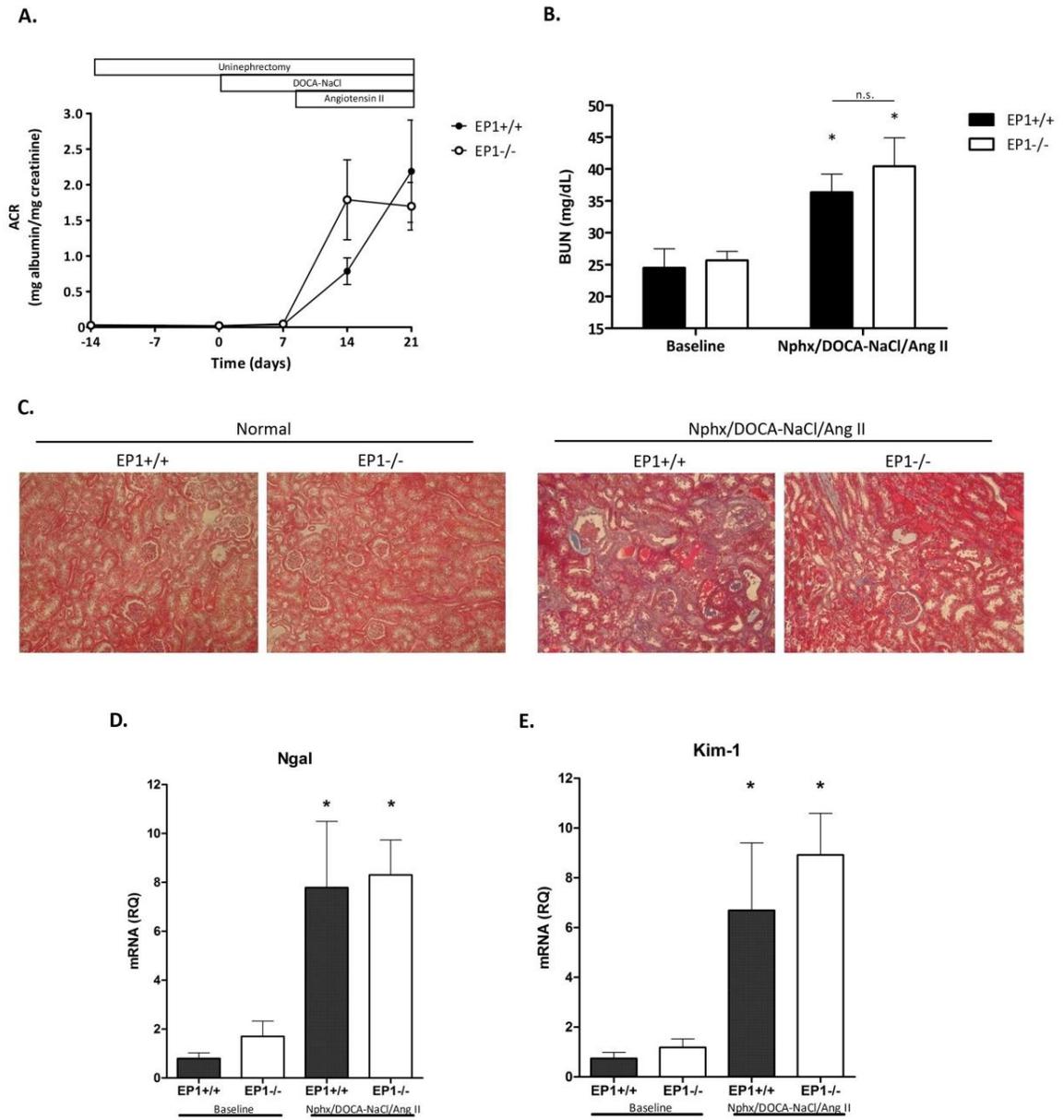


Figure 2.6. Plasma alanine transaminase activity and body weight showed no correlation in EP1+/+ mice. ALT activity was not elevated above expected baseline values (< 100 mU per mL) and no correlation with body weight was observed ($R^2 = 0.1368$).

Modest renal injury was induced in EP1^{+/+} and EP1^{-/-} mice

The Nphx/DOCA-NaCl/Ang II model was initially developed to induce hypertensive renal damage on the C57BL/6 background (163). To quantify renal damage in the EP1^{+/+} and EP1^{-/-} mice, we monitored urinary albumin excretion, blood urea nitrogen, renal histopathology, and biomarkers of acute kidney injury, neutrophil gelatinase-associated lipocalin (Ngal) and kidney injury molecule-1 (Kim-1) mRNA expression (Figure 2.7). ACR and BUN were elevated but no significant differences were observed between genotypes (Figure 2.7A, B). Renal histopathology showed modest hypertensive renal damage compared to the contralateral kidney removed at time of uninephrectomy (Figure 2.7C). Dilated tubules with moderate glomerulosclerosis and tubulointerstitial fibrosis were observed. Significant increases in Ngal and Kim-1 mRNA expression in the kidney of EP1^{+/+} and EP1^{-/-} mice were observed, though no differences were detected between genotypes (Figure 2.7D-E).

Figure 2.7. Modest renal damage is observed in EP1^{+/+} and EP1^{-/-} mice. A. Urinary albumin excretion. ACR was determined on spot urine collected throughout the duration of the experiment. High albumin levels were detected in the urine after the addition of Ang II and no significant differences were observed in ACR between the genotypes. Fisher's Exact test, $P = 0.4559$. B. Blood urea nitrogen levels were modestly increased by the end of the full model as compared to baseline in both genotypes (EP1^{+/+} $P = 0.0382$, EP1^{-/-} $P = 0.0132$), though no differences were observed in the degree of BUN levels between the genotypes ($P = 0.5867$). C. Renal histopathology. Masson's trichrome stain revealed modest hypertensive renal damage observed in both genotypes following full model treatment (right panels) and compared to normal kidneys removed at time of uninephrectomy (left panels). Specifically, dilated tubules with moderate glomerular sclerosis and tubulointerstitial fibrosis were observed. D and E. Quantification of Ngal and Kim-1 mRNA expression in whole kidneys. Expression of clinically used renal injury biomarkers, Ngal and Kim-1, revealed significant increases in both genotypes treated with the full model (Ngal: EP1^{+/+} $P = 0.0029$, EP1^{-/-} $P = 0.0008$; Kim-1: EP1^{+/+} $P = 0.007$, EP1^{-/-} $P = 0.0002$), though no significant differences between EP1^{+/+} and EP1^{-/-} mRNA levels were observed (Ngal $P = 0.8609$, Kim-1 $P = 0.4931$).



Cardiac function is reduced in EP1^{+/+} and EP1^{-/-} mice

No structural differences in the heart were observed by echocardiography between EP1^{+/+} and EP1^{-/-} mice at baseline. A modest increase in ejection fraction and fractional shortening was observed in EP1^{-/-} mice at baseline. At five days post-Ang II administration, EP1^{+/+} had increased left ventricular posterior wall (LVPWd) and interventricular septum diastolic diameters (IVSd). EP1^{-/-} mice had increased IVSd, though no significant change in LVPWd was observed. EP1^{+/+} and EP1^{-/-} mice displayed increased left ventricular interior diameter, though no differences were observed between genotypes. Cardiac function was significantly reduced in both genotypes upon treatment with Nphx/DOCA-NaCl/Ang II, as demonstrated by decreased fractional shortening and ejection fraction (Table 2.1). Additionally, heart weights of EP1^{+/+} and EP1^{-/-} mice after treatment with Nphx/DOCA-NaCl/Ang II showed no significant difference between the genotypes (EP1^{+/+}: 198.9 ± 9.6 grams N = 21, EP1^{-/-}: 190.0 ± 4.3 grams N = 17, P = 0.461).

Table 1. Echocardiography data at baseline and 5 days post Ang II treatment.

Parameter	Baseline		Nphx/DOCA-NaCl/Ang II	
	EP1+/+ (N = 9)	EP1-/- (N = 8)	EP1+/+ (N = 6)	EP1-/- (N = 8)
LVPWd (mm)	0.760 ± 0.015	0.772 ± 0.021	0.773 ± 0.021 †	0.794 ± 0.028
IVSd (mm)	0.812 ± 0.011	0.790 ± 0.019	0.921 ± 0.038 †	0.895 ± 0.010 †
LVIDd (mm)	3.058 ± 0.043	2.985 ± 0.040	3.551 ± 0.24 †	3.482 ± 0.13 †
FS (%)	49.225 ± 0.28	50.847 ± 0.13 ‡	31.784 ± 2.98 †	33.268 ± 2.31 †
EF (%)	81.917 ± 0.27	83.462 ± 0.15 ‡	60.228 ± 4.22 †	62.467 ± 3.20 †

† P < 0.05, Baseline vs Nphx/DOCA-NaCl/Ang II

‡ P < 0.05, EP1+/+ vs EP1-/-

Hypertension was less severe in EP1^{-/-} mice than EP1^{+/+} mice

To determine the effect of disruption of the EP1 receptor on blood pressure in Nphx/DOCA-NaCl/Ang II treated animals, intracarotid blood pressure was determined in EP1^{+/+} and EP1^{-/-} mice two days post-Ang II administration (Figure 2.8). MAP was significantly increased compared to untreated animals in both EP1^{+/+} (76.79 +/- 5.33 mm Hg baseline vs. 128.8 +/- 5.08 mm Hg, P = 0.0004) and EP1^{-/-} mice (74.36 +/- 5.61 mm Hg baseline vs. 102.4 +/- 7.77 mm Hg, P = 0.0423). However, the rise in MAP was significantly lower in EP1^{-/-} compared to EP1^{+/+} mice (P = 0.0295).

Reduction of blood pressure protected against mortality

Fifteen EP1^{+/+} and EP1^{-/-} mice treated with Nphx/DOCA-NaCl/Ang II were administered the antihypertensive agent hydralazine. Hydralazine treatment significantly reduced MAP in EP1^{+/+} mice (106.1 +/- 5.76 mm Hg vs. 128.8 +/- 5.08 mm Hg, P = 0.024), and but had no significant effect on EP1^{-/-} mice (Figure 2.9A, 99.47 +/- 4.82 mm Hg vs. 102.4 +/- 7.77 mm Hg, P = 0.762). A significant decrease in the incidence of mortality was observed in EP1^{+/+} but not EP1^{-/-} mice (Figure 2.9B, P = 0.007 and P = 0.642 respectively). Hydralazine treatment reduced aneurysm incidence and severity in EP1^{+/+} though this did not achieve statistical significance (Figure 2.9C, P > 0.05). Anasarca was not observed in hydralazine treated EP1^{+/+} (Body weight 5 days post-Ang II: 21.40 +/- 0.65 grams) as compared to untreated EP1^{+/+} mice (Body weight 5 days post-Ang II: 32.28 +/- 1.59 grams, P < 0.0001).

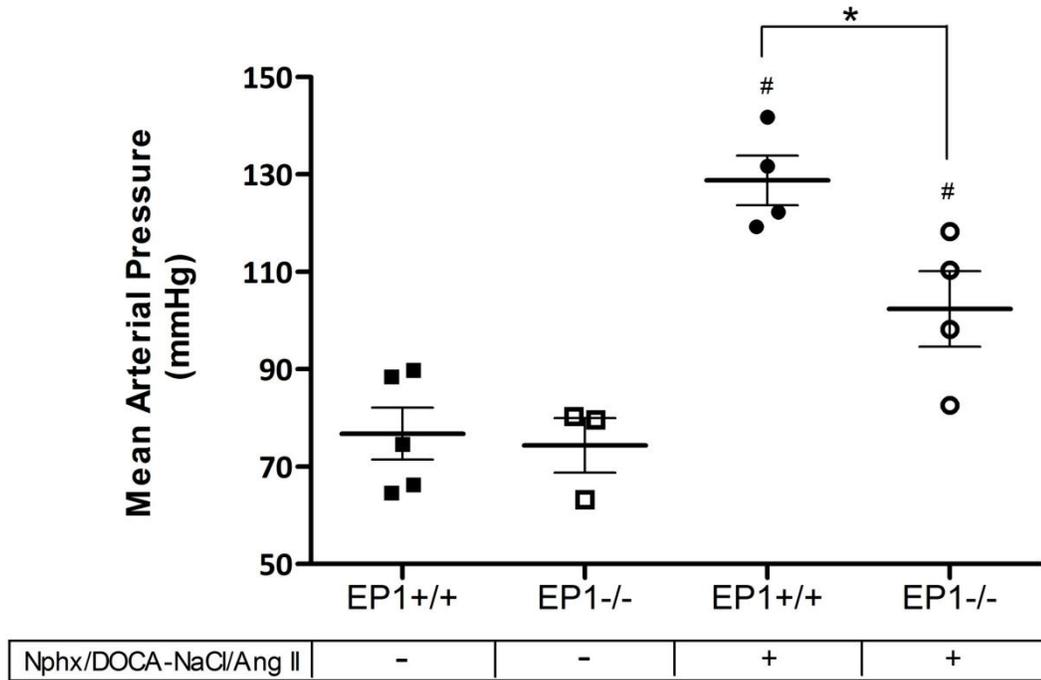
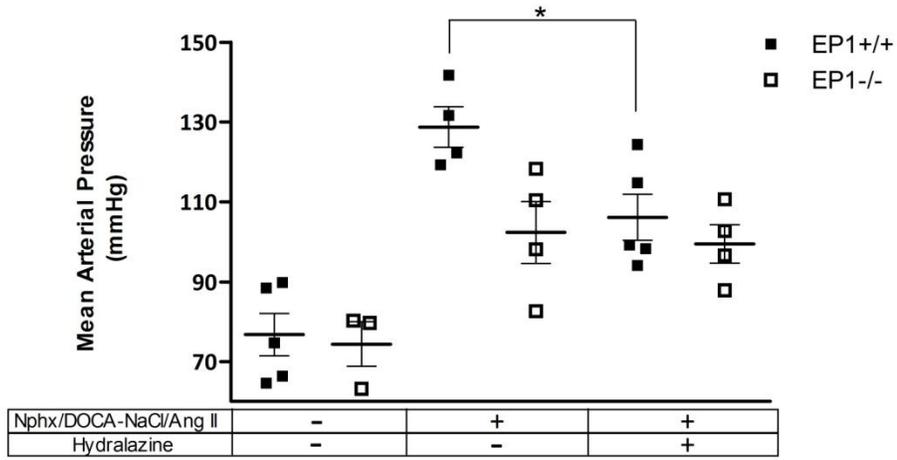


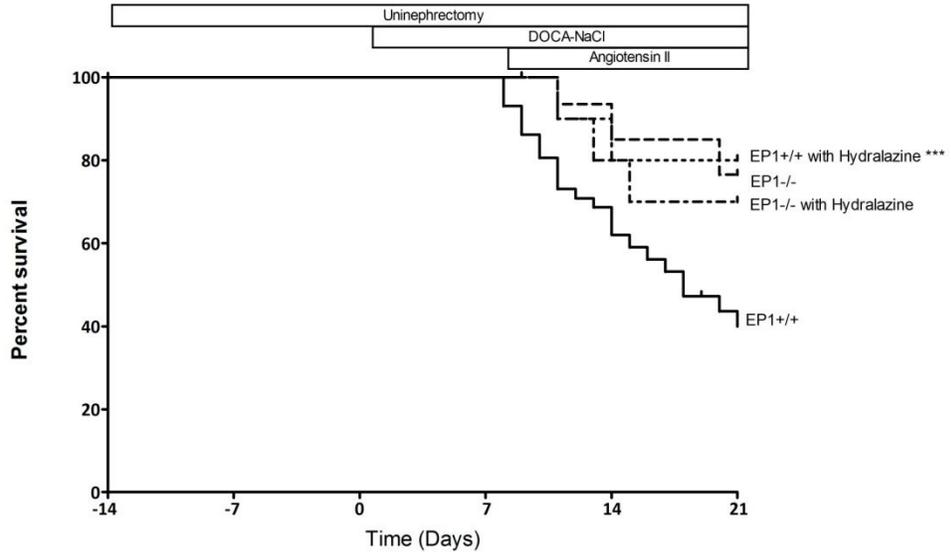
Figure 2.8. Mean Arterial Pressure is increased in EP1+/+, and blunted in EP1-/-, mice. Intracarotid blood pressure was measured in EP1+/+ and EP1-/- mice two days post-Ang II administration under ketamine and inactin anesthesia. MAP was elevated in both groups compared to baseline (EP1+/+ $^{##}P = 0.0004$, EP1-/- $^{#}P = 0.0423$) but was significantly lower in EP1-/- compared to EP1+/+ two days post-Ang II ($^{*}P = 0.0295$).

Figure 2.9. Hydralazine treatment reduced mortality in EP1+/+ but not EP1-/- mice. A. Mean arterial pressure was measured in EP1+/+ and EP1-/- mice treated with hydralazine at 2 days after Ang II administration and compared to existing data shown in figure 2.7. EP1+/+ with and without hydralazine, $P = 0.024$. EP1-/- with and without hydralazine, $P = 0.762$. B. Mortality observed in EP1+/+ mice was significantly reduced by treatment with hydralazine, $***P = 0.007$. No change was observed in EP1-/- mice, $P = 0.642$. C. Aortic aneurysm severity. Aneurysm severity was decreased in EP1+/+ mice treated with hydralazine ($P > 0.05$).

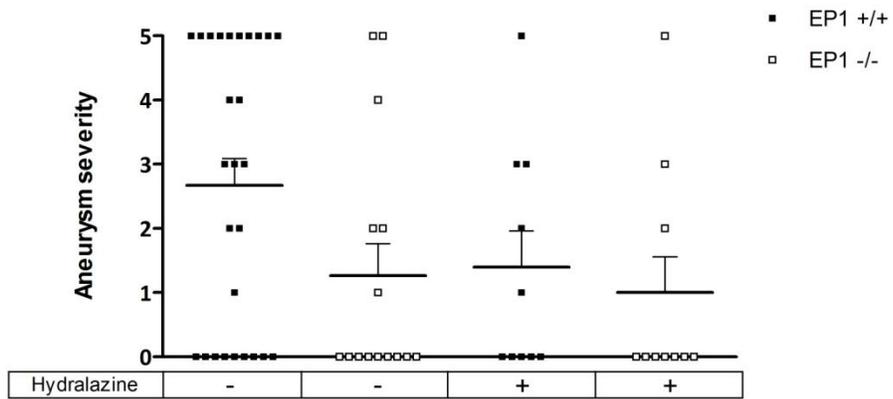
A.



B.



C.



Discussion

Disruption of the EP1 receptor affords substantial protection in the Nphx/DOCA-NaCl/Ang II evoked hypertension. The incidence of mortality was significantly decreased and appeared to result from reduction in MAP. Mortality was a result of ruptured aortic aneurysm or occurred after developing anasarca. The results presented here are consistent with previous studies demonstrating the role of EP1 in modulating the rise in MAP in response to Ang II (85), and further reveal the protective effect disruption of the EP1 receptor has on end-organ damage.

There are several limitations to these studies which deserve mention. First, high mortality observed in EP1+/+ mice confounds the analysis of measurements taken after implantation of Ang II, such as cardiac and renal function, since the analyses are only performed on surviving mice. Second, there was a modest (<10 %), though statistically significant difference in body weight observed between EP1+/+ and EP1-/- mice. Although the dosage of Ang II was adjusted by weight, the dosage of the DOCA pellet was not. However, one would predict the genotype receiving the greater dose/weight (EP1-/-) would have the worse phenotype, and this is opposite what we observed. Lastly, EP1-/- mice were observed to have lower blood pressure than EP1+/+ mice following treatment with the Nphx/DOCA-NaCl/ Ang II model. This is consistent with our previously published data (170) suggesting EP1 mediates part of Ang II-induced hypertension. In this model we measured MAP at baseline or after

treatment with all three model components; it is possible that EP1 also contributes to hypertension induced by Nphx or DOCA-NaCl as well.

Current models of aortic aneurysm include a combination of hyperlipidemic mice or high fat diet with modulation of the renin-angiotensin-aldosterone axis, or aberrant production of extracellular matrix components (171). Ang II-induced aortic aneurysms are characterized by accumulation of macrophages in the adventia and media, disruption of elastin fibers, expansion of the lumen, thrombus formation and disordered extracellular matrix deposition (172). These characteristics were also observed in the Nphx/DOCA-NaCl/Ang II model, although no significant differences in macrophage accumulation, matrix deposition, or COX-2 mRNA expression were detected between the two genotypes. It should be noted that the aneurysms and dissections observed in this model occur after acute severe hypertension and although the pathology appears similar to that observed in human disease, the disease genesis may not be. In humans, development of a true aneurysm is a slowly progressing disease initiating with local inflammation, disruption of the connective tissue matrix, and is often associated with atherosclerosis. In contrast, development of false aneurysm, or dissection as a result of a tear in the intima, can occur more acutely by a sudden large rise in blood pressure or direct injury and may be more representative of the damage induced by the Nphx/DOCA-NaCl/Ang II model. Our data demonstrate that protection observed when EP1 is disrupted is likely due to the prevention of a large rise in blood pressure, since treatment with

hydralazine phenocopied EP1^{-/-} mice. This does not eliminate the possibility that EP1 receptors might also provide protection directly at the target tissue.

Data exist suggesting a role for prostaglandins, in particular PGE₂, in aortic aneurysm formation. COX-2 initiates the production of prostaglandins, and its expression is induced by infusion of angiotensin II in the smooth muscle of the aorta surrounding aneurysms (166). Furthermore, either selective inhibition of COX-2 or genetic deletion of COX-2 significantly reduced aortic aneurysm formation and macrophage infiltration (166,167). Deletion of microsomal PGE synthase-1, which transforms the product of COX-2 metabolism into PGE₂, has also been demonstrated to reduce aortic aneurysm formation and oxidative stress in LDLR^{-/-} mice with an angiotensin II infusion (160), suggesting PGE₂ plays an important role in development of aneurysms and the EP receptors may be viable targets for treatment of aneurysm progression.

Previous reports of the role of EP1 in renal injury are contradictory. In spontaneously hypertensive rats, treatment with an EP1 antagonist reduced proteinuria and tubulointerstitial damage (87), while in anti-GBM nephrotoxic serum nephritis genetic deletion of EP1^{-/-} in mice resulted in enhanced mesangial expansion and tubular dilation and increased blood urea nitrogen and serum creatinine (173). In our studies, modest hypertensive renal damage was observed, although no significant differences in renal function were detected between genotypes. However, our interpretation was confounded by the differential mortalities in EP1^{+/+} and EP1^{-/-} mice, potentially biasing our results. Examination of renal histopathology at time points prior to significant mortality

failed to detect any severe renal damage or differences between the genotypes. This suggests the role of EP1 in renal damage is highly context dependent.

Anasarca, or extreme generalized edema, can occur in many disease settings. It is commonly a result of liver failure, nephrotic syndrome or heart failure (168,169). In our Nphx/DOCA-NaCl/Ang II model, a subset of EP1+/+ mice developed severe anasarca prior to mortality, while EP1-/- mice were protected. The EP1 receptor has previously been shown to be natriuretic (85). With this paradigm, one might predict EP1-/- mice would retain more salt and water; however in our results we demonstrate that EP1+/+ mice gain excessive fluid volume that is not observed in EP1-/- mice. This contradiction leads us to conclude that alterations in kidney function by disruption of EP1 do not play a dominant role in development of the observed edema. Additionally, cardiac function was reduced to similar degrees in EP1+/+ and EP1-/- mice. Edema was prevented by treatment with hydralazine, suggesting elevation in blood pressure was responsible for development of edema. We hypothesize that hypertension induced by DOCA-NaCl and Ang II results in volume loading and enhanced vasoconstriction, which places excessive stress on the vascular wall leading to enhanced permeability, resulting in edema and susceptibility to dissections and rupture. Future experiments will be required to identify whether vascular permeability differences are observed between EP1+/+ and EP1-/-mice.

The EP1 receptor plays an important role in the development of hypertensive damage. In the Nphx/DOCA-NaCl/Ang II model, disruption of EP1 results in increased incidence of survival, lessened aneurysm severity and the

absence of anasarca. This effect is a result of a reduced rise in blood pressure observed in EP1^{-/-} mice, and suggests the EP1 receptor may be a viable pharmaceutical target for the treatment of hypertension and subsequent organ damage. Furthermore the Nphx/DOCA-NaCl/Ang II model may prove to be a useful tool for studying the pathology of aortic aneurysm and dissection formation in a setting of acute severe hypertension.

CHAPTER III

DISRUPTION OF EP3 IS PROTECTIVE AGAINST MORTALITY IN A MOUSE MODEL OF HYPERTENSION

Introduction

There is a strong association between hypertension and progressive renal failure, and mitigation of hypertension is a major therapeutic goal for the prevention of end-stage renal disease. A number of pharmacologic agents are available for the treatment of hypertension including those that affect the renin-angiotensin aldosterone system such as angiotensin converting enzyme inhibitors, angiotensin receptor blockers and more recently, the introduction of renin inhibitors (11,174-178). Angiotensin II mediates its effects via two GPCRs, designated AT1 and AT2, which are distinguished by their pharmacology and the signal transduction pathways that they activate (175,176,179). The AT1 receptor is the target for the anti-hypertensive receptor antagonist ARBs (180). AT1 receptor activation leads to a number of signal transduction pathways including increases in intracellular Ca^{++} and activation of cPLA₂ (180-188), a critical regulatory step in the formation of PGs. PGs, which are oxygenated metabolites of the essential fatty acid arachidonic acid, are themselves modulators of blood pressure and evidence suggests that blockade of one or more prostaglandin receptors may be useful for the treatment of hypertension. Recent evidence

suggests that at least one PGE₂ receptor may play a role in the actions of the RAAS on blood pressure (85).

PGs are potent mediators of a wide range of physiological actions including inflammation, modulation of smooth muscle tone and water and ion transport in the kidney (98,189). The five primary bio-active prostanoids PGE₂, PGF_{2α}, PGD₂, PGI₂, and TXA₂, activate a family of specific GPCRs, EP for E-prostanoid receptors, FP, DP, IP and TP receptors respectively (98). The EP receptors are unique among the PG receptors, in that four receptors, designated EP1 through EP4, have been described for PGE₂ each encoded by a distinct gene. PGE₂ has been demonstrated to act as either a pressor or depressor depending upon the EP receptor activated (17,19,79,99,157,190-193). Upon acute infusion, PGE₂ is a vasodepressor in both humans and mice (17,138,150). Previous studies with knockout mice have shown that this depressor effect is primarily due to the activation of the EP2 receptor (139). When the EP2 receptor is deleted, the loss of the depressor response unmasks a PGE₂ pressor response, suggesting a balance of pressor and depressor receptors activated by PGE₂. The pressor response is mediated by the EP3 and EP1 receptors (85,139).

Development of models for the study of hypertension and renal damage that recapitulate human disease have been challenging, particularly on the C57BL/6 background (194,195). A model of hypertension that causes renal damage on the C57BL/6 background has been recently reported (163). This model which employs a combination of uninephrectomy, deoxycorticosterone

acetate (DOCA), high sodium intake and Ang II, was reported to initiate renal damage over a relatively short time course of five weeks. Using this model, the effects of the EP3 receptor on hypertension and renal damage were evaluated.

Experimental Procedures

Materials

Angiotensin II was purchased from EMD Gibbstown, NJ, and DOCA was purchased from Innovative Research of America, Sarasota, FL, USA. Osmotic minipumps were purchased from DURECT Corporation, USA.

Induction of Hypertension

EP3^{-/-} or EP3^{+/+} mice were uninephrectomized under ketamine/xylazine anesthesia two weeks before the start of the study. At day 0, mice received subcutaneous implantation of a 50 mg DOCA pellet and were given ad libitum access drinking water containing 1 % NaCl. At day 7, an osmotic mini-pump (Alzet model 1002) delivering 1.5 ng/min/gram body weight angiotensin II was implanted subcutaneously. At day 19 or 21, the animals were euthanized. Tissues were removed for examination by histological study or snap frozen for RNA and protein analysis. Four separate studies were performed with six to eight EP3^{-/-} animals in each study and seven or eight EP3^{+/+} littermates or commercially obtained C57BL/6J mice (The Jackson Laboratory, Bar Harbor,

ME). A total of 34 EP3^{+/+} mice and 27 EP3^{-/-} mice were used. All experimental studies were approved by the IACUC of Vanderbilt University Medical Center.

Measurement of Systolic Blood Pressure

Systolic BP was determined in conscious mice using a computerized tail-cuff system (Visitech systems BP-2000 Blood Pressure Analysis System, Apex NC, USA) in the Mouse Metabolic Phenotyping Core at Vanderbilt University Medical Center. Mice were trained for four days minimizing physiologically apparent stress. Each measurement is the average of at least 10 consecutive readings after stabilization of blood pressure.

Intracarotid blood pressure measurement

Intracarotid blood pressure was measured under ketamine (25 mg/kg) and inactin (100 mg/kg) anesthesia delivered intraperitoneally. Mice were placed on a thermal pad and a PE-10 catheter was inserted into the left carotid artery. The catheter was connected to a TXD-310 transducer and blood pressure was measured using a Digi-Med BPA 400 (Micromed). Mice were equilibrated 30-60 minutes until stable values were attained. Ten minute blood pressure measurements were collected and average mean arterial pressure (MAP) is plotted.

Determination of Blood Urea Nitrogen levels

To assess renal function, BUN levels were determined using Infinity Urea liquid stable reagent (Thermo scientific, USA). Heparinized blood was obtained from saphenous vein and plasma was stored at -80 °C until assayed.

Assessment of Renal Histopathology

Mice were humanely euthanized and tissues fixed overnight in 10 % neutral buffered formalin. Tissues were then processed routinely, embedded in paraffin, sectioned at 5 microns, stained with hematoxylin and eosin and evaluated by light microscopy. Kidney injury was scored by a renal pathologist who calculated the percent of tubules with cell necrosis, loss of brush border, cast formation, and tubular dilation as follows: 0, none; 1, <10 %; 2, 11-25 %; 3, 26-45 %; 4, 46-75 %; 5, >76 %. At least 10 fields (x200) were reviewed for each slide in a blinded fashion.

Statistical Analysis

Data were analysed using ANOVA or Student's *t* test were performed with GraphPad Prism software (GraphPad Software Inc., CA, USA) and are shown as means \pm SEM. Survival curves were evaluated by the Kaplan-Meier method. $P < 0.05$ was considered statistically significant for all studies.

Results

Uninephrectomy/DOCA-NaCl/Ang II model of Hypertension caused significant mortality.

C57BL/6 animals were treated using the recently reported uninephrectomy/DOCA-NaCl/Ang II model of hypertension (163). In contrast to the published report, this model resulted in significant mortality after implantation of the Ang II minipumps. Deletion of the EP3 receptor was protective; EP3^{+/+} mice having 36 % survival while EP3^{-/-} mice had 53% survival (P =0.037; Fig. 3.1).

In the EP3^{+/+} group, mice displayed severe anasarca, while EP3^{-/-} rarely developed anasarca even in the subset of mice that died. This is reflected in the change in body weight of mice from the initiation of the study to time of death, or the end of the study (Fig. 3.2A). While EP3^{+/+} had a marked increase in body weight, the EP3^{-/-} mice had no significant change in body weight. This difference in body weight was transient over the course of the study (Fig 3.2B). The transient change in body weight observed in the EP3^{+/+} group was a result of higher mortality observed in mice with the greatest increase in body weight (Figs. 3.2).

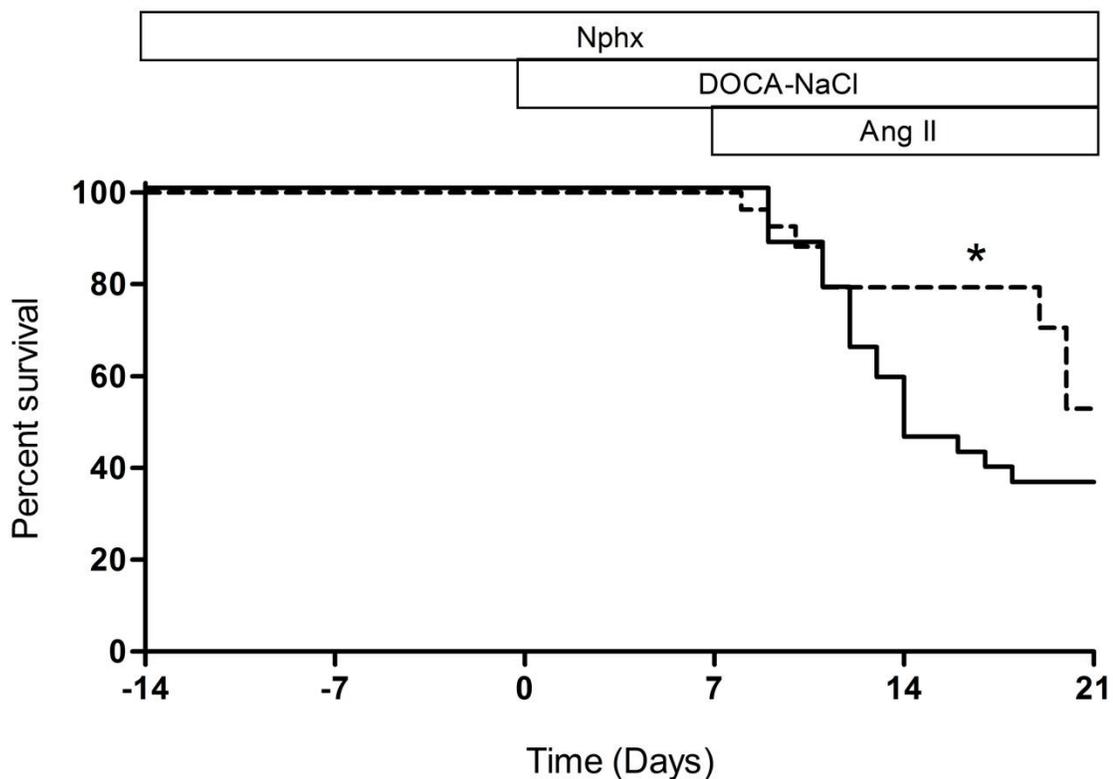


Figure 3.1. The uninephrectomy/DOCA-NaCl/Ang II model resulted in significant mortality. Significant mortality was observed in both EP3+/+ (solid line), and EP3-/- genotypes (broken line). The survival curves of two groups were compared using Kaplan-Meier analysis. Initial EP3+/+, N = 34, EP3-/-, N = 27, P = 0.037.

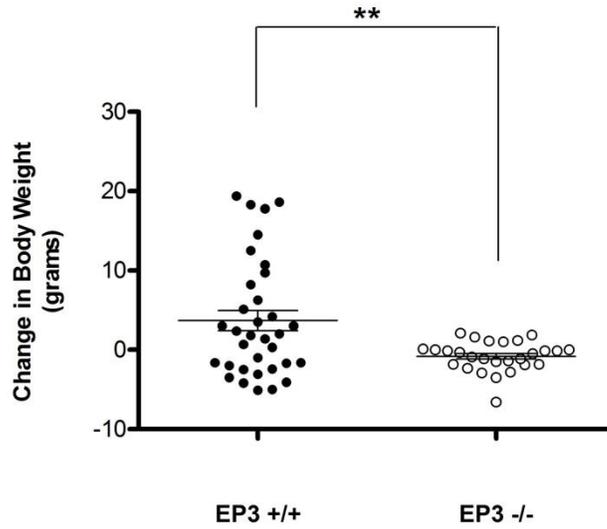
Blood pressure was elevated in both genotypes, but less in EP3^{-/-} mice

The EP3 receptor has vasopressor action, and therefore we hypothesized that significant changes in blood pressure between the two groups might underlie the decreased mortality in the EP3^{-/-} mice. Baseline blood pressure was not significantly different between genotypes in untreated animals (EP3^{+/+} 108.8 ± 1.7 mm Hg, EP3^{-/-} 107.1 ± 1.6 mm Hg P = 0.47, Figure 3.3A). Concurrent treatment with DOCA salt water and Ang II for two weeks resulted in elevated blood pressure as reported previously in this model. Although there was a dramatic increase in systemic blood pressure over time in both experimental groups terminal SBP was not different between the two groups (EP3^{+/+} 182.6 ± 5.0 mm Hg, EP3^{-/-} 193.1 ± 4.5 mm Hg P = 0.131; Fig. 3.3B). However, measurement of blood pressure by direct carotid catheterization under anesthesia at 2 or 14 days post Ang II revealed a significantly lower MAP in EP3^{-/-} mice as compared to EP3^{+/+} mice (EP3^{+/+} 129.7 ± 2.8 mm Hg, EP3^{-/-} 116.6 ± 5.4 mm Hg, P = 0.0314, Figure 3.3C).

EP3^{+/+} and EP3^{-/-} mice have modest renal damage

The uninephrectomy/DOCA-NaCl/Ang II model was developed to induce hypertensive renal damage on the C57BL/6 background. Histologic analyses indicated similar degrees of renal injury in the EP3^{+/+} and the EP3^{-/-} mice. The kidneys were characterized by multifocal, segmental to global glomerular microangiopathy and acute renal tubular necrosis with protein casts. Distinction between the EP3^{+/+} and EP3^{-/-} renal damage was limited to the renal tubules,

A



B

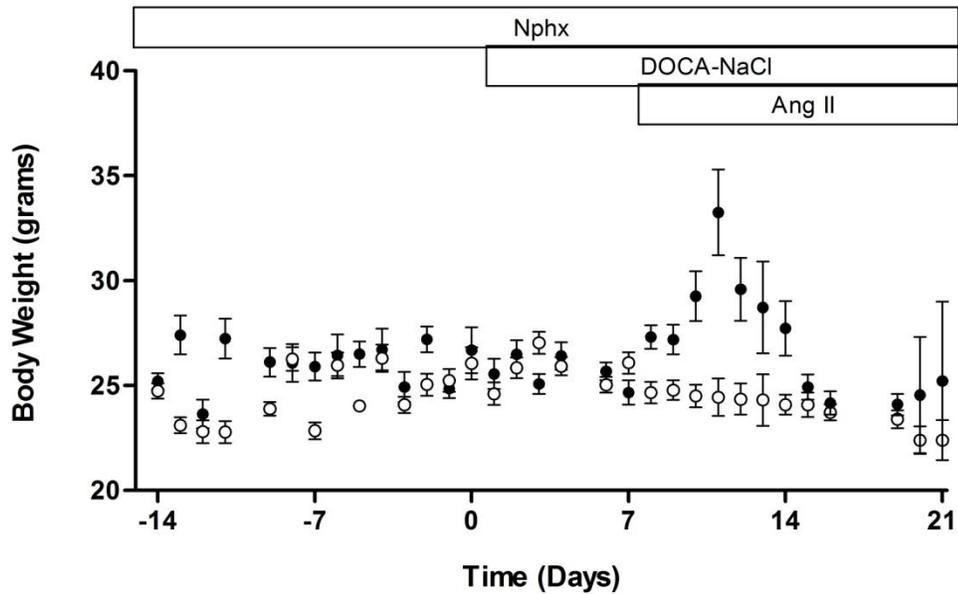
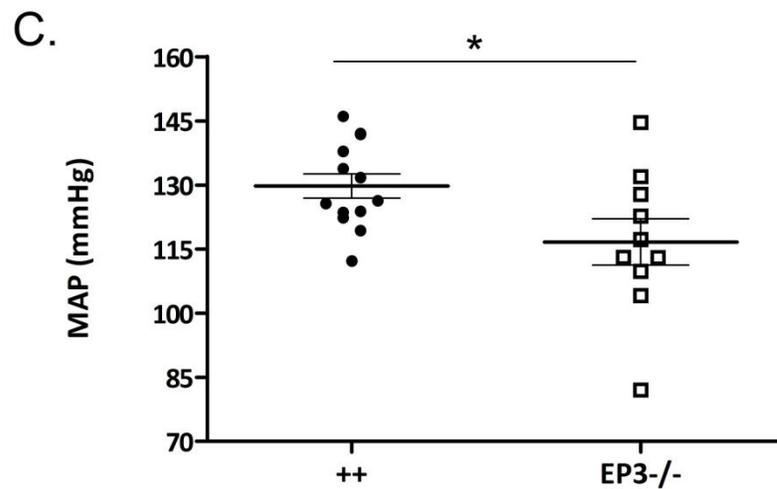
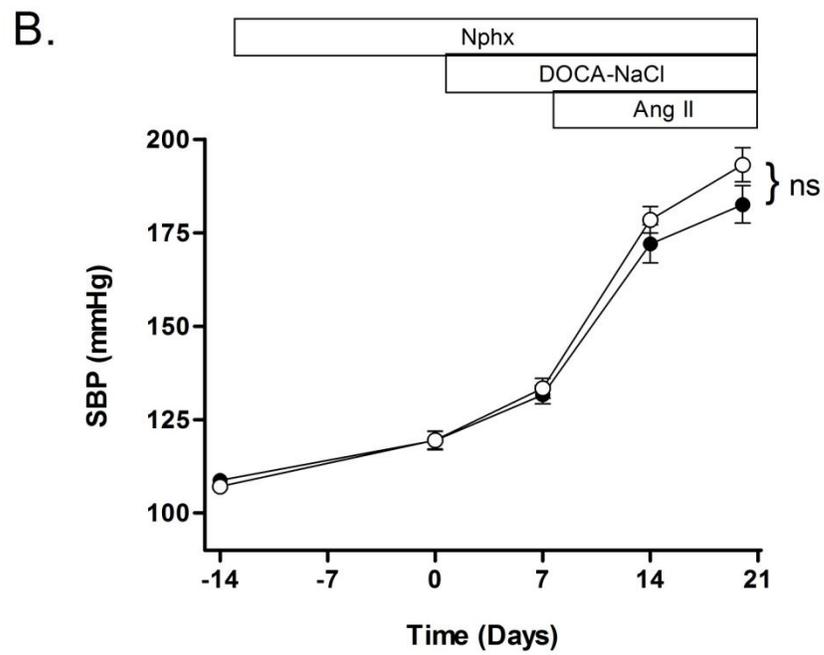
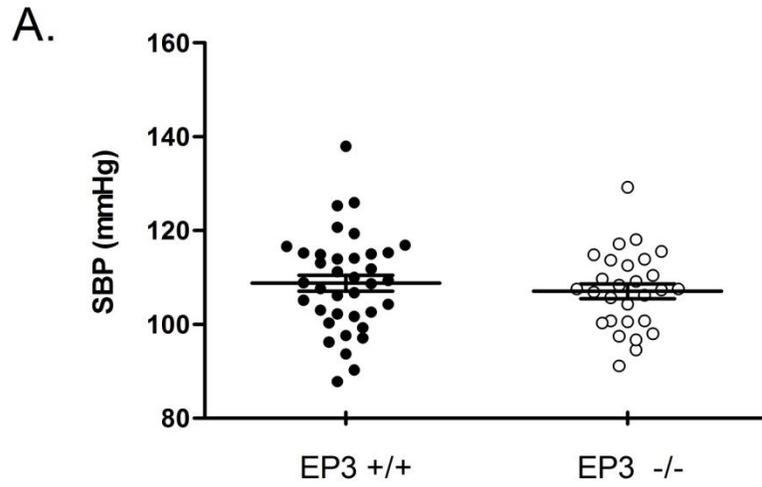


Figure 3.2. Body weight increased in EP3+/+ mice but not EP3-/- mice. A. The difference in terminal body weight and initial body weight was determined. EP3+/+ gained significantly more weight than EP3-/- mice. EP3 +/+, n = 34, EP3-/- n= 27 ** P = 0.0032. B. After implantation of the Ang II pump EP3+/+ mice increased body weight. Initial EP3+/+, n = 34, EP3-/-, N = 27.

Figure 3.3. Both EP3^{+/+} and EP3^{-/-} animals had significant increases in systolic blood pressure (SBP). A. Baseline SBP was measured in untreated EP3^{+/+} or EP3^{-/-} mice by the tail cuff method (EP3^{+/+}, n = 37, EP3^{-/-}, n= 28, P = 0.47). B. SBP for the two groups over the course of the hypertension study are shown. Data were analyzed by two-way ANOVA (genotype and time). Initial EP3^{+/+}, n = 34, EP3^{-/-}, n= 27, all surviving mice were analyzed at each time point. There was a statistically significant change in blood pressure over baseline with time in each genotype (P < 0.0001). There was no effect of the genotype on SBP over time (P = 0.29). C. Intracarotid blood pressure was measured at 2 and 14 days post Ang II. MAP was significantly lower in EP3^{-/-} mice as compared to EP3^{+/+} mice (P = 0.0314). Wildtype data from figure 2.7 are replotted as EP3^{+/+} MAP.



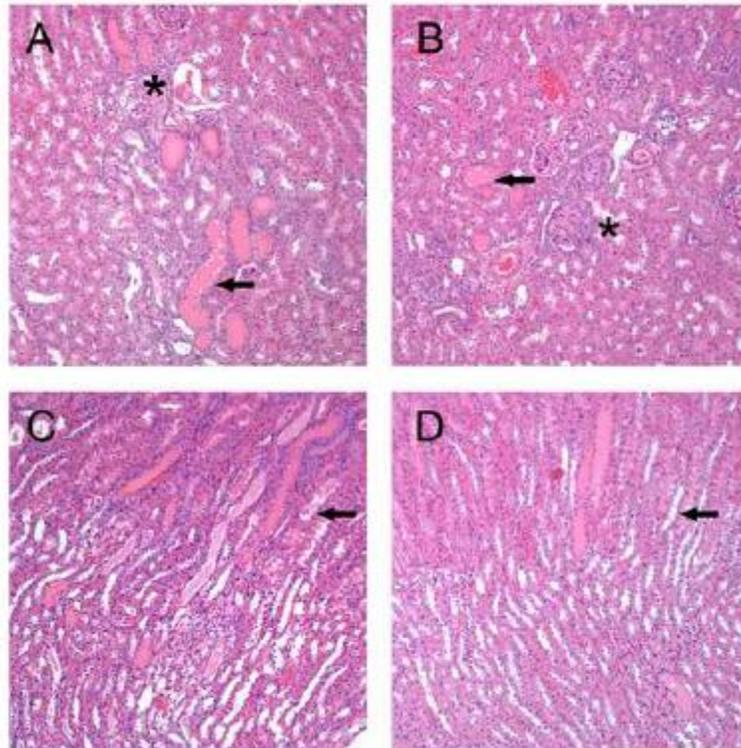


Figure 3.4. Modest end organ renal damage is observed in EP1+/+ and EP1-/- mice. A. H/E staining reveals glomerular microangiopathy and proteinaceous casts in the EP3+/+ and B. EP3-/- renal cortex (200x). *Asterisks* indicate glomeruli with segmental microangiopathies. *Arrows* indicate proteinaceous tubular casts. There is a mild decrease in tubular disease in the EP3-/- as compared to EP3+/+ kidneys. C. H/E staining of the corticomedullary junction in the EP3+/+ mouse kidney shows acute tubular necrosis and proteinaceous cast formation. In the more severely affected areas, tubular epithelial cells are swollen and vacuolated and lose intercellular and basement membrane attachments (*Arrows*). D. There is a modest decrease in damage seen in the EP3-/- mice (200x). Kidney tissue from eight EP3+/+ and ten EP3-/- mice were examined.

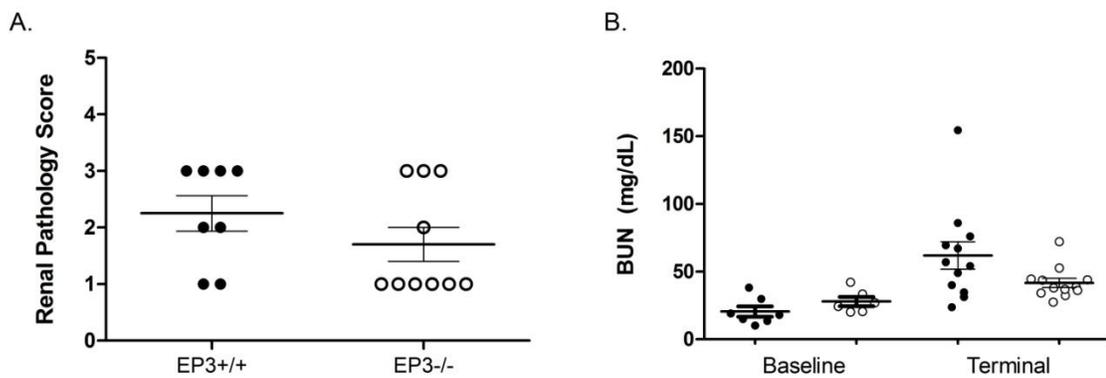


Figure 3.5. Quantitation of renal damage. A. Kidney injury was scored by pathologist Kelli Boyd, DVM as described in the Methods section. At least 10 fields (x200) were reviewed for each slide in a blinded fashion. No difference was seen between EP3+/+ and EP3-/-. B. Blood Urea Nitrogen (BUN) was determined as described in the Methods section. A significant increase in BUN was observed for both genotypes, comparing terminal BUN (n = 11 or 12 in each group) to baseline BUN values (n = 6 or 7 in each group, 2 way ANOVA P < 0.0001). The contribution of genotype to the BUN value was not significant.

where the tubular injury was slightly decreased in the EP3^{-/-} mice as compared to EP3^{+/+} mice (Fig. 3.4). Overall renal damage was quantitated as described in the methods section, and no significant differences were observed in renal histopathological score (Fig 3.5A). Similarly, although both EP3^{+/+} and EP3^{-/-} genotypes exhibited elevated BUN levels when subjected to the Nphx/DOCA-NaCl/AngII model ($P < 0.0001$) no statistically significant increase in terminal BUN of EP3^{+/+} as compared to EP3^{-/-} animals was observed (baseline: EP3^{+/+} BUN 20.6 ± 3.8 mg/dL, EP3^{-/-} 28.0 ± 3.5 mg/dL; terminal: EP3^{+/+} BUN = 53.7 ± 6.0 mg/dL, EP3^{-/-} 41.8 ± 3.4 mg/dL 2 way ANOVA $P > 0.05$; Fig. 3.5B).

Discussion

These studies demonstrated that deletion of the EP3 receptor resulted in decreased mortality and modestly reduced hypertension in a mouse model of severe hypertension. Although both EP3^{+/+} and EP3^{-/-} animals displayed increased BUN and pathological changes in the kidney, these changes were not significantly different between the genotypes. These studies were carried out on the C57BL/6 background, which is normally resistant to renal damage. A recent report described this model to induce hypertensive end organ damage on the C57BL/6 background strain (163). We were able to induce renal damage in the present studies, although we observed substantial differences compared to the reported findings. Most strikingly the EP3^{+/+} mice developed severe edema and mice in both groups experienced significant mortality. This was surprising as no

mortality was reported in the original description of the model, nor was a gain in weight reported. The EP3 receptor opposes the diuretic action of vasopressin in the renal cortical collecting duct (196,197). Loss of EP3 receptor might be expected to affect water balance, but in this case it should increase water retention in the EP3^{-/-} animals, the opposite of the observed effect of anasarca in the EP3^{+/+} mice. Previous studies have shown that mice with a disruption of the EP3 receptor concentrated urine normally in response to a range of physiologic stimuli, again suggesting that renal effects of the loss of the EP3 receptor are not playing a critical role in the phenotypic differences observed here (117).

Using tail cuff measurements, we observed SBP of near 200 mm Hg, almost 60 mm Hg higher than observed in the previous report. No deaths were observed in our study until after implantation of the Ang II pump, suggesting that the presence of exogenous Ang II is critical to the cause of death. Deletion of the EP3 receptor was associated with increased survival from 36 % in the EP3^{+/+} group to 53 % in the EP3^{-/-} group. The EP3 receptor has been implicated in the pressor effect of PGE₂ (139). It might be anticipated that deletion of this receptor would lead to lower blood pressure, and this would underlie its protective effect. In the present studies we noted that the EP3^{-/-} mice had modestly reduced hypertension compared to EP3^{+/+} mice. This suggests the protective effect of EP3 may be due to changes in blood pressure. These results are consistent with studies presented in Chapter II. Furthermore, the degree of hypertension correlates with protection against mortality. Given these results, these studies

that blockade of the EP3 and/or EP1 receptor would be protective from hypertension and its downstream consequences.

CHAPTER IV

CONTRIBUTION OF THE EP1 RECEPTOR IN HYPERTENSIVE RENAL DAMAGE

Introduction

Chronic kidney disease (CKD) is a major public health concern. The 2010 annual report from the United States Renal Data Systems estimated approximately 600,000 patients had end-stage renal disease (ESRD) in 2008, which cost the US \$39.5 billion/year. Additionally, the incidence and prevalence of CKD continues to rise in the United States creating a need for novel therapeutic agents (198).

Development of new onset kidney disease is strongly associated with hypertension, diabetes, smoking and low HDL cholesterol (199). In men, baseline blood pressure $\geq 120/80$ mm Hg strongly increased the development of ESRD as demonstrated by the Multiple Risk Factor Intervention Trial (200). Several other randomized clinical trials have also demonstrated correlations between hypertension and CKD (201-208), including the RENAAL (Reduction of Endpoints in NIDDM with Angiotension II Antagonist Losartan) trial which showed that each 10 mm Hg rise in systolic blood pressure resulted in an increased risk of ESRD or death of 11 % (209). Therefore, the treatment of renal disease is very closely tied to the treatment of hypertension.

Kidney disease is characterized by changes that involve renal inflammation followed by interstitial fibrosis, atrophy of the renal tubules, and glomerulosclerosis (210). Agents that block the renin-angiotensin-aldosterone system, such as ARBs, ACEi, and renin inhibitors, are among the most commonly used therapeutics for treatment of hypertension and renal damage. AT1 receptors mediate the majority of Ang II induced effects. AT1 is a G protein coupled receptor, which canonically couples to G_q G-proteins, but is also known to activate multiple second messenger signal transduction pathways including extracellular signal-regulated kinases 1 and 2, activation of phospholipases, inhibition of adenylate cyclase, and stimulation of tyrosine phosphorylation and Akt (211). Although classically Ang II has been thought to primarily modulate blood pressure and renal damage through vasoconstriction and aldosterone-induced sodium retention, it is now appreciated to also have effects on proteinuria, inflammation, proliferation, apoptosis, and fibrosis (212,213), thus the benefits of blockade of RAAS often extend beyond that of just blood pressure control (178,214-222). In these studies, the ARBs and ACEi had effects that occur in the absence of a blood pressure change or had a greater change than observed with a drug which reduced blood pressure comparably. One such example utilized a 5/6th nephrectomy model on male Munich-Wistar rats. Rats were treated with either an ACEi, enalapril, or a triple therapy of reserpine, hydralazine and hydrochlorothiazide (RAAS independent). Both treatments normalized blood pressure, although only enalapril was able to reduce the development of proteinuria and glomerulosclerosis, suggesting the

renoprotective effects of enalapril are independent of blood pressure reduction (216).

As mentioned previously, agonism of AT1 receptors can activate phospholipases, which yields products such as prostaglandins. PGE₂ is a known modulator of blood pressure and renal disease. PGE₂ can act as either a vasopressor or a vasodepressor (17-19,138). This antagonistic property of PGE₂ can be explained by the existence of four PGE₂ receptors, designated EP1 through EP4, each with distinct tissue localization and characteristic signal transduction pathways. Using pharmacologic agents and genetics, it has been demonstrated that the EP1 and EP3 receptors primarily mediate the pressor response of PGE₂, while the EP2 and EP4 receptors mediate the depressor response (17,79,85,99,134,138-140). Of most interest, the EP1 receptor has been shown to mediate Ang II-induced hypertension (85). EP1^{-/-} mice have blunted pressor responses to both acute and chronic Ang II administration (85). Furthermore, in isolated vessel preparations pre-treatment with the EP1 selective antagonist SC51322 reduced Ang II mediated vasoconstriction (85). Given that EP1 mediates some of the blood pressure and vasoconstrictor effects of Ang II and RAAS, it is of great interest whether EP1 contributes to renal fibrosis and CKD, possibly in a blood pressure independent manner similar to that of AT1.

EP1 blockade has been shown to positively affect renal function in a rodent model of human malignant hypertension (87). Stroke-prone spontaneously hypertensive rats were treated with vehicle or a selective EP1 antagonist. Although no differences were observed in blood pressure, treatment

with the EP1 antagonist reduced tubulointerstitial fibrosis, lessened urinary protein excretion, and blunted the drop in plasma creatinine levels. This suggests blockade of the EP1 receptor has a positive effect on renal function and may be a therapeutically relevant target (87). Nonetheless, it is imperative to continue investigating the contribution of EP1 to renal damage in order to demonstrate whether this is model specific and can be observed with both pharmacologic as well as genetic manipulation. Therefore, EP1^{+/+} and EP1^{-/-} mice were studied in a model hypertensive renal damage.

Experimental Procedures

Animal procedures

Hypertension-induced renal damage was obtained utilizing a previously published model involving uninephrectomy and Ang II administration (214), with the exception of using the 129S6 background instead of FVB/N. Twelve week old 129S6/SvEvTac (EP1^{+/+}, Taconic, USA) and EP1^{-/-} mice (85) underwent unilateral nephrectomy and implantation of a subcutaneous osmotic minipump delivering Ang II (1.4 mg/kg/day) for 6 weeks as illustrated in Figure 4.1. In some cases, uninephrectomy was performed in the absence of Ang II administration. Blood was collected at baseline and 6 weeks post-treatment for measurement of BUN and GFR. Every two weeks, systolic blood pressure was measured and urine collected. Animals were maintained in an AAALAC accredited rodent facility in individually ventilated microisolator cages on a 12:12 light dark cycle.

All procedures were done in accordance with the policies of the Institutional Animal Care and Use Committee at Vanderbilt University.

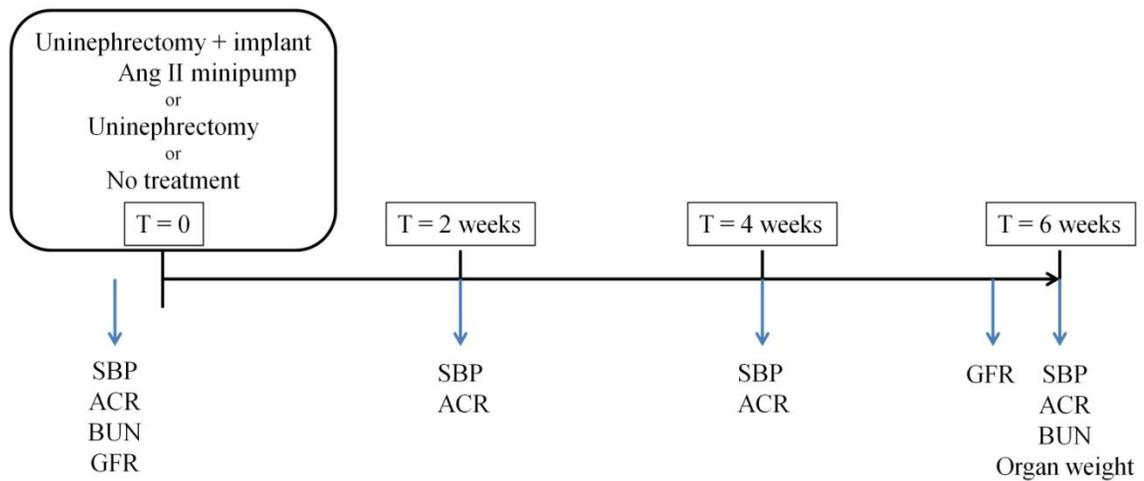


Figure 4.1 Experimental Design of Nphx + Ang II induced renal damage. EP1^{+/+} and EP1^{-/-} mice underwent uninephrectomy + implantation of an Ang II osmotic minipump delivering 1.4 mg/kg/day, uninephrectomy only, or no treatment and followed for 6 weeks.

Measurement of Systolic Blood Pressure

Systolic BP was determined in conscious mice using a computerized tail-cuff system (Visitech systems BP-2000 Blood Pressure Analysis System, Apex NC, USA) in the Mouse Metabolic Phenotyping Core at Vanderbilt University Medical Center. Mice were trained for four days minimizing physiologically apparent stress. Each measurement is the average of at least 10 consecutive readings after stabilization of blood pressure.

Determination of Blood Urea Nitrogen levels

To assess the renal function, blood urea nitrogen (BUN) was determined using an iSTAT-1 analyzer (Abbott Point of Care Inc., New Jersey, USA). Whole blood was obtained from saphenous vein and immediately assayed utilizing Chem8+ cartridges.

Determination of urinary albumin/creatinine ratios

Albumin/Creatinine ratios (ACR; expressed as mg albumin/mg creatinine) were measured from 20-200 μ L volumes of spot urine using Albuwell M ELISA kit, and urinary creatinine was measured using the Creatinine Companion (Exocell, Philadelphia, USA).

Glomerular filtration rate determination

GFR was determined based on plasma FITC–inulin clearance following a single bolus injection (223). Briefly, dialyzed FITC-inulin solution was injected into tail

vein. Approximately 20 μ L of blood was collected via saphenous venesection at 3, 7, 10, 15, 35, 55 and 75 minutes post injection for measurement of FITC concentration. Plasma samples were buffered to pH 7.4 with 500 mM HEPES and fluorescence was determined using 485-nm excitation and read at 538-nm emission. Parameters were estimated by two-phase exponential decay nonlinear regression of plasma fluorescence data (GraphPad Prism). GFR was calculated using the equation $GFR = I/(A/\alpha + B/\beta)$.

Statistical Analysis

Data are presented as means \pm SEM, using GraphPad Prism software (GraphPad Software Inc., USA). Analysis utilized Student's t test and two-way ANOVA. $P < 0.05$ was considered statistically significant for all studies.

Results

Hypertension was induced by uninephrectomy and Ang II

We employed a model of hypertension (214) to investigate the contribution of EP1 receptors to the pathogenesis of hypertension-induced renal damage. At baseline, SBP in EP1^{-/-} mice was comparable to that of EP1^{+/+} mice (EP1^{+/+}: 117.8 ± 2.485 mm Hg, EP1^{-/-}: 114.3 ± 3.853 mm Hg, Figure 4.2A). Following uninephrectomy and Ang II administration, SBP increased in both EP1^{+/+} and EP1^{-/-} mice, though no differences were detected between the two genotypes at any time point (Figure 4.2B).

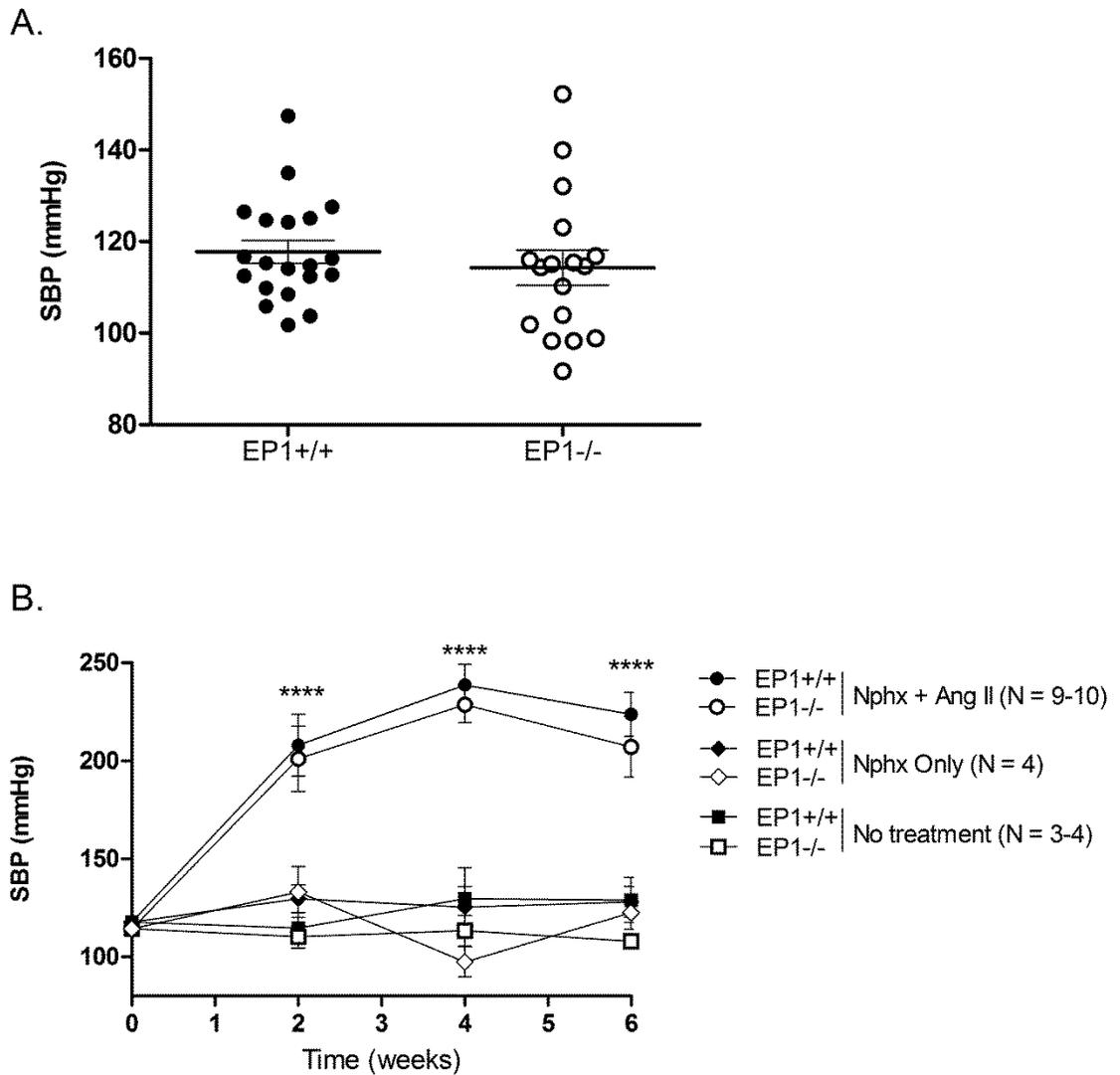


Figure 4.2 Hypertension was induced by Nphx + Ang II administration. A. Baseline systolic blood pressure was similar in EP1+/+ and EP1-/- mice ($P = 0.44$, $N = 17$ or 20 mice). B. Treatment with the Nphx + Ang II model induced comparable hypertension in both EP1+/+ (solid symbols) and EP1-/- (open symbols) mice. ($P < 0.0001$ vs baseline). No differences were detected between EP1+/+ and EP1-/- SBP.

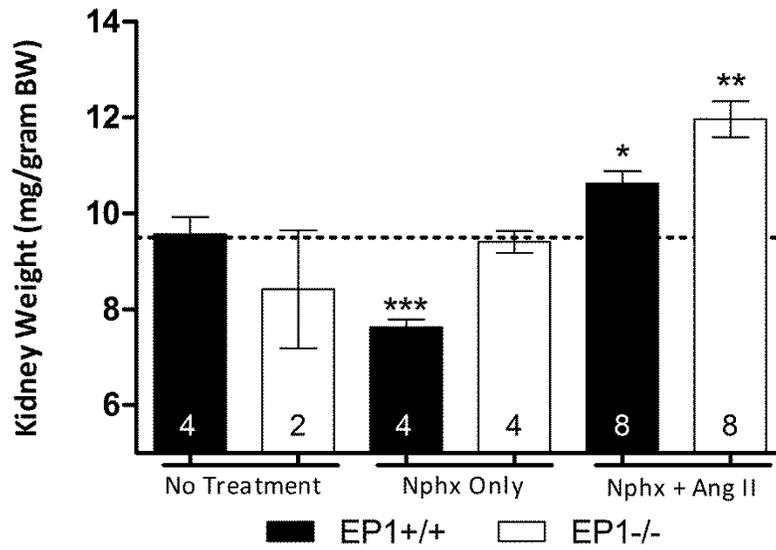
Organ weight was increased in EP1^{+/+} and EP1^{-/-} mice

Heart and kidney weight were increased in both EP1^{+/+} and EP1^{-/-} mice after 6 weeks treatment with Nphx + Ang II. Kidney weight was modestly increased by treatment with Nphx + Ang II in EP1^{+/+} and EP1^{-/-} mice compared to baseline (Figure 4.3A). Uninephrectomy alone decreased kidney weight in EP1^{+/+} mice, though no difference was observed in EP1^{-/-} mice. Heart weight was also significantly greater in Nphx + Ang II treated animals as compared to Nphx only, or untreated animals (Figure 4.3B). No significant differences in heart weight were observed between EP1^{+/+} and EP1^{-/-} mice, indicative of a similar degree of hypertension.

Renal damage was induced by Nphx + Ang II treatment and reduced in EP1^{-/-} mice

Blood urea nitrogen was significantly elevated in EP1^{+/+} and EP1^{-/-} mice with Nphx + Ang II (Figure 4.4A). Interestingly, EP1^{-/-} mice had modestly elevated BUN at baseline as compared to EP1^{+/+} mice. Over the 6 week treatment period, EP1^{+/+} mice experienced a rise in BUN, even in the absence of Nphx or Ang II, while EP1^{-/-} mouse BUN remained consistent unchanged. The difference between genotypes at baseline complicates the interpretation of the data. Therefore, change in BUN was determined using paired data from mice treated with Nphx + Ang II. Compared to baseline, six weeks post treatment EP1^{+/+} mice had a rise in BUN of 37 mg/dL, whereas EP1^{-/-} mouse BUN rose only 17 mg/dL (P = 0.007, Figure 4.4B). Urinary ACR was measured every other week throughout the experiment, and revealed significantly lower urinary protein

A.



B.

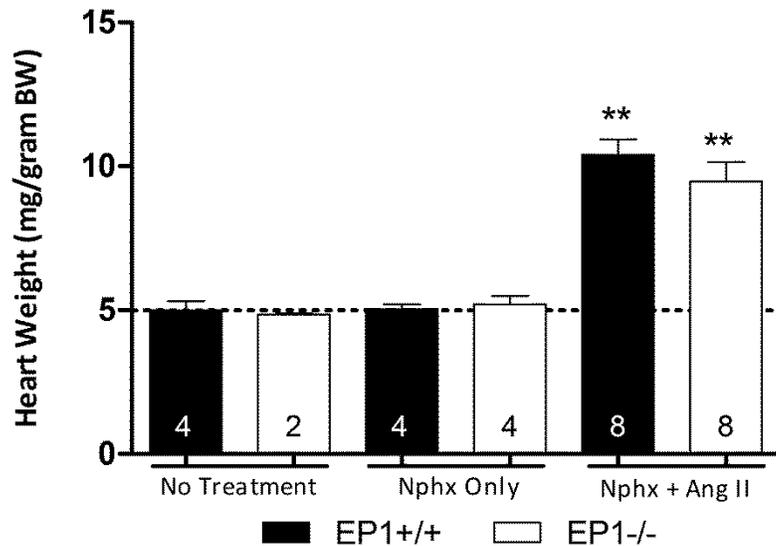


Figure 4.3 Heart and kidney weight were increased in both EP1+/+ and EP1-/- mice after 6 weeks treatment with Nphx + Ang II. A. Kidney weight, expressed mg/gram body weight, was increased in EP1+/+ and EP1-/- mice compared to baseline by treatment with Nphx + Ang II (EP1+/+ P = 0.04, EP1-/- P = 0.001). Nphx alone decreased kidney weight in EP1+/+ mice (P = 0.005), though no difference was observed in EP1-/- mice. B. Heart weight, expressed mg/gram body weight, was significantly greater in Nphx + Ang II treated animals as compared to Nphx only, or untreated animals (P < 0.01 for both genotypes). No significant differences were observed between EP1+/+ and EP1-/- mice (P > 0.05).

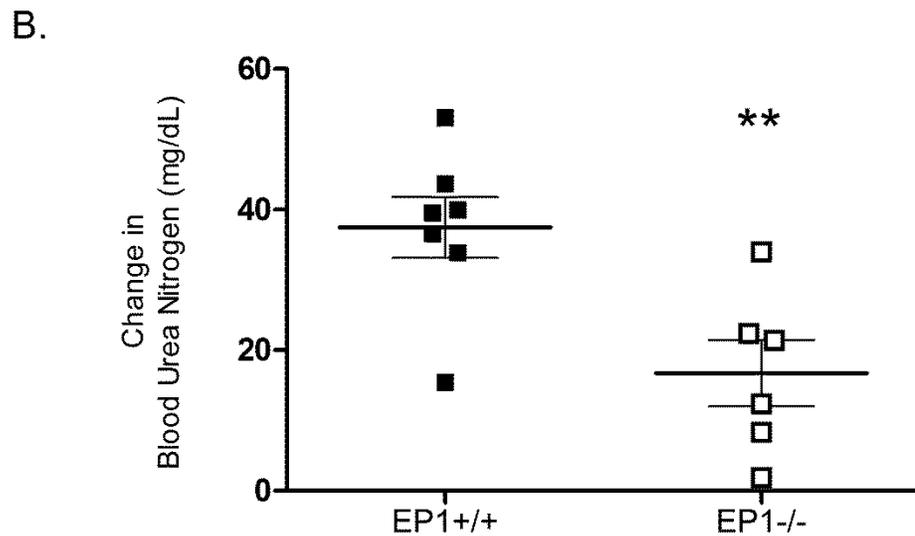
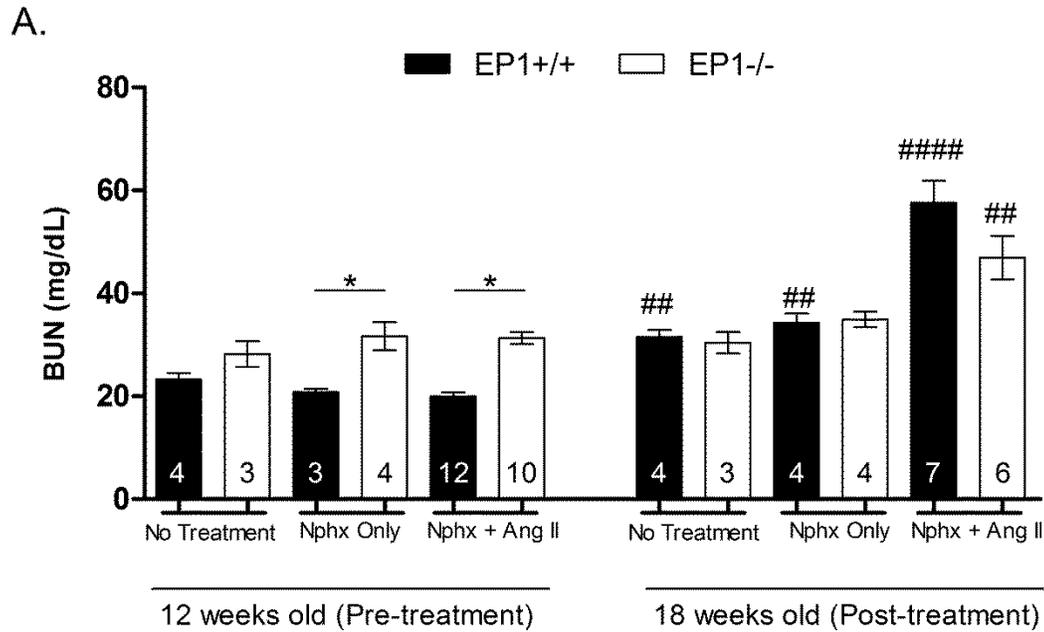


Figure 4.4 BUN was increased by Nphx + Ang II treatment. A. Blood urea nitrogen was measured prior to start of the model or six weeks after treatment. At baseline EP1-/- mice had elevated BUN compared to EP1+/+ mice (* $P < 0.05$). Six weeks later, BUN was elevated over baseline in all treatment groups of EP1+/+, and Nphx + Ang II treated EP1-/- mice (## $P < 0.004$, #### $P < 0.0001$). B. The change in BUN from baseline to six weeks treatment with Nphx + Ang II revealed EP1-/- mice had an attenuated rise in BUN compared to EP1+/+ mice (** $P = 0.007$)

in EP1^{-/-} mice compared to EP1^{+/+} mice ($P < 0.0001$, Figure 4.5A). GFR was decreased in EP1^{+/+} and EP^{-/-} mice treated with Nphx + Ang II, compared to untreated 18 week old mice, dropping 60 % in EP1^{+/+} mice and 40 % in EP1^{-/-} mice (Figure 4.5B). However, aging from 12 to 18 weeks of age resulted in a reduction in GFR in EP1^{-/-} mice only.

Discussion

In the present study we examined whether genetic ablation of the EP1 receptor affords renal protection in a model of hypertensive end-organ damage. The uninephrectomy and Ang II model resulted in functional renal damage including a rise in BUN, elevated ACR and reduced GFR, while genetic ablation of EP1 reduced the rise in BUN and decreased ACR. These studies suggest the EP1 receptor plays an important role in hypertensive renal disease.

Our results are consistent with Suganami et al., who showed that pharmacological blockade of EP1 reduced proteinuria and tubulointerstitial damage in stroke prone spontaneously hypertensive rats (87). However, in anti-GBM nephrotoxic serum nephritis, which occurs in the absence of elevated blood pressure, genetic deletion of EP1^{-/-} in mice resulted in increased mesangial expansion, tubular dilation, BUN and serum creatinine (173). Furthermore, no differences were observed in renal damage with genetic ablation of EP1 in the Nphx/DOCA-NaCl/Ang II model, although in this case only modest renal damage was induced and ablation of EP1 did substantially increase survival and protect

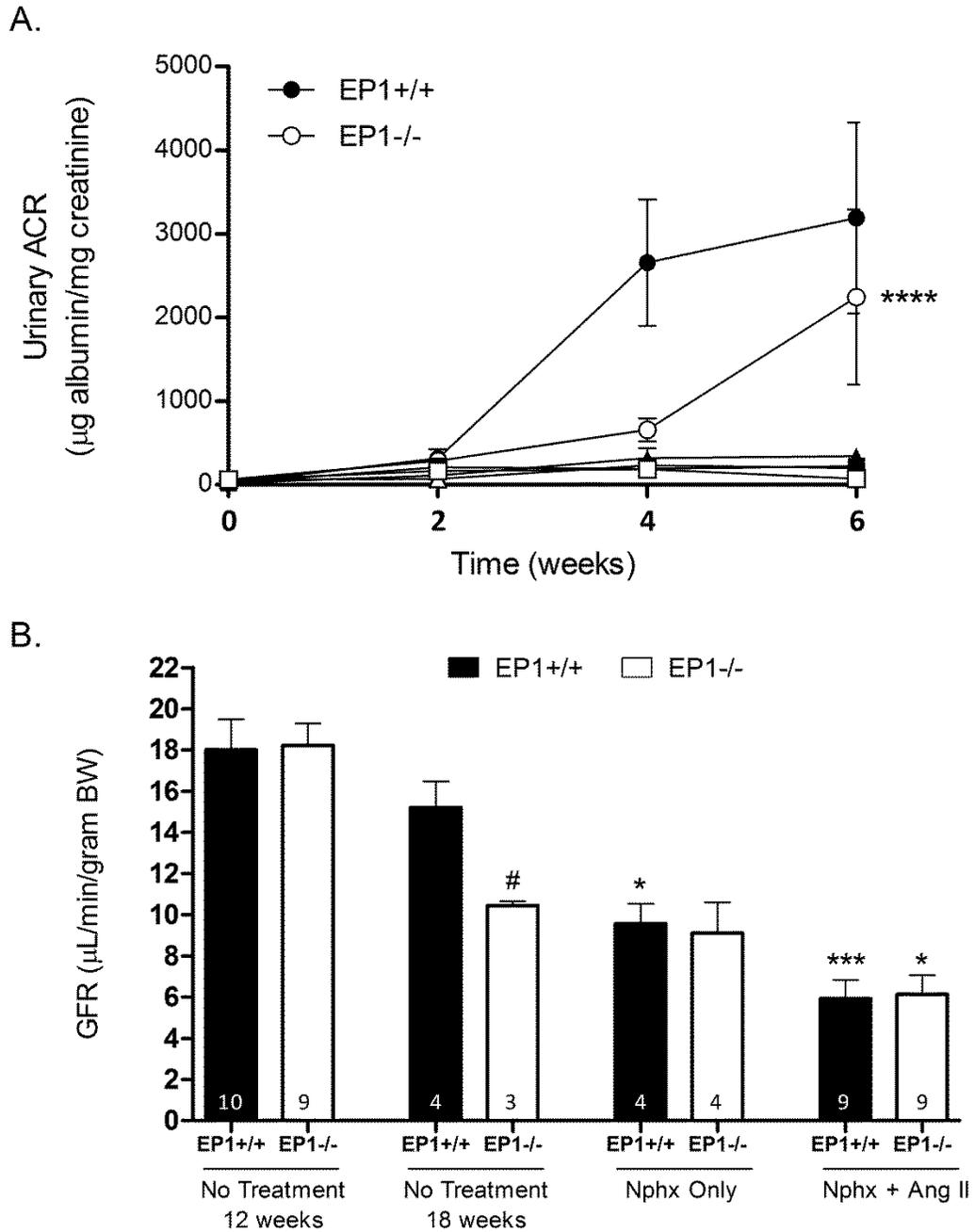


Figure 4.5 ACR and GFR following treatment with Nphx + Ang II. A. Urinary ACR was measured every other week throughout the experiment, revealing significantly lower urinary protein in EP1^{-/-} mice compared to EP1^{+/+} mice. (2 way ANOVA, ****P < 0.0001, N = 3-12 samples per point). B. GFR was decreased in EP1^{+/+} and EP1^{-/-} mice treated with Nphx + Ang II, compared to untreated 18 week old mice (EP1^{+/+} P = 0.001, EP1^{-/-} P = 0.028). Aging from 12 to 18 weeks of age resulted in a reduction in GFR in EP1^{-/-} mice only (#P = 0.002).

against vascular defects (224). This suggests the role of EP1 in renal damage is highly context dependent.

Throughout these studies blood pressure was measured by tail cuff plethysmography. Treatment with uninephrectomy and Ang II raised SBP as compared to uninephrectomy alone or no treatment. However, EP1^{-/-} mice had a similar degree of hypertension compared to EP1^{+/+} mice. This is inconsistent with previous reports demonstrating that pharmacological blockade and genetic ablation of EP1 reduces blood pressure, modestly at baseline and exaggerated in a setting of hypertension, and administration of EP1 agonists results in an increase in blood pressure (79,85,224). It is possible that blood pressure differences were not observed in our studies due to the lack of sensitivity of the tail cuff technique. To address this, more accurate blood pressure measurements would need to be made utilizing a direct arterial catheterization either anesthetized or by telemetry.

EP1^{-/-} mice had a reduction in the rise in BUN from baseline vs 6 weeks post uninephrectomy and Ang II, as compared to EP1^{+/+} mice. However, the raw values for BUN at 6 weeks post treatment failed to reach significance. Change in BUN reached significance because EP1^{-/-} began the study with significantly higher BUN as compared to EP1^{+/+} mice. Similarly GFR declined with age in untreated EP1^{-/-} mice and not EP1^{+/+} mice although the sample size was small in this case. This effect has not been observed previously, and it would be important to determine if this trend holds true in repeated experiments or is observed with prolonged treatment by pharmacological blockade in a

healthy animal. This could indicate that treatment with EP1 antagonists would be better utilized for slowing the progression of renal damage and not advised for use in patients without renal damage.

In summary, genetic ablation of the PGE₂ receptor EP1 affords protection against renal function decline as a result of hypertension. This protection may be independent of blood pressure reduction. These results suggest the EP1 receptor may be a viable target for the treatment of renal damage.

CHAPTER V

CONTRIBUTION OF THE EP1 RECEPTOR IN DIABETIC RENAL DAMAGE

Introduction

Diabetes is a disease characterized by high circulating blood glucose levels. This can occur as a result of inadequate insulin production, the inability to properly respond to insulin, or both. The prevalence of diabetes increases with age. In 2010, approximately 11.3 % of adults over 20 years of age had diabetes, while 26.9 % of adults over 65 years of age were diabetic. According to the CDC, in 2007 diabetes cost the US \$174 billion, \$116 billion of that composed of direct medical costs. Diabetes is the leading cause of blindness, lower-limb amputation, and kidney failure, and also increases the risk of coronary heart disease, peripheral vascular disease and stroke (225).

Diabetic nephropathy (DN) is a progressive disease which results in irreversible loss of kidney function. The initial mechanism of damage occurs due to adaptive hyperfiltration which eventually leads to long term damage of the nephrons. In type I diabetes, progression of DN begins within 5 years of onset of diabetes with glomerular hyperfiltration, which transitions into the presence of glomerular lesions without clinical disease or urinary albumin excretion (226-228). These lesions include thickening of the glomerular basement membrane and expansion of the mesangial cells. After about 10-15 years, incipient diabetic

nephropathy develops. The patient now has microalbuminuria along with the glomerular lesions. This is an important stage of DN because therapeutic intervention is started with the goal of preventing progression to overt DN. In overt DN there is worsening proteinuria, a drop in glomerular filtration rate and eventual progression to ESRD. The pathology of overt DN usually consists of glomerulosclerosis, fibrinoid caps and arteriolar hyaline sclerosis (226-228).

In 2008, 44 % of all new cases of kidney failure were a result of diabetes. It was estimated over 200,000 people with ESRD due to diabetes were on chronic dialysis or living with a transplant in 2008, even though the majority of diabetic patients with DN die of cardiovascular causes before progression to ESRD. Treatments for DN are aimed at controlling blood glucose and blood pressure (11,201,229). 75 % of adults with diabetes have blood pressure \geq 130/80 mm Hg. Treatment with ACEi or ARBs is more effective at reducing the decline in renal function than other blood pressure lowering drugs (217,218). According to the CDC, they reduce proteinuria, a risk factor for developing kidney disease, by 35 %. The AT1 receptor is the principal receptor mediating angiotensin II pressor effects and is the direct target of ARBs. AT1 receptor activation leads to a number of signal transduction pathways including increases in $[Ca^{++}]_i$ and activation of cPLA₂ (180,181,184-187). cPLA₂ is the rate limiting step in prostaglandin synthesis, freeing arachidonic acid from the plasma membrane. COX enzymes catalyze the oxidation and reduction of AA into PGH₂, which is rapidly converted into one of the principal prostanoids by tissue specific synthases. COX-2 expression has been shown to be upregulated in the thick

ascending limb and macula densa of both type I and type II diabetic rats (230-233). Increased COX-2 expression in the macula densa has also been seen in human diabetic kidneys (234). One product of COX-2 produced PGH₂ is the prostanoid PGE₂, which is of particular interest in diabetes.

PGE₂ has been shown to be elevated in the urine of diabetics, suggesting they have increased renal PGE₂ production (235). In STZ and Akita diabetes models, renal EP1 and EP3 mRNA expression is increased, suggesting these receptors may play an important role in the pathogenesis of DN (236).

In the Lepr^{db/db} model of diabetes, a homozygous mutation of the leptin receptor, treatment with antagonist AH6809 reduced SBP and blocked the vasoconstriction of PGE₂ and 17-phenyl trinor PGE₂, which was enhanced by the diabetic phenotype (86). Furthermore, pharmacologic blockade of EP1 was able to ameliorate DN in a rat model of STZ induced diabetes (88). Treatment with the EP1 antagonist was able to reduce renal and glomerular hypertrophy, reduce mesangial expansion, and suppress proteinuria (88). Therefore, we sought to determine the contribution of EP1 to DN utilizing a genetic disruption of EP1 in mice.

Experimental Procedures

Animal procedures

Diabetes-induced renal damage was obtained utilizing a previously published model involving low dose streptozotocin (STZ) treatment on an eNOS^{-/-}

background (195). Eight week old eNOS^{-/-} mice (EP1^{+/+}, eNOS^{-/-} Jackson Labs, USA), EP1^{-/-}, eNOS^{-/-} mice, EP1^{+/+}, eNOS^{+/+} mice and EP1^{-/-}, eNOS^{-/-} mice underwent five consecutive daily i.p. doses of STZ (50 mg/kg). Two weeks after STZ treatment blood glucose was measured by saphenous venesection. Diabetic mice with blood glucose greater than 300 mg/dL at 10 weeks of age were included in the studies. Blood glucose, blood pressure, urinary albumin excretion, and glomerular filtration rate were assessed 20 weeks after the onset of diabetes as illustrated in Figure 5.1. Animals were maintained in an AAALAC accredited rodent facility in individually ventilated microisolator cages on a 12:12 light dark cycle. All procedures were done in accordance with the policies of the Institutional Animal Care and Use Committee at Vanderbilt University.

Intracarotid blood pressure measurement

Intracarotid blood pressure was measured under ketamine (25 mg/kg) and inactin (100 mg/kg) anesthesia delivered intraperitoneally. Mice were placed on a thermal pad and a PE-10 catheter was inserted into the left carotid artery. The catheter was connected to a TXD-310 transducer and blood pressure was measured using a Digi-Med BPA 400 (Micromed). Mice were equilibrated 30-60 minutes until stable values were attained. Ten minute blood pressure measurements were collected and average mean arterial pressure (MAP) is plotted.

Determination of urinary albumin/creatinine ratios

Albumin/Creatinine ratios (ACR; expressed as mg albumin/mg creatinine) were measured from 20-200 μ L volumes of spot urine using Albuwell M ELISA kit, and urinary creatinine was measured using the Creatinine Companion (Exocell, Philadelphia, USA).

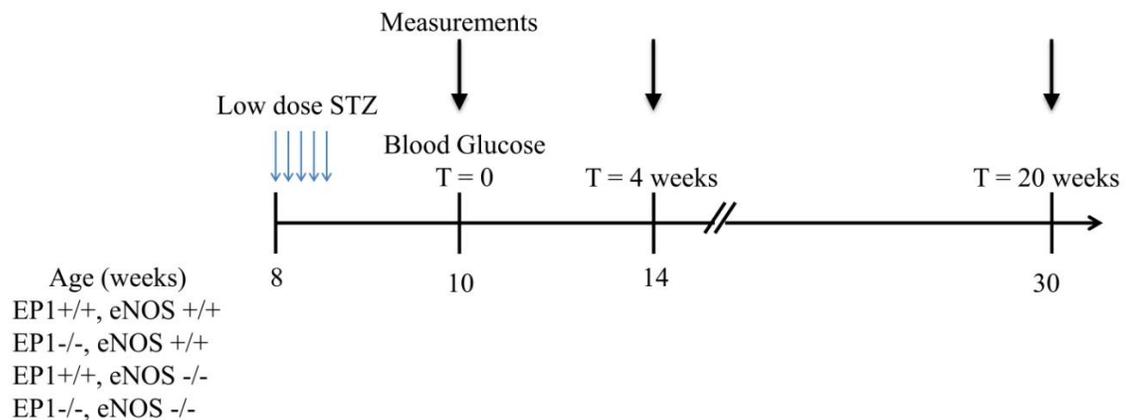


Figure 5.1 Induction of diabetes in mice. Eight week old EP1+/+, eNOS-/- mice, EP1-/- , eNOS-/- mice, EP1+/+, eNOS+/+ mice and EP1-/-, eNOS+/+ mice underwent five consecutive daily i.p.doses of STZ (50 mg/kg). Diabetic mice with blood glucose greater than 300 mg/dL at 10 weeks of age were included in the studies. Blood glucose, blood pressure, urinary albumin excretion, and glomerular filtration rate were assessed 20 weeks after the onset of diabetes.

Glomerular filtration rate determination

GFR was determined based on plasma FITC–inulin clearance following a single bolus injection (223). Briefly, dialyzed FITC-inulin solution was injected into tail vein. Approximately 20 μ L of blood was collected via saphenous venesection at 3, 7, 10, 15, 35, 55 and 75 minutes post injection for measurement of FITC concentration. Plasma samples were buffered to pH 7.4 with 500 mM HEPES and fluorescence was determined using 485-nm excitation and read at 538-nm emission. Parameters were estimated by two-phase exponential decay nonlinear regression of plasma fluorescence data (GraphPad Prism). GFR was calculated using the equation $GFR = I/(A/\alpha + B/\beta)$.

Statistical Analysis

Data are means \pm SEM, using GraphPad Prism software (GraphPad Software Inc., USA). Analysis utilized Student's t test. $P < 0.05$ was considered statistically significant for all studies.

Results

Diabetes was induced by low dose STZ treatment

Diabetic nephropathy was induced in EP1^{+/+}, eNOS^{-/-} mice, EP1^{-/-}, eNOS^{-/-} mice, EP1^{+/+}, eNOS^{+/+} mice and EP1^{-/-}, eNOS^{+/+} mice by five consecutive daily administrations of low dose STZ. Two weeks after STZ treatment, blood glucose was measured and diabetic mice with blood glucose greater than 300

mg/dL at 10 weeks of age were included in the studies. Four and 20 weeks after onset of diabetes, blood glucose was measured. In all genotypes blood glucose increased over time, and no differences were observed between the four genotypes (Figure 5.2).

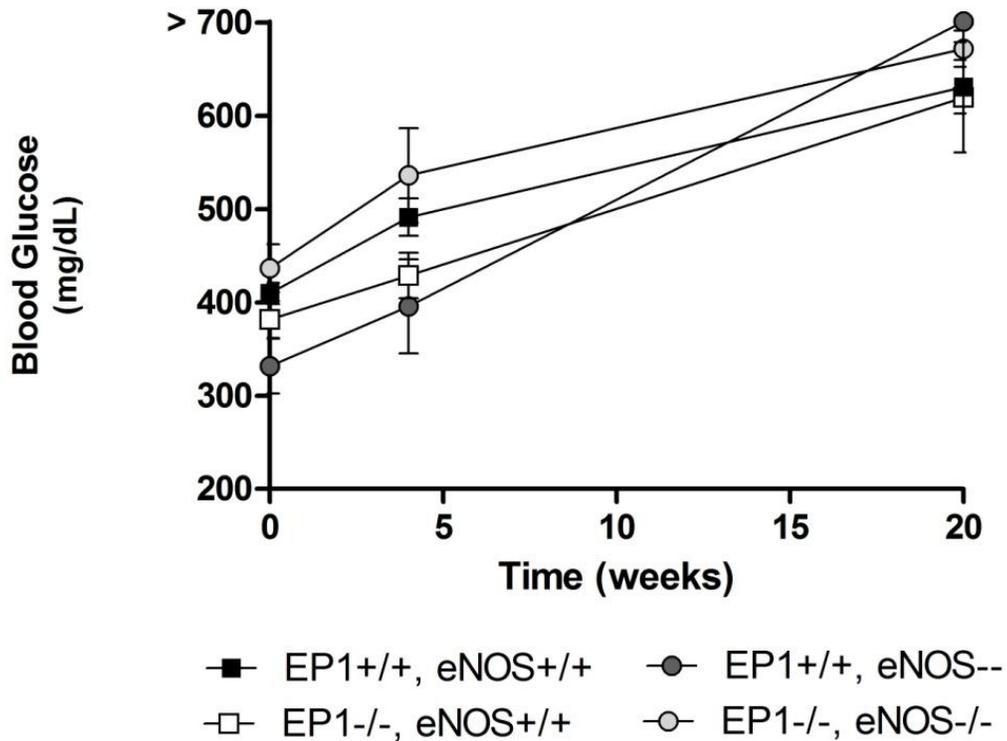


Figure 5.2 Diabetes was induced in all genotypes with low dose STZ treatment. Blood glucose was measured at onset of diabetes, four weeks diabetic and twenty weeks diabetic. Blood glucose increased with time in all groups, and no differences were observed between genotypes (EP1+/+, eNOS+/+ N = 5; EP1-/-, eNOS+/+ N = 7; EP1+/+, eNOS-/- N = 3; EP1-/-, eNOS-/- N = 10).

MAP was altered by genetic disruption of eNOS and not EP1

Anesthetized intracarotid blood pressure was measured at 20 weeks post onset of diabetes. No significant differences were observed between any of the genotypes (Figure 5.3A). Genetic deletion of eNOS has been well characterized to modestly increase MAP. When MAP was stratified based on eNOS genotype, regardless of the status of the EP1 gene, a significant increase in MAP was observed ($P = 0.027$, Figure 5.3B).

Urinary albumin excretion was reduced by genetic ablation of EP1

Low dose STZ has been demonstrated to induce diabetic nephropathy in C57BL/6 mice when eNOS has been deleted (195). In our study, we also observed increased protein excretion in EP1+/+, eNOS-/- mice as compared to EP1+/+, eNOS+/+ mice ($P = 0.009$). Deletion of the EP1 receptor on the background of eNOS-/- reduced urinary protein to a level similar to that observed in eNOS+/+ mice (EP1+/+, eNOS-/- vs. EP1-/-, eNOS-/- $P = 0.028$, Figure 5.4).

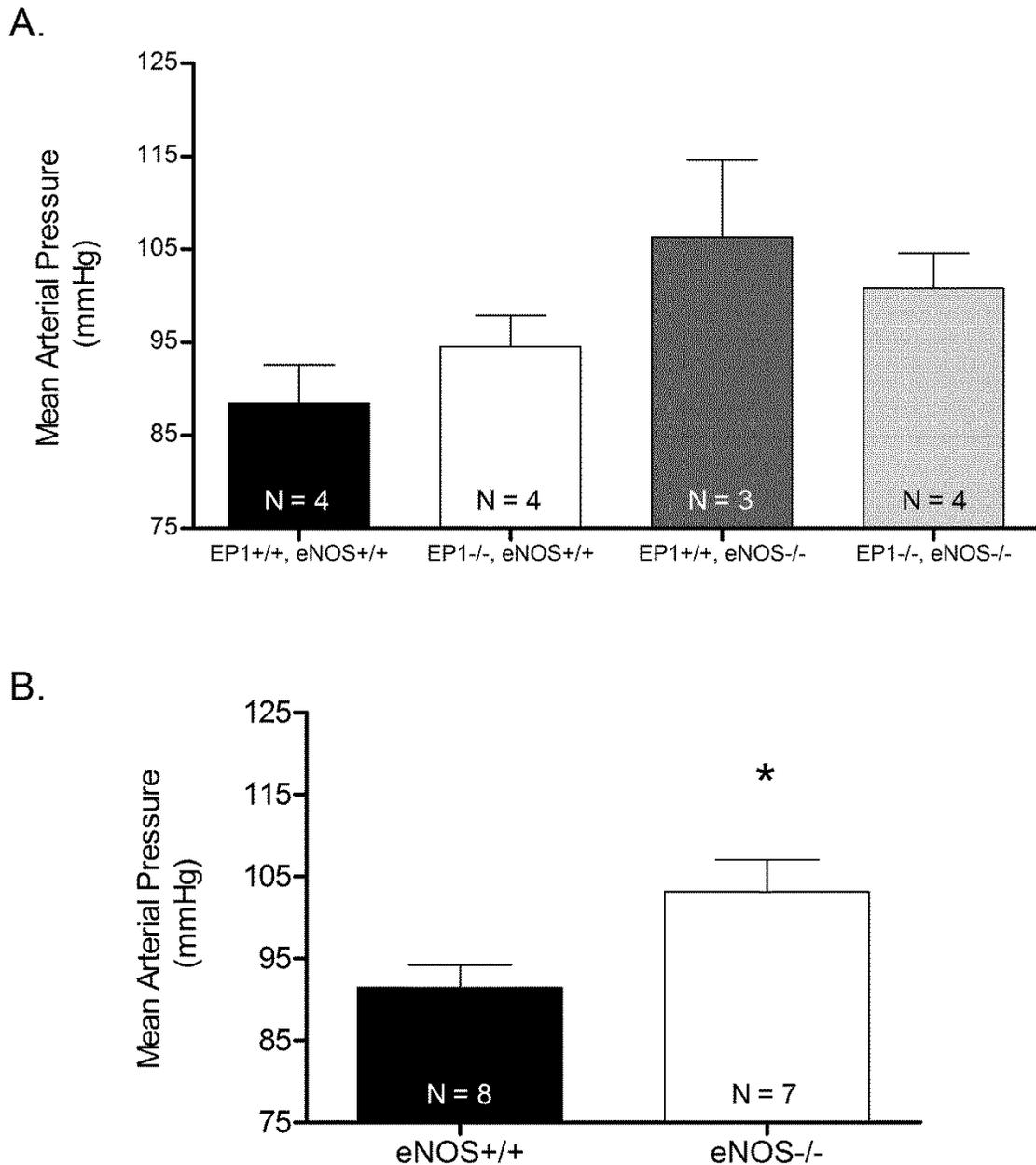


Figure 5.3 MAP was increased by genetic deletion of eNOS, but no change was observed due to EP1 gene disruption. A. Average MAP at 20 weeks diabetic. Anesthetized intracarotid blood pressure demonstrated no significant differences observed between any genotype (EP1^{+/+}, eNOS^{+/+} vs. EP1^{-/-}, eNOS^{+/+} P = 0.294; EP1^{+/+}, eNOS^{-/-} vs. EP1^{-/-}, eNOS^{-/-} P = 0.536; EP1^{+/+}, eNOS^{+/+} vs. EP1^{+/+}, eNOS^{-/-} P = 0.090; EP1^{-/-}, eNOS^{+/+} vs. EP1^{-/-}, eNOS^{-/-} P = 0.263 by t-test). B. MAP stratified based on eNOS genotype. MAP, regardless of EP1 gene status, was significantly increased with eNOS genetic ablation (P = 0.027).

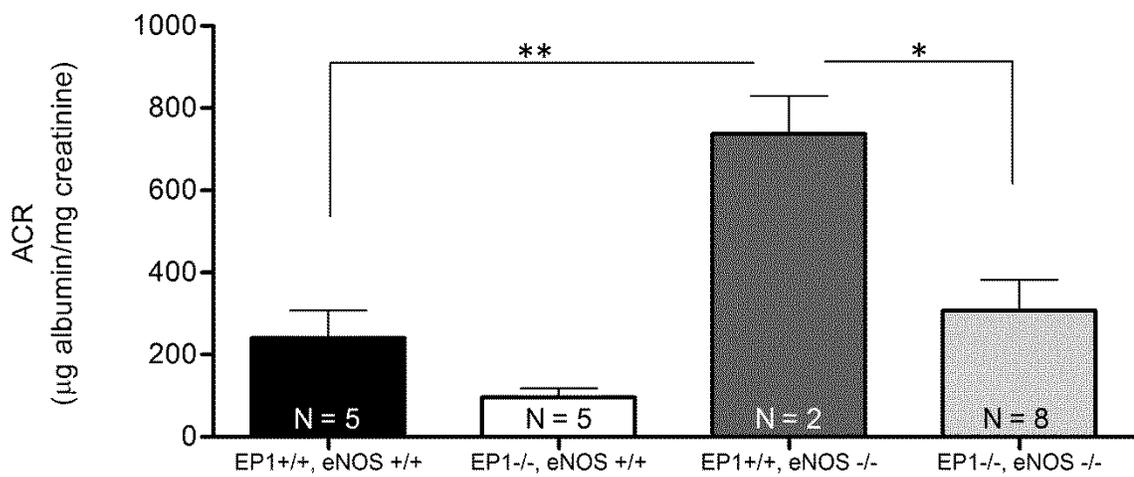


Figure 5.4 Albuminuria was significantly reduced by genetic disruption of EP1 in DN. Urinary albumin excretion was measured at 20 weeks post diabetes. Genetic deletion of eNOS significantly increased ACR on the background of EP1+/+ (P = 0.009), while no statistically significant increase was observed on the EP1-/- background (P = 0.054). Deletion of EP1 reduced ACR on the eNOS-/- background (P = 0.028).

GFR was reduced by deletion of eNOS in the presence, but not absence of EP1

Hyperfiltration is often observed in diabetes. In our study, we observed GFR of approximately 40 $\mu\text{L}/\text{min}/\text{gram}$ body weight in EP1^{+/+}, eNOS^{+/+} mice (Figure 5.5). Genetic deletion of eNOS significantly reduced GFR to approximately 15 $\mu\text{L}/\text{min}/\text{gram}$ ($P = 0.0001$), indicating compromised renal function. Genetic deletion of EP1 on the eNOS^{-/-} background had no statistically significant effect, but trended toward a slightly higher GFR (20 $\mu\text{L}/\text{min}/\text{gram}$, $P = 0.051$). However, genetic deletion of EP1 on the eNOS^{+/+} background significantly reduced GFR ($P = 0.011$). No significant reduction in GFR was observed between EP1^{-/-}, eNOS^{+/+} and EP1^{-/-}, eNOS^{-/-} mice ($P = 0.301$).

Discussion

In the present study we examined whether genetic disruption of EP1 protected eNOS^{-/-} mice from STZ induced diabetic nephropathy. We demonstrated EP1^{-/-}, eNOS^{-/-} mice have a significant reduction in proteinuria as compared to EP1^{+/+}, eNOS^{-/-} control mice. No significant difference in blood pressure was conferred by disruption of EP1 in this model. Our data are consistent a previous report that showed treatment of STZ induced diabetes in Wistar rats with EP1 antagonist ONO-8713 reduced renal hypertrophy and suppressed proteinuria (88), suggesting the EP1 receptor contributes to renal decline in the setting of diabetes.

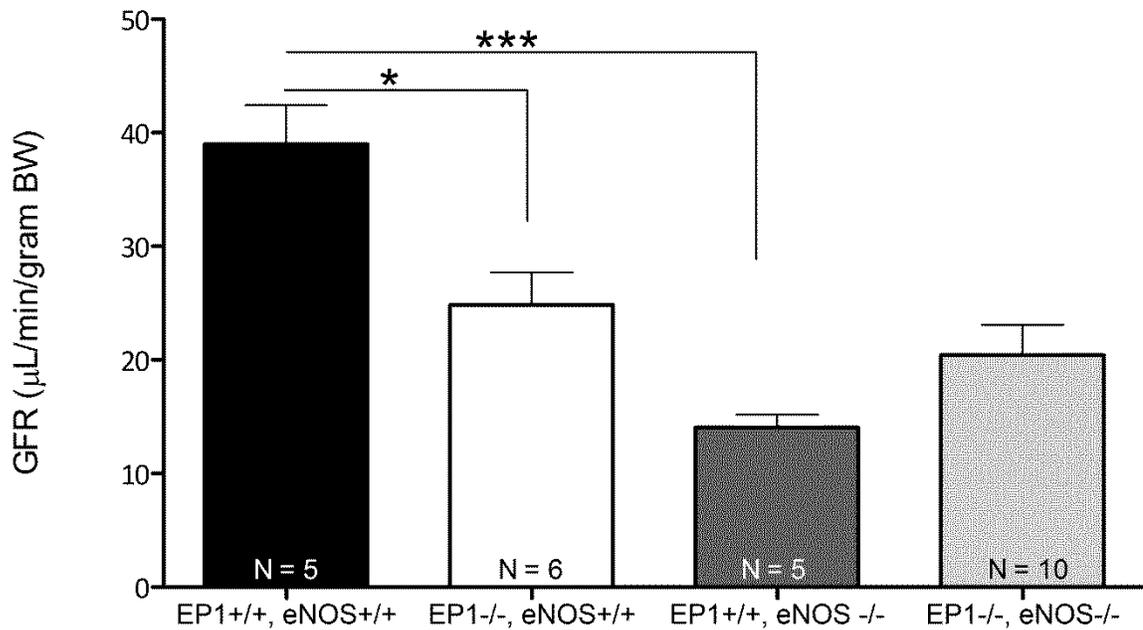


Figure 5.5 GFR was significantly reduced by deletion of eNOS in the presence, but not absence of EP1 receptor expression. GFR was measured at 20 weeks post diabetes. High GFR was observed in EP1+/+, eNOS+/+ mice, and was significantly reduced by deletion of eNOS (P = 0.0001) and deletion of EP1-/- (P = 0.011). Deletion of eNOS on the background of EP1-/- had no significant effect (P = 0.301). No statistically significant difference was observed between EP1+/+, eNOS-/- and EP1-/-, eNOS-/- mice (P = 0.051).

In our studies we were unable to detect a reduction in blood pressure due to disruption of EP1 suggesting suppression of proteinuria occurred by a mechanism other than blood pressure regulation. However, this may be a result of small sample size (3-4 mice per group). Deletion of eNOS conferred an average rise in BP of 10 mm Hg, and it may be expected that EP1 disruption reduce BP to an intermediate level. Such small changes would require larger sample sizes to measure with statistical significance. Rutkai et al demonstrated that BP and diabetes enhanced vasoconstriction could be reduced in the db/db diabetic model by treatment with AH6809 (86). The authors attribute the results to blockade of EP1 receptors although this particular antagonist is not very selective. In *Xenopus* oocytes expressing human EP1 receptors, AH6809 is able to inhibit calcium accumulation (21); in CHO cells stably expressing mouse prostanoid receptors AH6809 failed to compete $^3\text{H-PGE}_2$ for mouse EP1 (30 % displacement at 10 μM), while demonstrating a K_i of 350 nM at mouse EP2 (34). It would be interesting whether their results could be reproduced with a more selective antagonist such as ONO-8713.

Another interesting finding uncovered in this study was lower GFR observed in EP1^{-/-}, eNOS^{+/+} mice as compared to EP1^{+/+}, eNOS^{+/+}. C57BL/6J eNOS^{+/+} mice are resistant to development of diabetic nephropathy, while deletion of eNOS renders the strain susceptible (195,237). We predicted GFR would be elevated in both genotypes to a similar degree due to hyperfiltration. It may be possible that EP1^{-/-} mice do not hyperfilter to the same extent as EP1^{+/+} mice. Alternatively it is possible that EP1 plays an important role in an age

dependent reduction in GFR. At the time GFR was measured, the mice were > 6 months old. This hypothesis is consistent with results shown in Chapter IV, Figure 4.5B. EP1^{-/-} 129S6 mice, which were aged from 12 to 18 weeks without uninephrectomy or Ang II, displayed a significant reduction in GFR while no reduction was observed in EP1^{+/+} mice. It would be interesting to determine if this trend holds true in repeated experiments or is observed with prolonged treatment by pharmacological blockade in a healthy animal. Use of control animals, receiving no STZ, would be of importance in future experiment to eliminate hyperfiltration as a confounding variable.

In summary, our data demonstrate a detrimental role of EP1 in diabetes-induced proteinuria. This protection may be independent of blood pressure reduction. In addition, it would be of great interest to determine differences renal morphology as well. These results suggest the EP1 receptor may be a viable target for the treatment of diabetic renal damage.

CHAPTER VI

SUMMARY AND FUTURE DIRECTIONS

Summary

Hypertension is a prevalent disease affecting one in three adults in the United States. It is estimated that 27.5 % of the adult population is either not receiving therapy for their hypertension or is unable to control their blood pressure with the current therapies, making treatment of hypertension an important public health goal.

PGE₂, a biologically active lipid-derived autacoid, contributes to the regulation of blood pressure and is able to exert vasopressor or vasodepressor effects depending upon the setting (17-19). EP1 does not appear to play a significant role in the blood pressure effects of systemically administered PGE₂, however, it does contribute to hypertension. EP1^{-/-} mice have blunted pressor responses to both acute and chronic Ang II administration (85). In isolated vessel preparations, pre-treatment with the EP1/EP3 antagonist SC51322 reduced Ang II mediated vasoconstriction (85). Treatment of spontaneously hypertensive rats with SC51322 significantly reduces blood pressure (85), indicating blockade of the EP1/EP3 receptors may be a target for the treatment of hypertension.

EP1 blockade has been shown to positively affect renal function in stroke-prone spontaneously hypertensive rats (87), as well as cerebrovascular dysfunction induced by Ang II (161), implicating the EP1 receptor in hypertension and resultant end-organ damage. No data exists regarding the contribution of the EP3 receptor to hypertensive end-organ damage.

Three-quarters of diabetic patients have blood pressure \geq 130 mm Hg, and diabetes is the leading cause of chronic kidney failure. In STZ and Akita diabetes models, renal EP1 and EP3 mRNA expression is increased, suggesting these receptors may play an important role in the pathogenesis of DN (236). Pharmacologic blockade of EP1 was able to ameliorate DN in a rat model of STZ induced diabetes (88). Treatment with the EP1 antagonist was able to reduce renal and glomerular hypertrophy, reduce mesangial expansion, and suppress proteinuria (88). However, little advance has been made on the role of EP1 in hypertensive and diabetic renal damage, as these studies were published about 10 years ago, nor have the effects been reproduced with a genetic approach or another antagonist. Therefore, we sought to determine the contribution of EP1 and/or EP3 receptors to hypertensive end-organ damage and DN using a genetic approach in mice.

Using the Nphx/DOCA-NaCl/Ang II model of hypertension, we have demonstrated that disruption of EP1 or EP3 can afford substantial protection from end-organ damage and reduce incidence of mortality (Chapter II and Chapter III). The beneficial effects of EP1 disruption, and likely EP3 disruption, appeared to be a result of reduction in MAP, since treatment with the

antihypertensive agent hydralazine was able to phenocopy the effect observed in EP1^{-/-} mice. Mortality in the Nphx/DOCA-NaCl/Ang II model occurred primarily as a result of aortic aneurysm rupture, or after development of anasarca. We hypothesize that hypertension induced by DOCA-NaCl and Ang II results in volume loading and enhanced vasoconstriction, which places excessive stress on the vascular wall leading to enhanced permeability, resulting in edema and susceptibility to dissections and rupture. While we have shown that protection against end-organ damage is likely a result of reduced blood pressure, this does not eliminate the possibility that EP1 receptors might also provide protection directly at the target tissue, especially given the acute, severe nature of damage in this model.

Previous reports of the role of EP1 in renal injury are contradictory. In spontaneously hypertensive rats, treatment with an EP1 antagonist reduced proteinuria and tubulointerstitial damage (87), while in anti-GBM nephrotoxic serum nephritis genetic deletion of EP1^{-/-} in mice resulted in enhanced mesangial expansion and tubular dilation and increased blood urea nitrogen and serum creatinine (173). In our studies with the Nphx/DOCA-NaCl/Ang II model, modest hypertensive renal damage was observed, although no significant differences in renal function were detected between genotypes. However, our interpretation was confounded by the differential mortalities in EP1^{+/+} and EP1^{-/-} mice, potentially biasing our results. Examination of renal histopathology at time points prior to significant mortality failed to detect any severe renal damage or

differences between the genotypes, suggesting the role of EP1 in renal damage is highly context dependent.

Due to the high mortality and lack of severe renal damage observed in the Nphx/DOCA-NaCl/Ang II model, we pursued the use of another model involving uninephrectomy and Ang II on a 129S6 background (Chapter IV). This genetic background is more susceptible to development of renal damage, and it therefore requires less manipulation to achieve more renal damage. The uninephrectomy and Ang II model resulted in functional renal damage including a rise in BUN, elevated ACR and reduced GFR. Genetic ablation of EP1 reduced the rise in BUN and ACR while no change in hypertension was observed. This suggests the EP1 receptor plays an important role in hypertensive renal disease independent of blood pressure reduction. However, throughout these studies blood pressure was measured by tail cuff plethysmography. It is possible that blood pressure differences were not observed in our studies due to the lack of sensitivity of the tail cuff technique.

Lastly, we examined whether genetic disruption of EP1 protected eNOS^{-/-} mice from STZ induced diabetic nephropathy (Chapter V). We demonstrated EP1^{-/-}, eNOS^{-/-} mice have a significant reduction in proteinuria as compared to EP1^{+/+}, eNOS^{-/-} control mice. No significant difference in blood pressure was conferred by disruption of EP1 in this model. Our data are consistent a previous report that showed treatment of STZ induced diabetes in Wistar rats with EP1 antagonist ONO-8713 reduced renal hypertrophy and suppressed proteinuria (88), suggesting the EP1 receptor contributes to renal decline in the setting of

diabetes. Again, we were unable to detect a reduction in blood pressure due to disruption of EP1 suggesting suppression of proteinuria occurred by a mechanism other than blood pressure regulation. Although blood pressure was measured using a direct intracarotid catheter, the lack of a difference may be a result of small sample size (3-4 mice per group).

Another interesting finding uncovered in these studies points to a potential role of the EP1 receptor in age-dependent renal decline. In the diabetic renal model (Chapter V), lower GFR was observed in EP1^{-/-}, eNOS^{+/+} mice as compared to EP1^{+/+}, eNOS^{+/+}. At the time GFR was measured, the mice were > 6 months old. Furthermore, in the hypertensive renal model (Chapter IV), EP1^{-/-} mice began the study with significantly higher BUN as compared to EP1^{+/+} mice. Similarly GFR declined with age in untreated EP1^{-/-} mice and not EP1^{+/+} mice. This effect has not been observed previously, and it would be important to determine if this trend holds true in repeated experiments or is observed with prolonged treatment by pharmacological blockade in a healthy animal. This could indicate that treatment with EP1 antagonists would be better utilized for slowing the progression of renal damage and not advised for use in patients without renal damage.

In summary, our data demonstrate a detrimental role of EP1 in hypertensive and diabetic end-organ damage. Further characterization of EP1 in these diseases will be essential as the EP1 receptor may be a viable pharmaceutical target for the treatment of hypertension and subsequent organ damage.

Future directions

The data presented in this thesis advances our knowledge of the role of EP1 and EP3 receptors in hypertension and subsequent sequelae. Several new questions have emerged as a result including: Does the EP1 receptor mediate the BP- independent actions of Ang II? What is the therapeutic relevance of EP1 blockade? What is the contribution of EP1 in vascular permeability? Which EP receptors contribute to aortic aneurysm formation? What is the mechanism of EP1/Angiotensin II hypertension? Future experiments should be designed to address these issues.

Drugs which block the renin-angiotensin-aldosterone pathway are considered superior to other anti-hypertensive treatments due to their beneficial actions directly on the kidney which occur independent of blood pressure reduction. Additionally, PGE₂ and the EP1 receptor have been demonstrated to mediate at least part of the actions of angiotensin II. It would be of great interest to determine whether genetic disruption of EP1 also confers similar protection in the kidney. Preliminary studies presented in this thesis (Chapters IV and V) suggest this may be the case. Disruption of EP receptors suppressed increases in proteinuria and BUN or prevented reductions in GFR, while no significant differences in blood pressure were detected. However, throughout these studies blood pressure was measured by tail cuff plethysmography or contained small sample sizes of direct intracarotid measurements. It is possible that blood pressure differences were not observed in our studies due to the lack of

sensitivity or low power. To address this, future experiments should include more accurate blood pressure measurements using a direct arterial catheterization, in either anesthetized mice or by telemetry. Additionally, the therapeutic relevance of EP1 blockade should be more thoroughly characterized. One major limitation associated with the use of genetic disruption versus pharmacologic blockade is the chronic disruption of receptor action. Lack of receptor action throughout development or in the absence of disease may result in compensatory changes which may not be observed with use of pharmacologic agents. Therefore, determining whether similar results can be obtained by treatment with an EP1 antagonist would be essential for verifying the therapeutic benefit of EP1 blockade. Furthermore, it would be of interest to determine whether there is additional benefit from dual EP1/EP3 blockade compared with EP1 or EP3 blockade only. The mechanism by which EP1 and EP3 mediate hypertension is unknown, however it is well understood that the EP1 and EP3 can mediate distinct signal transduction pathways. It is conceivable that blockade of EP1 and EP3 may have an additive effect and provide enhanced protection.

For the uninephrectomy/ DOCA-NaCl/Ang II model, we hypothesized that hypertension resulted in enhanced vascular permeability, causing edema and susceptibility to dissections and rupture (Chapter II). This hypothesis was based on eliminating the most common causes of edema. Furthermore both aneurysms and edema were reduced via treatment with hydralazine, supporting hypertension as the major mitigating factor for development of these pathologies.

Future experiments will be required to identify whether vascular permeability differences are observed between EP1^{+/+} and EP1^{-/-} mice. In this model, vascular permeability could be examined using Evans Blue dye or dextran-rhodamine permeability. If differences were observed in vivo, the molecular mechanism could be identified using transwell vascular permeability assays, with cells isolated from EP1^{+/+} and EP1^{-/-} mice or heterologously expressing EP1.

It has been previously shown that either selective inhibition of COX-2 or genetic deletion of COX-2 significantly reduced aortic aneurysm formation and macrophage infiltration (166,167). Furthermore, deletion of microsomal PGE synthase-1 has also been demonstrated to reduce aortic aneurysm formation and oxidative stress in LDLR^{-/-} mice with an angiotensin II infusion (160), suggesting PGE₂ plays an important role in development of aneurysms and the EP receptors may be viable targets for treatment of aneurysm progression. It would be of interest to investigate the contribution of EP1 in vascular damage, utilizing models which are independent of increased blood pressure, such as wire injury.

Lastly, it is imperative that we identify the mechanism by which EP1 and EP3 mediate Ang II-induced hypertension. It has been demonstrated that Ang II-induced vasoconstriction can be blocked by treatment with EP1/EP3 antagonist SC-51322. EP1 receptors in the sub-fornical organ of the brain also play a critical role in slow pressor Ang II hypertension (238). Ang II/EP1 signaling in the subfornical organ is dependent of COX activity, suggesting that Ang II results in local production of PGE₂, which then activates EP1 (238). It would be interesting

to determine whether a similar mechanism/interaction is observed in other target tissues such as the kidney. Furthermore, the mechanism of EP3 action in Ang II-mediated hypertension is still unknown but may have an important contribution in Ang II-induced vasoconstriction. Future experiment designed to address these remaining questions would increase our overall understanding of EP receptors in blood pressure regulation, and may also uncover new therapeutic targets for the treatment of hypertension and end-organ damage.

BIBLIOGRAPHY

1. Moser, M. (2006) Historical perspectives on the management of hypertension. *J Clin Hypertens (Greenwich)* **8**, 15-20
2. FM, A. (1922) Treatment of arterial hypertension. *Journal of Metabolic Research* **2**, 429-545
3. W, K. (1944) Treatment of kidney disease and hypertensive vascular disease with rice diet. *North Carolina Medical Journal* **5**, 125-127
4. Moser, M. (1997) Evolution of the treatment of hypertension from the 1940s to JNC V. *Am J Hypertens* **10**, 2S-8S
5. Perera, G. A. (1955) Hypertensive vascular disease; description and natural history. *J Chronic Dis* **1**, 33-42
6. Moser, M., and Macaulay, A. I. (1959) Chlorothiazide as an adjunct in the treatment of essential hypertension. *Am J Cardiol* **3**, 214-219
7. Kannel, W. B., Schwartz, M. J., and McNamara, P. M. (1969) Blood pressure and risk of coronary heart disease: the Framingham study. *Dis Chest* **56**, 43-52
8. (1967) Veterans Administration Cooperative Study Group on Anti-hypertensive Agents. Effects of treatment on morbidity in hypertension. Results in patients with diastolic blood pressures averaging 115 through 129 mm Hg. *JAMA* **202**, 1028-1034
9. (1977) Report of the Joint National Committee on Detection, Evaluation, and Treatment of High Blood Pressure. A cooperative study. *JAMA* **237**, 255-261
10. (2012) *National Center for Health Statistics. Health, United States, 2011: With Special Feature on Socioeconomic Status and Health.*, Hyattsville, MD.
11. Chobanian, A. V., Bakris, G. L., Black, H. R., Cushman, W. C., Green, L. A., Izzo, J. L., Jones, D. W., Materson, B. J., Oparil, S., Wright, J. T., Rocella, E. J., National Heart, L., and Blood Institute Joint National Committee on Prevention, Detection, Evaluation, and Treatment of High Blood Pressure, and Committee, N. H. B. P. E. P. C. (2003) The Seventh Report of the Joint National Committee on Prevention, Detection, Evaluation, and Treatment of High Blood Pressure: the JNC 7 report. *JAMA* **289**, 2560-2572

12. Foex, P. (2004) Hypertension: pathophysiology and treatment. *Contin Educ Anaesth Crit Care Pain* **4**, 71-75
13. Sever, P. S., and Poulter, N. R. (1989) A hypothesis for the pathogenesis of essential hypertension: the initiating factors. *J Hypertens Suppl* **7**, S9-12
14. (1988) Intersalt: an international study of electrolyte excretion and blood pressure. Results for 24 hour urinary sodium and potassium excretion. Intersalt Cooperative Research Group. *BMJ* **297**, 319-328
15. Brunton, L. L., Chabner, B. A., and Knollmann, B. C. (2010) *Goodman & Gilman's The Pharmacological Basis of Therapeutics, 12e*, McGraw-Hill Professional
16. Tai, H. H., Ensor, C. M., Tong, M., Zhou, H., and Yan, F. (2002) Prostaglandin catabolizing enzymes. *Prostaglandins Other Lipid Mediat* **68-69**, 483-493
17. Kennedy, C. R., Zhang, Y., Brandon, S., Guan, Y., Coffee, K., Funk, C. D., Magnuson, M. A., Oates, J. A., Breyer, M. D., and Breyer, R. M. (1999) Salt-sensitive hypertension and reduced fertility in mice lacking the prostaglandin EP2 receptor. *Nat Med* **5**, 217-220
18. Schaaf, T. K., and Hess, H. J. (1979) Synthesis and biological activity of carboxyl-terminus modified prostaglandin analogues. *J Med Chem* **22**, 1340-1346
19. Hockel, G. M., and Cowley, A. W., Jr. (1979) Prostaglandin E2-induced hypertension in conscious dogs. *Am J Physiol* **237**, H449-454
20. Sugimoto, Y., Namba, T., Honda, A., Hayashi, Y., Negishi, M., Ichikawa, A., and Narumiya, S. (1992) Cloning and expression of a cDNA for mouse prostaglandin E receptor EP3 subtype. *J Biol Chem* **267**, 6463-6466
21. Funk, C. D., Furci, L., FitzGerald, G. A., Grygorczyk, R., Rochette, C., Bayne, M. A., Abramovitz, M., Adam, M., and Metters, K. M. (1993) Cloning and expression of a cDNA for the human prostaglandin E receptor EP1 subtype. *J Biol Chem* **268**, 26767-26772
22. Honda, A., Sugimoto, Y., Namba, T., Watabe, A., Irie, A., Negishi, M., Narumiya, S., and Ichikawa, A. (1993) Cloning and expression of a cDNA for mouse prostaglandin E receptor EP2 subtype. *J Biol Chem* **268**, 7759-7762

23. Regan, J. W., Bailey, T. J., Pepperl, D. J., Pierce, K. L., Bogardus, A. M., Donello, J. E., Fairbairn, C. E., Kedzie, K. M., Woodward, D. F., and Gil, D. W. (1994) Cloning of a novel human prostaglandin receptor with characteristics of the pharmacologically defined EP2 subtype. *Mol Pharmacol* **46**, 213-220
24. Narumiya, S., Sugimoto, Y., and Ushikubi, F. (1999) Prostanoid Receptors: Structures, Properties, and Functions. *Physiol Rev* **79**, 1193-1226
25. Sugimoto, Y., Negishi, M., Hayashi, Y., Namba, T., Honda, A., Watabe, A., Hirata, M., Narumiya, S., and Ichikawa, A. (1993) Two isoforms of the EP3 receptor with different carboxyl-terminal domains. Identical ligand binding properties and different coupling properties with Gi proteins. *J Biol Chem* **268**, 2712-2718
26. Irie, A., Sugimoto, Y., Namba, T., Harazono, A., Honda, A., Watabe, A., Negishi, M., Narumiya, S., and Ichikawa, A. (1993) Third isoform of the prostaglandin-E-receptor EP3 subtype with different C-terminal tail coupling to both stimulation and inhibition of adenylate cyclase. *Eur J Biochem* **217**, 313-318
27. Okuda-Ashitaka, E., Sakamoto, K., Ezashi, T., Miwa, K., Ito, S., and Hayaishi, O. (1996) Suppression of prostaglandin E receptor signaling by the variant form of EP1 subtype. *J Biol Chem* **271**, 31255-31261
28. Abramovitz, M., Adam, M., Boie, Y., Carriere, M., Denis, D., Godbout, C., Lamontagne, S., Rochette, C., Sawyer, N., Tremblay, N. M., Belley, M., Gallant, M., Dufresne, C., Gareau, Y., Ruel, R., Juteau, H., Labelle, M., Ouimet, N., and Metters, K. M. (2000) The utilization of recombinant prostanoid receptors to determine the affinities and selectivities of prostaglandins and related analogs. *Biochim Biophys Acta* **1483**, 285-293
29. Sugimoto, Y., and Narumiya, S. (2007) Prostaglandin E receptors. *J Biol Chem* **282**, 11613-11617
30. Watabe, A., Sugimoto, Y., Honda, A., Irie, A., Namba, T., Negishi, M., Ito, S., Narumiya, S., and Ichikawa, A. (1993) Cloning and expression of cDNA for a mouse EP1 subtype of prostaglandin E receptor. *J Biol Chem* **268**, 20175-20178
31. Katoh, H., Watabe, A., Sugimoto, Y., Ichikawa, A., and Negishi, M. (1995) Characterization of the signal transduction of prostaglandin E receptor EP1 subtype in cDNA-transfected Chinese hamster ovary cells. *Biochim Biophys Acta* **1244**, 41-48

32. Ji, R., Chou, C. L., Xu, W., Chen, X. B., Woodward, D. F., and Regan, J. W. (2010) EP1 prostanoid receptor coupling to G i/o up-regulates the expression of hypoxia-inducible factor-1 alpha through activation of a phosphoinositide-3 kinase signaling pathway. *Mol Pharmacol* **77**, 1025-1036
33. Regard, J. B., Sato, I. T., and Coughlin, S. R. (2008) Anatomical profiling of G protein-coupled receptor expression. *Cell* **135**, 561-571
34. Kiriya, M., Ushikubi, F., Kobayashi, T., Hirata, M., Sugimoto, Y., and Narumiya, S. (1997) Ligand binding specificities of the eight types and subtypes of the mouse prostanoid receptors expressed in Chinese hamster ovary cells. *Br J Pharmacol* **122**, 217-224
35. Lawrence, R. A., Jones, R. L., and Wilson, N. H. (1992) Characterization of receptors involved in the direct and indirect actions of prostaglandins E and I on the guinea-pig ileum. *Br J Pharmacol* **105**, 271-278
36. Dong, Y. J., Jones, R. L., and Wilson, N. H. (1986) Prostaglandin E receptor subtypes in smooth muscle: agonist activities of stable prostacyclin analogues. *Br J Pharmacol* **87**, 97-107
37. Suzawa, T., Miyaura, C., Inada, M., Maruyama, T., Sugimoto, Y., Ushikubi, F., Ichikawa, A., Narumiya, S., and Suda, T. (2000) The role of prostaglandin E receptor subtypes (EP1, EP2, EP3, and EP4) in bone resorption: an analysis using specific agonists for the respective EPs. *Endocrinology* **141**, 1554-1559
38. Watanabe, K., Kawamori, T., Nakatsugi, S., Ohta, T., Ohuchida, S., Yamamoto, H., Maruyama, T., Kondo, K., Narumiya, S., Sugimura, T., and Wakabayashi, K. (2000) Inhibitory effect of a prostaglandin E receptor subtype EP(1) selective antagonist, ONO-8713, on development of azoxymethane-induced aberrant crypt foci in mice. *Cancer Lett* **156**, 57-61
39. Watanabe, K., Kawamori, T., Nakatsugi, S., Ohta, T., Ohuchida, S., Yamamoto, H., Maruyama, T., Kondo, K., Ushikubi, F., Narumiya, S., Sugimura, T., and Wakabayashi, K. (1999) Role of the prostaglandin E receptor subtype EP1 in colon carcinogenesis. *Cancer Res* **59**, 5093-5096
40. Gardiner, P. J. (1986) Characterization of prostanoid relaxant/inhibitory receptors (psi) using a highly selective agonist, TR4979. *Br J Pharmacol* **87**, 45-56
41. Schwaner, I., Offermanns, S., Spicher, K., Seifert, R., and Schultz, G. (1995) Differential activation of Gi and Gs proteins by E- and I-type

prostaglandins in membranes from the human erythroleukaemia cell line, HEL. *Biochim Biophys Acta* **1265**, 8-14.

42. Sugimoto, Y., Nakato, T., Kita, A., Hatae, N., Tabata, H., Tanaka, S., and Ichikawa, A. (2003) Functional domains essential for Gs activity in prostaglandin EP2 and EP3 receptors. *Life Sci* **74**, 135-141
43. Fujino, H., West, K. A., and Regan, J. W. (2002) Phosphorylation of glycogen synthase kinase-3 and stimulation of T-cell factor signaling following activation of EP2 and EP4 prostanoid receptors by prostaglandin E2. *J Biol Chem* **277**, 2614-2619
44. Chun, K. S., Lao, H. C., Trempus, C. S., Okada, M., and Langenbach, R. (2009) The prostaglandin receptor EP2 activates multiple signaling pathways and beta-arrestin1 complex formation during mouse skin papilloma development. *Carcinogenesis* **30**, 1620-1627
45. Kim, J. I., Lakshmikanthan, V., Frilot, N., and Daaka, Y. (2010) Prostaglandin E2 promotes lung cancer cell migration via EP4-betaArrestin1-c-Src signalsome. *Mol Cancer Res* **8**, 569-577
46. Chun, K. S., Lao, H. C., and Langenbach, R. (2010) The prostaglandin E2 receptor, EP2, stimulates keratinocyte proliferation in mouse skin by G protein-dependent and {beta}-arrestin1-dependent signaling pathways. *J Biol Chem* **285**, 39672-39681
47. Breyer, M. D., and Breyer, R. M. (2001) G Protein-Coupled Prostanoid Receptors and the Kidney. *Annu Rev Physiol* **63**, 579-605
48. Sharif, N. A., and Davis, T. L. (2002) Cloned human EP1 prostanoid receptor pharmacology characterized using radioligand binding techniques. *J Pharm Pharmacol* **54**, 539-547
49. af Forselles, K. J., Root, J., Clarke, T., Davey, D., Aughton, K., Dack, K., and Pullen, N. (2011) In vitro and in vivo characterization of PF-04418948, a novel, potent and selective prostaglandin EP₂ receptor antagonist. *Br J Pharmacol* **164**, 1847-1856
50. Birrell, M. A., and Nials, A. T. (2011) At last, a truly selective EP₂ receptor antagonist. *Br J Pharmacol* **164**, 1845-1846
51. Birrell, M. A., Maher, S., Buckley, J., Dale, N., Bonvini, S., Raemdonck, K., Pullen, N., Giembycz, M. A., and Belvisi, M. G. (2012) Selectivity profiling of the novel EP(2) -receptor antagonist, PF-04418948, in functional bioassay systems: atypical affinity at the guinea pig EP(2) -receptor. *Br J Pharmacol*

52. An, S., Yang, J., So, S. W., Zeng, L., and Goetzl, E. J. (1994) Isoforms of the EP3 subtype of human prostaglandin E2 receptor transduce both intracellular calcium and cAMP signals. *Biochemistry* **33**, 14496-14502
53. Asboth, G., Phaneuf, S., Europe-Finner, G. N., Toth, M., and Bernal, A. L. (1996) Prostaglandin E2 activates phospholipase C and elevates intracellular calcium in cultured myometrial cells: involvement of EP1 and EP3 receptor subtypes. *Endocrinology* **137**, 2572-2579
54. Macias-Perez, I. M., Zent, R., Carmosino, M., Breyer, M. D., Breyer, R. M., and Pozzi, A. (2008) Mouse EP3 alpha, beta, and gamma receptor variants reduce tumor cell proliferation and tumorigenesis in vivo. *J Biol Chem* **283**, 12538-12545
55. Regan, J. W., Bailey, T. J., Donello, J. E., Pierce, K. L., Pepperl, D. J., Zhang, D., Kedzie, K. M., Fairbairn, C. E., Bogardus, A. M., Woodward, D. F., and et al. (1994) Molecular cloning and expression of human EP3 receptors: evidence of three variants with differing carboxyl termini. *Br J Pharmacol* **112**, 377-385
56. Schmid, A., Thierauch, K. H., Schleuning, W. D., and Dinter, H. (1995) Splice variants of the human EP3 receptor for prostaglandin E2. *Eur J Biochem* **228**, 23-30
57. Oldfield, S., Grubb, B. D., and Donaldson, L. F. (2001) Identification of a prostaglandin E2 receptor splice variant and its expression in rat tissues. *Prostaglandins Other Lipid Mediat* **63**, 165-173
58. Namba, T., Sugimoto, Y., Negishi, M., Irie, A., Ushikubi, F., Kakizuka, A., Ito, S., Ichikawa, A., and Narumiya, S. (1993) Alternative splicing of C-terminal tail of prostaglandin E receptor subtype EP3 determines G-protein specificity. *Nature* **365**, 166-170
59. Hasegawa, H., Negishi, M., and Ichikawa, A. (1996) Two isoforms of the prostaglandin E receptor EP3 subtype different in agonist-independent constitutive activity. *J Biol Chem* **271**, 1857-1860
60. Negishi, M., Hasegawa, H., and Ichikawa, A. (1996) Prostaglandin E receptor EP3gamma isoform, with mostly full constitutive Gi activity and agonist-dependent Gs activity. *FEBS Lett* **386**, 165-168
61. Coleman, R. A., Smith, W. L., and Narumiya, S. (1994) International Union of Pharmacology classification of prostanoid receptors: properties, distribution, and structure of the receptors and their subtypes. *Pharmacol Rev* **46**, 205-229

62. Sugimoto, Y., Shigemoto, R., Namba, T., Negishi, M., Mizuno, N., Narumiya, S., and Ichikawa, A. (1994) Distribution of the messenger RNA for the prostaglandin E receptor subtype EP3 in the mouse nervous system. *Neuroscience* **62**, 919-928
63. Cao, J., Shayibuzhati, M., Tajima, T., Kitazawa, T., and Taneike, T. (2002) In vitro pharmacological characterization of the prostanoid receptor population in the non-pregnant porcine myometrium. *Eur J Pharmacol* **442**, 115-123
64. Singh, J., Zeller, W., Zhou, N., Hategen, G., Mishra, R., Polozov, A., Yu, P., Onua, E., Zhang, J., Zembower, D., Kiselyov, A., Ramirez, J. L., Sigthorsson, G., Bjornsson, J. M., Thorsteinsdottir, M., Andresson, T., Bjarnadottir, M., Magnusson, O., Fabre, J. E., Stefansson, K., and Gurney, M. E. (2009) Antagonists of the EP(3) Receptor for Prostaglandin E(2) Are Novel Antiplatelet Agents That Do Not Prolong Bleeding. *ACS Chem Biol*
65. Amano, H., Hayashi, I., Endo, H., Kitasato, H., Yamashina, S., Maruyama, T., Kobayashi, M., Satoh, K., Narita, M., Sugimoto, Y., Murata, T., Yoshimura, H., Narumiya, S., and Majima, M. (2003) Host prostaglandin E(2)-EP3 signaling regulates tumor-associated angiogenesis and tumor growth. *J Exp Med* **197**, 221-232
66. Juteau, H., Gareau, Y., Labelle, M., Sturino, C. F., Sawyer, N., Tremblay, N., Lamontagne, S., Carriere, M. C., Denis, D., and Metters, K. M. (2001) Structure-activity relationship of cinnamic acylsulfonamide analogues on the human EP3 prostanoid receptor. *Bioorg Med Chem* **9**, 1977-1984
67. Kawahara, H., Sakamoto, A., Takeda, S., Onodera, H., Imaki, J., and Ogawa, R. (2001) A prostaglandin E2 receptor subtype EP1 receptor antagonist (ONO-8711) reduces hyperalgesia, allodynia, and c-fos gene expression in rats with chronic nerve constriction. *Anesth Analg* **93**, 1012-1017
68. Fujino, H., Xu, W., and Regan, J. W. (2003) Prostaglandin E2 induced functional expression of early growth response factor-1 by EP4, but not EP2, prostanoid receptors via the phosphatidylinositol 3-kinase and extracellular signal-regulated kinases. *J Biol Chem* **278**, 12151-12156
69. Fujino, H., and Regan, J. W. (2006) EP(4) prostanoid receptor coupling to a pertussis toxin-sensitive inhibitory G protein. *Mol Pharmacol* **69**, 5-10
70. Yoshida, K., Oida, H., Kobayashi, T., Maruyama, T., Tanaka, M., Katayama, T., Yamaguchi, K., Segi, E., Tsuboyama, T., Matsushita, M., Ito, K., Ito, Y., Sugimoto, Y., Ushikubi, F., Ohuchida, S., Kondo, K., Nakamura, T., and Narumiya, S. (2002) Stimulation of bone formation and

prevention of bone loss by prostaglandin E EP4 receptor activation. *Proc Natl Acad Sci U S A* **99**, 4580-4585

71. Davis, T. L., and Sharif, N. A. (2000) Pharmacological characterization of [(3)H]-prostaglandin E(2) binding to the cloned human EP(4) prostanoid receptor. *Br J Pharmacol* **130**, 1919-1926
72. Boie, Y., Stocco, R., Sawyer, N., Slipetz, D. M., Ungrin, M. D., Neuschafer-Rube, F., Puschel, G. P., Metters, K. M., and Abramovitz, M. (1997) Molecular cloning and characterization of the four rat prostaglandin E2 prostanoid receptor subtypes. *Eur J Pharmacol* **340**, 227-241
73. Nishigaki, N., Negishi, M., Honda, A., Sugimoto, Y., Namba, T., Narumiya, S., and Ichikawa, A. (1995) Identification of prostaglandin E receptor 'EP2' cloned from mastocytoma cells as EP4 subtype. *FEBS Lett* **364**, 339-341
74. Kabashima, K., Saji, T., Murata, T., Nagamachi, M., Matsuoka, T., Segi, E., Tsuboi, K., Sugimoto, Y., Kobayashi, T., Miyachi, Y., Ichikawa, A., and Narumiya, S. (2002) The prostaglandin receptor EP4 suppresses colitis, mucosal damage and CD4 cell activation in the gut. *J Clin Invest* **109**, 883-893
75. Machwate, M., Harada, S., Leu, C. T., Sedor, G., Labelle, M., Gallant, M., Hutchins, S., Lachance, N., Sawyer, N., Slipetz, D., Metters, K. M., Rodan, S. B., Young, R., and Rodan, G. A. (2001) Prostaglandin receptor EP(4) mediates the bone anabolic effects of PGE(2). *Mol Pharmacol* **60**, 36-41
76. Clark, P., Rowland, S. E., Denis, D., Mathieu, M. C., Stocco, R., Poirier, H., Burch, J., Han, Y., Audoly, L., Therien, A. G., and Xu, D. (2008) MF498 [N-{[4-(5,9-Diethoxy-6-oxo-6,8-dihydro-7H-pyrrolo[3,4-g]quinolin-7-yl)-3-m ethylbenzyl]sulfonyl}-2-(2-methoxyphenyl)acetamide], a selective E prostanoid receptor 4 antagonist, relieves joint inflammation and pain in rodent models of rheumatoid and osteoarthritis. *J Pharmacol Exp Ther* **325**, 425-434
77. Wilson, R. J., Giblin, G. M., Roomans, S., Rhodes, S. A., Cartwright, K. A., Shield, V. J., Brown, J., Wise, A., Chowdhury, J., Pritchard, S., Coote, J., Noel, L. S., Kenakin, T., Burns-Kurtis, C. L., Morrison, V., Gray, D. W., and Giles, H. (2006) GW627368X ((N-{2-[4-(4,9-diethoxy-1-oxo-1,3-dihydro-2H-benzo[f]isoindol-2-yl)phenyl]acetyl} benzene sulphonamide): a novel, potent and selective prostanoid EP4 receptor antagonist. *Br J Pharmacol* **148**, 326-339
78. Hizaki, H., Segi, E., Sugimoto, Y., Hirose, M., Saji, T., Ushikubi, F., Matsuoka, T., Noda, Y., Tanaka, T., Yoshida, N., Narumiya, S., and

- Ichikawa, A. (1999) Abortive expansion of the cumulus and impaired fertility in mice lacking the prostaglandin E receptor subtype EP(2). *Proc Natl Acad Sci U S A* **96**, 10501-10506
79. Stock, J. L., Shinjo, K., Burkhardt, J., Roach, M., Taniguchi, K., Ishikawa, T., Kim, H. S., Flannery, P. J., Coffman, T. M., McNeish, J. D., and Audoly, L. P. (2001) The prostaglandin E2 EP1 receptor mediates pain perception and regulates blood pressure. *J Clin Invest* **107**, 325-331
80. Minami, T., Nakano, H., Kobayashi, T., Sugimoto, Y., Ushikubi, F., Ichikawa, A., Narumiya, S., and Ito, S. (2001) Characterization of EP receptor subtypes responsible for prostaglandin E2-induced pain responses by use of EP1 and EP3 receptor knockout mice. *Br J Pharmacol* **133**, 438-444
81. Jones, R. L., Giembycz, M. A., and Woodward, D. F. (2009) Prostanoid receptor antagonists: development strategies and therapeutic applications. *Br J Pharmacol* **158**, 104-145
82. Drower, E. J., Stapelfeld, A., Mueller, R. A., and Hammond, D. L. (1987) The antinociceptive effects of prostaglandin antagonists in the rat. *Eur J Pharmacol* **133**, 249-256
83. Omote, K., Kawamata, T., Nakayama, Y., Kawamata, M., Hazama, K., and Namiki, A. (2001) The effects of peripheral administration of a novel selective antagonist for prostaglandin E receptor subtype EP(1), ONO-8711, in a rat model of postoperative pain. *Anesth Analg* **92**, 233-238
84. Syriatowicz, J. P., Hu, D., Walker, J. S., and Tracey, D. J. (1999) Hyperalgesia due to nerve injury: role of prostaglandins. *Neuroscience* **94**, 587-594
85. Guan, Y., Zhang, Y., Wu, J., Qi, Z., Yang, G., Dou, D., Gao, Y., Chen, L., Zhang, X., Davis, L. S., Wei, M., Fan, X., Carmosino, M., Hao, C., Imig, J. D., Breyer, R. M., and Breyer, M. D. (2007) Antihypertensive effects of selective prostaglandin E(2) receptor subtype 1 targeting. *J Clin Invest* **117**, 2496-2505
86. Rutkai, I., Feher, A., Erdei, N., Henrion, D., Papp, Z., Edes, I., Koller, A., Kaley, G., and Bagi, Z. (2009) Activation of prostaglandin E2 EP1 receptor increases arteriolar tone and blood pressure in mice with type 2 diabetes. *Cardiovasc Res* **83**, 148-154
87. Suganami, T., Mori, K., Tanaka, I., Mukoyama, M., Sugawara, A., Makino, H., Muro, S., Yahata, K., Ohuchida, S., Maruyama, T., Narumiya, S., and Nakao, K. (2003) Role of prostaglandin E receptor EP1 subtype in the

development of renal injury in genetically hypertensive rats. *Hypertension* **42**, 1183-1190

88. Makino, H., Tanaka, I., Mukoyama, M., Sugawara, A., Mori, K., Muro, S., Suganami, T., Yahata, K., Ishibashi, R., Ohuchida, S., Maruyama, T., Narumiya, S., and Nakao, K. (2002) Prevention of diabetic nephropathy in rats by prostaglandin E receptor EP1-selective antagonist. *J Am Soc Nephrol* **13**, 1757-1765
89. Tilley, S. L., Hartney, J. M., Erikson, C. J., Jania, C., Nguyen, M., Stock, J., McNeisch, J., Valancius, C., Panettieri, R. A., Jr., Penn, R. B., and Koller, B. H. (2003) Receptors and pathways mediating the effects of prostaglandin E2 on airway tone. *Am J Physiol Lung Cell Mol Physiol* **284**, L599-L606
90. Morsy, M. A., Isohama, Y., and Miyata, T. (2001) Prostaglandin E(2) increases surfactant secretion via the EP(1) receptor in rat alveolar type II cells. *Eur J Pharmacol* **426**, 21-24
91. Takeuchi, K., Ukawa, H., Furukawa, O., Kawauchi, S., Araki, H., Sugimoto, Y., Ishikawa, A., Ushikubi, F., and Narumiya, S. (1999) Prostaglandin E receptor subtypes involved in stimulation of gastroduodenal bicarbonate secretion in rats and mice. *J Physiol Pharmacol* **50**, 155-167
92. Takeuchi, K., Aihara, E., Hayashi, M., and Sasaki, Y. (2005) Role of prostaglandin E receptor subtypes in gastroduodenal HCO₃⁻ secretion. *Med Chem* **1**, 395-403
93. Kunikata, T., Araki, H., Takeeda, M., Kato, S., and Takeuchi, K. (2001) Prostaglandin E prevents indomethacin-induced gastric and intestinal damage through different EP receptor subtypes. *J Physiol Paris* **95**, 157-163
94. Takeuchi, K., Araki, H., Umeda, M., Komoike, Y., and Suzuki, K. (2001) Adaptive gastric cytoprotection is mediated by prostaglandin EP1 receptors: a study using rats and knockout mice. *J Pharmacol Exp Ther* **297**, 1160-1165
95. Kawamori, T., Kitamura, T., Watanabe, K., Uchiya, N., Maruyama, T., Narumiya, S., Sugimura, T., and Wakabayashi, K. (2005) Prostaglandin E receptor subtype EP(1) deficiency inhibits colon cancer development. *Carcinogenesis* **26**, 353-357
96. Kawamori, T., Uchiya, N., Nakatsugi, S., Watanabe, K., Ohuchida, S., Yamamoto, H., Maruyama, T., Kondo, K., Sugimura, T., and

- Wakabayashi, K. (2001) Chemopreventive effects of ONO-8711, a selective prostaglandin E receptor EP(1) antagonist, on breast cancer development. *Carcinogenesis* **22**, 2001-2004
97. Kunikata, T., Yamane, H., Segi, E., Matsuoka, T., Sugimoto, Y., Tanaka, S., Tanaka, H., Nagai, H., Ichikawa, A., and Narumiya, S. (2005) Suppression of allergic inflammation by the prostaglandin E receptor subtype EP3. *Nat Immunol* **6**, 524-531
 98. Hata, A. N., and Breyer, R. M. (2004) Pharmacology and signaling of prostaglandin receptors: Multiple roles in inflammation and immune modulation. *Pharmacol Ther* **103**, 147-166
 99. Tilley, S. L., Audoly, L. P., Hicks, E. H., Kim, H. S., Flannery, P. J., Coffman, T. M., and Koller, B. H. (1999) Reproductive failure and reduced blood pressure in mice lacking the EP2 prostaglandin E2 receptor. *J Clin Invest* **103**, 1539-1545
 100. Ochsner, S. A., Russell, D. L., Day, A. J., Breyer, R. M., and Richards, J. S. (2003) Decreased Expression of Tumor Necrosis Factor- α -Stimulated Gene 6 in Cumulus Cells of the Cyclooxygenase-2 and EP2 Null Mice. *Endocrinology* **144**, 1008-1019
 101. Armstrong, J. M., Boura, A. L., Hamberg, M., and Samuelsson, B. (1976) A comparison of the vasodepressor effects of the cyclic effects of the cyclic endoperoxides PGG₂, and PGH₂ with those of PGD₂ and PGE₂ in hypertensive and normotensive rats. *Eur J Pharmacol* **39**, 251-258
 102. Schweda, F., Klar, J., Narumiya, S., Nusing, R. M., and Kurtz, A. (2004) Stimulation of renin release by prostaglandin E₂ is mediated by EP2 and EP4 receptors in mouse kidneys. *Am J Physiol Renal Physiol* **287**, F427-433
 103. Friis, U. G., Stubbe, J., Uhrenholt, T. R., Svenningsen, P., Nusing, R. M., Skott, O., and Jensen, B. L. (2005) Prostaglandin E₂ EP2 and EP4 receptor activation mediates cAMP-dependent hyperpolarization and exocytosis of renin in juxtaglomerular cells. *Am J Physiol Renal Physiol* **289**, F989-997
 104. Imig, J. D., Breyer, M. D., and Breyer, R. M. (2002) Contribution of prostaglandin EP(2) receptors to renal microvascular reactivity in mice. *Am J Physiol Renal Physiol* **283**, F415-F422
 105. Chen, J., Zhao, M., He, W., Milne, G. L., Howard, J. R., Morrow, J., Hebert, R. L., Breyer, R. M., and Hao, C. M. (2008) Increased dietary

- NaCl induces renal medullary PGE₂ production and natriuresis via the EP₂ receptor. *Am J Physiol Renal Physiol* **295**, F818-825
106. Norel, X., Walch, L., Labat, C., Gascard, J. P., Dulmet, E., and Brink, C. (1999) Prostanoid receptors involved in the relaxation of human bronchial preparations. *Br J Pharmacol* **126**, 867-872
 107. Sheller, J. R., Mitchell, D., Meyrick, B., Oates, J., and Breyer, R. (2000) EP₂ receptor mediates bronchodilation by PGE₂ in mice. *J Appl Physiol* **88**, 2214-2218
 108. Hartney, J. M., Coggins, K. G., Tilley, S. L., Jania, L. A., Lovgren, A. K., Audoly, L. P., and Koller, B. H. (2006) Prostaglandin E₂ protects lower airways against bronchoconstriction. *Am J Physiol Lung Cell Mol Physiol* **290**, L105-113
 109. Kay, L. J., Yeo, W. W., and Peachell, P. T. (2006) Prostaglandin E₂ activates EP₂ receptors to inhibit human lung mast cell degranulation. *Br J Pharmacol* **147**, 707-713
 110. Nials, A., Vardey, C., Denyer, L., Thomas, M., Sparrow, S., Shepherd, G., and Coleman, R. (1993) AH 13205, a selective prostanoid EP₂-receptor agonist. *Cardiovasc Drug Rev* **11**, 165
 111. Sung, Y. M., He, G., and Fischer, S. M. (2005) Lack of expression of the EP₂ but not EP₃ receptor for prostaglandin E₂ results in suppression of skin tumor development. *Cancer Res* **65**, 9304-9311
 112. Sung, Y. M., He, G., Hwang, D. H., and Fischer, S. M. (2006) Overexpression of the prostaglandin E₂ receptor EP₂ results in enhanced skin tumor development. *Oncogene* **25**, 5507-5516
 113. Kamiyama, M., Pozzi, A., Yang, L., Debusk, L. M., Breyer, R. M., and Lin, P. C. (2006) EP₂, a receptor for PGE₂, regulates tumor angiogenesis through direct effects on endothelial cell motility and survival. *Oncogene*
 114. Tian, M., and Schiemann, W. P. (2010) PGE₂ receptor EP₂ mediates the antagonistic effect of COX-2 on TGF- β signaling during mammary tumorigenesis. *FASEB J* **24**, 1105-1116
 115. Konger, R. L., Scott, G. A., Landt, Y., Ladenson, J. H., and Pentland, A. P. (2002) Loss of the EP₂ prostaglandin E₂ receptor in immortalized human keratinocytes results in increased invasiveness and decreased paxillin expression. *Am J Pathol* **161**, 2065-2078

116. Brouxhon, S., Konger, R. L., VanBuskirk, J., Sheu, T. J., Ryan, J., Erdle, B., Almudevar, A., Breyer, R. M., Scott, G., and Pentland, A. P. (2007) Deletion of prostaglandin E2 EP2 receptor protects against ultraviolet-induced carcinogenesis, but increases tumor aggressiveness. *J Invest Dermatol* **127**, 439-446
117. Fleming, E. F., Athirakul, K., Oliverio, M. I., Key, M., Goulet, J., Koller, B. H., and Coffman, T. M. (1998) Urinary concentrating function in mice lacking EP3 receptors for prostaglandin E2. *Am J Physiol* **275**, F955-961
118. Ushikubi, F., Segi, E., Sugimoto, Y., Murata, T., Matsuoka, T., Kobayashi, T., Hizaki, H., Tuboi, K., Katsuyama, M., Ichikawa, A., Tanaka, T., Yoshida, N., and Narumiya, S. (1998) Impaired febrile response in mice lacking the prostaglandin E receptor subtype EP3. *Nature* **395**, 281-284
119. Takasaki, I., Nojima, H., Shiraki, K., Sugimoto, Y., Ichikawa, A., Ushikubi, F., Narumiya, S., and Kuraishi, Y. (2005) Involvement of cyclooxygenase-2 and EP3 prostaglandin receptor in acute herpetic but not postherpetic pain in mice. *Neuropharmacology* **49**, 283-292
120. Armstrong, R. A. (1996) Platelet prostanoid receptors. *Pharmacol Ther* **72**, 171-191
121. Matthews, J. S., and Jones, R. L. (1993) Potentiation of aggregation and inhibition of adenylate cyclase in human platelets by prostaglandin E analogues. *Br J Pharmacol* **108**, 363-369
122. Fabre, J. E., Nguyen, M., Athirakul, K., Coggins, K., McNeish, J. D., Austin, S., Parise, L. K., FitzGerald, G. A., Coffman, T. M., and Koller, B. H. (2001) Activation of the murine EP3 receptor for PGE2 inhibits cAMP production and promotes platelet aggregation. *J Clin Invest* **107**, 603-610
123. Ma, H., Hara, A., Xiao, C. Y., Okada, Y., Takahata, O., Nakaya, K., Sugimoto, Y., Ichikawa, A., Narumiya, S., and Ushikubi, F. (2001) Increased bleeding tendency and decreased susceptibility to thromboembolism in mice lacking the prostaglandin E receptor subtype EP(3). *Circulation* **104**, 1176-1180
124. Gross, S., Tilly, P., Hentsch, D., Vonesch, J. L., and Fabre, J. E. (2007) Vascular wall-produced prostaglandin E2 exacerbates arterial thrombosis and atherothrombosis through platelet EP3 receptors. *J Exp Med*
125. Su, X., Leon, L. A., Wu, C. W., Morrow, D. M., Jaworski, J. P., Hieble, J. P., Lashinger, E. S., Jin, J., Edwards, R. M., and Laping, N. J. (2008) Modulation of bladder function by prostaglandin EP3 receptors in the central nervous system. *Am J Physiol Renal Physiol* **295**, F984-994

126. McCafferty, G. P., Misajet, B. A., Laping, N. J., Edwards, R. M., and Thorneloe, K. S. (2008) Enhanced bladder capacity and reduced prostaglandin E2-mediated bladder hyperactivity in EP3 receptor knockout mice. *Am J Physiol Renal Physiol* **295**, F507-514
127. Sanchez-Alavez, M., Klein, I., Brownell, S. E., Tabarean, I. V., Davis, C. N., Conti, B., and Bartfai, T. (2007) Night eating and obesity in the EP3R-deficient mouse. *Proc Natl Acad Sci U S A* **104**, 3009-3014
128. Shi, J., Wang, Q., Johansson, J. U., Liang, X., Woodling, N. S., Priyam, P., Loui, T. M., Merchant, M., Breyer, R. M., Montine, T. J., and Andreasson, K. (2012) Inflammatory prostaglandin E2 signaling in a mouse model of Alzheimer's disease. *Annals of Neurology*
129. Wood, H. (2012) Alzheimer disease: Prostaglandin E2 signalling is implicated in inflammation early in the Alzheimer disease course. *Nat Rev Neurol* **advance online publication**
130. Shoji, Y., Takahashi, M., Kitamura, T., Watanabe, K., Kawamori, T., Maruyama, T., Sugimoto, Y., Negishi, M., Narumiya, S., Sugimura, T., and Wakabayashi, K. (2004) Downregulation of prostaglandin E receptor subtype EP3 during colon cancer development. *Gut* **53**, 1151-1158
131. Richards, J. A., and Brueggemeier, R. W. (2003) Prostaglandin E2 regulates aromatase activity and expression in human adipose stromal cells via two distinct receptor subtypes. *J Clin Endocrinol Metab* **88**, 2810-2816
132. Nguyen, M., Camenisch, T., Snouwaert, J. N., Hicks, E., Coffman, T. M., Anderson, P. A., Malouf, N. N., and Koller, B. H. (1997) The prostaglandin receptor EP4 triggers remodelling of the cardiovascular system at birth. *Nature* **390**, 78-81
133. Segi, E., Sugimoto, Y., Yamasaki, A., Aze, Y., Oida, H., Nishimura, T., Murata, T., Matsuoka, T., Ushikubi, F., Hirose, M., Tanaka, T., Yoshida, N., Narumiya, S., and Ichikawa, A. (1998) Patent ductus arteriosus and neonatal death in prostaglandin receptor EP4-deficient mice. *Biochem Biophys Res Commun* **246**, 7-12
134. Breyer, M. D., and Breyer, R. M. (2000) Prostaglandin E receptors and the kidney. *Am J Physiol Renal Physiol* **279**, F12-F23
135. Peti-Peterdi, J., Komlosi, P., Fuson, A. L., Guan, Y., Schneider, A., Qi, Z., Redha, R., Rosivall, L., Breyer, M. D., and Bell, P. D. (2003) Luminal NaCl delivery regulates basolateral PGE2 release from macula densa cells. *J Clin Invest* **112**, 76-82

136. Li, M., Healy, D. R., Li, Y., Simmons, H. A., Crawford, D. T., Ke, H. Z., Pan, L. C., Brown, T. A., and Thompson, D. D. (2005) Osteopenia and impaired fracture healing in aged EP4 receptor knockout mice. *Bone* **37**, 46-54
137. Akhter, M. P., Cullen, D. M., Gong, G., and Recker, R. R. (2001) Bone biomechanical properties in prostaglandin EP1 and EP2 knockout mice. *Bone* **29**, 121-125
138. Audoly, L. P., Tilley, S. L., Goulet, J., Key, M., Nguyen, M., Stock, J. L., McNeish, J. D., Koller, B. H., and Coffman, T. M. (1999) Identification of specific EP receptors responsible for the hemodynamic effects of PGE₂. *Am J Physiol* **277**, H924-930
139. Zhang, Y., Guan, Y., Schneider, A., Brandon, S., Breyer, R. M., and Breyer, M. D. (2000) Characterization of Murine Vasopressor and Vasodepressor Prostaglandin E(2) Receptors. *Hypertension* **35**, 1129-1134
140. Audoly, L. P., Ruan, X., Wagner, V. A., Goulet, J. L., Tilley, S. L., Koller, B. H., Coffman, T. M., and Arendshorst, W. J. (2001) Role of EP(2) and EP(3) PGE(2) receptors in control of murine renal hemodynamics. *Am J Physiol Heart Circ Physiol* **280**, H327-333
141. Gurwitz, J., Avorn, J., Bohn, R., Glynn, R., Monane, M., and Mogun, H. (1994) Initiation of antihypertensive treatment during nonsteroidal anti-inflammatory drug therapy. *J. Am. Med. Assoc.* **272**, 781-786
142. Gurwitz, J. H., Avorn, J., Bohn, R. L., Glynn, R. J., Monane, M., and Mogun, H. (1994) Initiation of antihypertensive treatment during nonsteroidal anti-inflammatory drug therapy. *JAMA* **272**, 781-786
143. Imanishi, M., Kawamura, M., Akabane, S., Matsushima, Y., Kuramochi, M., Ito, K., Ohta, M., Kimura, K., Takamiya, M., and Omae, T. (1989) Aspirin lowers blood pressure in patients with renovascular hypertension. *Hypertension* **14**, 461-468
144. Breyer, M. D., and Breyer, R. M. (2000) Prostaglandin receptors: their role in regulating renal function. *Curr Opin Nephrol Hypertens* **9**, 23-29
145. Jakobsson, P. J., Thoren, S., Morgenstern, R., and Samuelsson, B. (1999) Identification of human prostaglandin E synthase: a microsomal, glutathione-dependent, inducible enzyme, constituting a potential novel drug target. *Proc Natl Acad Sci U S A* **96**, 7220-7225

146. Facemire, C. S., Griffiths, R., Audoly, L. P., Koller, B. H., and Coffman, T. M. (2010) The impact of microsomal prostaglandin e synthase 1 on blood pressure is determined by genetic background. *Hypertension* **55**, 531-538
147. Zhang, D. J., Chen, L. H., Zhang, Y. H., Yang, G. R., Dou, D., Gao, Y. S., Zhang, X. Y., Kong, X. M., Zhao, P., Pu, D., Wei, M. F., Breyer, M. D., and Guan, Y. F. (2010) Enhanced pressor response to acute Ang II infusion in mice lacking membrane-associated prostaglandin E2 synthase-1. *Acta Pharmacol Sin* **31**, 1284-1292
148. Jia, Z., Aoyagi, T., and Yang, T. (2010) mPGES-1 protects against DOCA-salt hypertension via inhibition of oxidative stress or stimulation of NO/cGMP. *Hypertension* **55**, 539-546
149. Ruilope, L., Garcia Robles, R., Barrientos, A., Bernis, C., Alcazar, J., Tresguerres, J. A., Mancheño, E., Millet, V. G., Sancho, J., and Rodicio, J. L. (1982) The role of urinary PGE2 and renin-angiotensin-aldosterone system in the pathogenesis of essential hypertension. *Clin Exp Hypertens A* **4**, 989-1000
150. Eklund, B., and Carlson, L. A. (1980) Central and peripheral circulatory effects and metabolic effects of different prostaglandins given I.V. to man. *Prostaglandins* **20**, 333-347
151. Murray, M., Breene, P., Brater, D., Manatunga, A., and Hall, S. (1992) Effect of flurbiprofen on renal function in patients with moderate renal insufficiency. *Br. J. Clin. Pharm.* **33**, 385-393
152. Johnson, A. G. (1997) NSAIDs and increased blood pressure. What is the clinical significance? *Drug Saf* **17**, 277-289
153. Cheng, H. F., and Harris, R. C. (2004) Cyclooxygenases, the kidney, and hypertension. *Hypertension* **43**, 525-530
154. Solomon, D. H., Schneeweiss, S., Levin, R., and Avorn, J. (2004) Relationship between COX-2 specific inhibitors and hypertension. *Hypertension* **44**, 140-145
155. Mori, A., Saito, M., Sakamoto, K., Narita, M., Nakahara, T., and Ishii, K. (2007) Stimulation of prostanoid IP and EP(2) receptors dilates retinal arterioles and increases retinal and choroidal blood flow in rats. *Eur J Pharmacol* **570**, 135-141
156. Hristovska, A. M., Rasmussen, L. E., Hansen, P. B., Nielsen, S. S., Nusing, R. M., Narumiya, S., Vanhoutte, P., Skott, O., and Jensen, B. L. (2007) Prostaglandin e2 induces vascular relaxation by e-prostanoid 4

- receptor-mediated activation of endothelial nitric oxide synthase. *Hypertension* **50**, 525-530
157. Ariumi, H., Takano, Y., Masumi, A., Takahashi, S., Hirabara, Y., Honda, K., Saito, R., and Kamiya, H. O. (2002) Roles of the central prostaglandin EP3 receptors in cardiovascular regulation in rats. *Neurosci Lett* **324**, 61-64
 158. Flack, J. M. (2007) Epidemiology and unmet needs in hypertension. *J Manag Care Pharm* **13**, 2-8
 159. Wang, M., Ihida-Stansbury, K., Kothapalli, D., Tamby, M. C., Yu, Z., Chen, L., Grant, G., Cheng, Y., Lawson, J. A., Assoian, R. K., Jones, P. L., and Fitzgerald, G. A. (2011) Microsomal prostaglandin e2 synthase-1 modulates the response to vascular injury. *Circulation* **123**, 631-639
 160. Wang, M., Lee, E., Song, W., Ricciotti, E., Rader, D. J., Lawson, J. A., Pure, E., and FitzGerald, G. A. (2008) Microsomal prostaglandin E synthase-1 deletion suppresses oxidative stress and angiotensin II-induced abdominal aortic aneurysm formation. *Circulation* **117**, 1302-1309
 161. Capone, C., Faraco, G., Anrather, J., Zhou, P., and Iadecola, C. (2010) Cyclooxygenase 1-derived prostaglandin E2 and EP1 receptors are required for the cerebrovascular dysfunction induced by angiotensin II. *Hypertension* **55**, 911-917
 162. Xiao, C. Y., Yuhki, K., Hara, A., Fujino, T., Kuriyama, S., Yamada, T., Takayama, K., Takahata, O., Karibe, H., Taniguchi, T., Narumiya, S., and Ushikubi, F. (2004) Prostaglandin E2 protects the heart from ischemia-reperfusion injury via its receptor subtype EP4. *Circulation* **109**, 2462-2468
 163. Kirchhoff, F., Krebs, C., Abdulhag, U. N., Meyer-Schwesinger, C., Maas, R., Helmchen, U., Hilgers, K. F., Wolf, G., Stahl, R. A., and Wenzel, U. (2008) Rapid development of severe end-organ damage in C57BL/6 mice by combining DOCA salt and angiotensin II. *Kidney Int* **73**, 643-650
 164. Manning, M. W., Cassi, L. A., Huang, J., Szilvassy, S. J., and Daugherty, A. (2002) Abdominal aortic aneurysms: fresh insights from a novel animal model of the disease. *Vasc Med* **7**, 45-54
 165. Guan, Y., Hao, C., Cha, D. R., Rao, R., Lu, W., Kohan, D. E., Magnuson, M. A., Redha, R., Zhang, Y., and Breyer, M. D. (2005) Thiazolidinediones expand body fluid volume through PPARgamma stimulation of ENaC-mediated renal salt absorption. *Nat Med* **11**, 861-866

166. Gitlin, J. M., Trivedi, D. B., Langenbach, R., and Loftin, C. D. (2007) Genetic deficiency of cyclooxygenase-2 attenuates abdominal aortic aneurysm formation in mice. *Cardiovasc Res* **73**, 227-236
167. King, V. L., Trivedi, D. B., Gitlin, J. M., and Loftin, C. D. (2006) Selective cyclooxygenase-2 inhibition with celecoxib decreases angiotensin II-induced abdominal aortic aneurysm formation in mice. *Arterioscler Thromb Vasc Biol* **26**, 1137-1143
168. Cho, S., and Atwood, J. E. (2002) Peripheral edema. *Am J Med* **113**, 580-586
169. Kemp, C. D., and Conte, J. V. (2012) The pathophysiology of heart failure. *Cardiovasc Pathol*
170. Guan, Y., Zhang, Y., Wu, J., Qi, Z., Yang, G., Dou, D., Gao, Y., Chen, L., Zhang, X., Davis, L., Wei, M., Fan, X., Carmosino, M., Hao, C., Imig, J., Breyer, R., and Breyer, M. (2007) Antihypertensive effects of selective prostaglandin E2 receptor subtype 1 targeting. *J Clin Invest* **117**, 2496-2505
171. Daugherty, A., and Cassis, L. A. (2004) Mouse models of abdominal aortic aneurysms. *Arterioscler Thromb Vasc Biol* **24**, 429-434
172. Daugherty, A., Cassis, L. A., and Lu, H. (2011) Complex pathologies of angiotensin II-induced abdominal aortic aneurysms. *J Zhejiang Univ Sci B* **12**, 624-628
173. Rahal, S., McVeigh, L. I., Zhang, Y., Guan, Y., Breyer, M. D., and Kennedy, C. R. (2006) Increased severity of renal impairment in nephritic mice lacking the EP1 receptor. *Can J Physiol Pharmacol* **84**, 877-885
174. Ashida, T., Nishioeda, Y., Kimura, G., Kojima, S., Kawamura, M., Imanishi, M., Abe, H., Kawano, Y., Yoshimi, H., Yoshida, K., and et al. (1989) Effects of salt, prostaglandin, and captopril on vascular responsiveness in essential hypertension. *Am J Hypertens* **2**, 640-642
175. Goodfriend, T. L., Elliott, M. E., and Catt, K. J. (1996) Angiotensin receptors and their antagonists. *N Engl J Med* **334**, 1649-1654
176. Griendling, K. K., Lassegue, B., and Alexander, R. W. (1996) Angiotensin receptors and their therapeutic implications. *Annu Rev Pharmacol Toxicol* **36**, 281-306

177. Morsing, P. (1999) Candesartan: a new-generation angiotensin II AT1 receptor blocker: pharmacology, antihypertensive efficacy, renal function, and renoprotection. *J Am Soc Nephrol* **10 Suppl 11**, S248-254
178. Lewis, E. J., Hunsicker, L. G., Clarke, W. R., Berl, T., Pohl, M. A., Lewis, J. B., Ritz, E., Atkins, R. C., Rohde, R., and Raz, I. (2001) Renoprotective effect of the angiotensin-receptor antagonist irbesartan in patients with nephropathy due to type 2 diabetes. *N Engl J Med* **345**, 851-860
179. Murphy, T. J., Alexander, R. W., Griendling, K. K., Runge, M. S., and Bernstein, K. E. (1991) Isolation of a cDNA encoding the vascular type-1 angiotensin II receptor. *Nature* **351**, 233-236
180. Sayeski, P. P., Ali, M. S., Semeniuk, D. J., Doan, T. N., and Bernstein, K. E. (1998) Angiotensin II signal transduction pathways. *Regul Pept* **78**, 19-29
181. Xie, Z., Gong, M. C., Su, W., Turk, J., and Guo, Z. (2007) Group VIA Phospholipase A2 (iPLA2beta) Participates in Angiotensin II-induced Transcriptional Up-regulation of Regulator of G-protein Signaling-2 in Vascular Smooth Muscle Cells. *J Biol Chem* **282**, 25278-25289
182. Vagnes, O. B., Iversen, B. M., and Arendshorst, W. J. (2007) Short-term ANG II produces renal vasoconstriction independent of TP receptor activation and TxA2/isoprostane production. *Am J Physiol Renal Physiol* **293**, F860-867
183. Higuchi, S., Ohtsu, H., Suzuki, H., Shirai, H., Frank, G. D., and Eguchi, S. (2007) Angiotensin II signal transduction through the AT1 receptor: novel insights into mechanisms and pathophysiology. *Clin Sci (Lond)* **112**, 417-428
184. Alvarez, Y., Perez-Giron, J. V., Hernanz, R., Briones, A. M., Garcia-Redondo, A., Beltran, A., Alonso, M. J., and Salaices, M. (2007) Losartan reduces the increased participation of cyclooxygenase-2-derived products in vascular responses of hypertensive rats. *J Pharmacol Exp Ther* **321**, 381-388
185. Chen, J., Chen, J. K., Neilson, E. G., and Harris, R. C. (2006) Role of EGF receptor activation in angiotensin II-induced renal epithelial cell hypertrophy. *J Am Soc Nephrol* **17**, 1615-1623
186. De Paolis, P., Porcellini, A., Savoia, C., Lombardi, A., Gigante, B., Frati, G., Rubattu, S., Musumeci, B., and Volpe, M. (2002) Functional cross-talk between angiotensin II and epidermal growth factor receptors in NIH3T3 fibroblasts. *J Hypertens* **20**, 693-699

187. Nataraj, C., Oliverio, M. I., Mannon, R. B., Mannon, P. J., Audoly, L. P., Amuchastegui, C. S., Ruiz, P., Smithies, O., and Coffman, T. M. (1999) Angiotensin II regulates cellular immune responses through a calcineurin-dependent pathway. *J Clin Invest* **104**, 1693-1701
188. Inagami, T. (1999) Molecular biology and signaling of angiotensin receptors: an overview. *J Am Soc Nephrol* **10 Suppl 11**, S2-7
189. Breyer, R. M., Bagdassarian, C. K., Myers, S. A., and Breyer, M. D. (2001) Prostanoid receptors: subtypes and signaling. *Annu Rev Pharmacol Toxicol* **41**, 661-690
190. Hocherl, K., Endemann, D., Kammerl, M. C., Grobecker, H. F., and Kurtz, A. (2002) Cyclo-oxygenase-2 inhibition increases blood pressure in rats. *Br J Pharmacol* **136**, 1117-1126
191. Kawada, N., Solis, G., Ivey, N., Connors, S., Dennehy, K., Modlinger, P., Hamel, R., Kawada, J. T., Imai, E., Langenbach, R., Welch, W. J., and Wilcox, C. S. (2005) Cyclooxygenase-1-deficient mice have high sleep-to-wake blood pressure ratios and renal vasoconstriction. *Hypertension* **45**, 1131-1138
192. Ye, W., Zhang, H., Hillas, E., Kohan, D. E., Miller, R. L., Nelson, R. D., Honeggar, M., and Yang, T. (2006) Expression and function of COX isoforms in renal medulla: evidence for regulation of salt sensitivity and blood pressure. *Am J Physiol Renal Physiol* **290**, F542-549
193. Mtabaji, J. P., Kihara, M., and Yamori, Y. (1985) The effects of indomethacin and PGE2 on vascular reactivity in spontaneously hypertensive rats: possible role of prostaglandins in the pathogenesis of hypertension. *Prostaglandins Leukot Med* **20**, 255-265
194. Breyer, M. D., Qi, Z., Tchekneva, E. E., and Harris, R. C. (2008) Insight into the genetics of diabetic nephropathy through the study of mice. *Curr Opin Nephrol Hypertens* **17**, 82-86
195. Kanetsuna, Y., Takahashi, K., Nagata, M., Gannon, M. A., Breyer, M. D., Harris, R. C., and Takahashi, T. (2007) Deficiency of endothelial nitric-oxide synthase confers susceptibility to diabetic nephropathy in nephropathy-resistant inbred mice. *Am J Pathol* **170**, 1473-1484
196. Grantham, J. J., and Orloff, J. (1968) Effect of prostaglandin E1 on the permeability response of the isolated collecting tubule to vasopressin, adenosine 3',5'-monophosphate, and theophylline. *J Clin Invest* **47**, 1154-1161

197. Hebert, R. L., Jacobson, H. R., Fredin, D., and Breyer, M. D. (1993) Evidence that separate PGE2 receptors modulate water and sodium transport in rabbit cortical collecting duct. *Am J Physiol* **265**, F643-650
198. Foundation, N. K. (2002) K/DOQI clinical practice guidelines for chronic kidney disease: evaluation, classification, and stratification. *Am J Kidney Dis* **39**, S1-266
199. Fox, C. S., Larson, M. G., Leip, E. P., Culeton, B., Wilson, P. W., and Levy, D. (2004) Predictors of new-onset kidney disease in a community-based population. *JAMA* **291**, 844-850
200. Klag, M. J., Whelton, P. K., Randall, B. L., Neaton, J. D., Brancati, F. L., Ford, C. E., Shulman, N. B., and Stamler, J. (1996) Blood pressure and end-stage renal disease in men. *N Engl J Med* **334**, 13-18
201. Bakris, G. L., Williams, M., Dworkin, L., Elliott, W. J., Epstein, M., Toto, R., Tuttle, K., Douglas, J., Hsueh, W., and Sowers, J. (2000) Preserving renal function in adults with hypertension and diabetes: a consensus approach. National Kidney Foundation Hypertension and Diabetes Executive Committees Working Group. *Am J Kidney Dis* **36**, 646-661
202. Parving, H. H., Hommel, E., Damkjaer Nielsen, M., and Giese, J. (1989) Effect of captopril on blood pressure and kidney function in normotensive insulin dependent diabetics with nephropathy. *BMJ* **299**, 533-536
203. Hebert, L. A., Bain, R. P., Verme, D., Cattran, D., Whittier, F. C., Tolchin, N., Rohde, R. D., and Lewis, E. J. (1994) Remission of nephrotic range proteinuria in type I diabetes. Collaborative Study Group. *Kidney Int* **46**, 1688-1693
204. Klahr, S., Levey, A. S., Beck, G. J., Caggiula, A. W., Hunsicker, L., Kusek, J. W., and Striker, G. (1994) The effects of dietary protein restriction and blood-pressure control on the progression of chronic renal disease. Modification of Diet in Renal Disease Study Group. *N Engl J Med* **330**, 877-884
205. Viberti, G., Mogensen, C. E., Groop, L. C., and Pauls, J. F. (1994) Effect of captopril on progression to clinical proteinuria in patients with insulin-dependent diabetes mellitus and microalbuminuria. European Microalbuminuria Captopril Study Group. *JAMA* **271**, 275-279
206. Lebovitz, H. E., Wiegmann, T. B., Cnaan, A., Shahinfar, S., Sica, D. A., Broadstone, V., Schwartz, S. L., Mengel, M. C., Segal, R., and Versaggi, J. A. (1994) Renal protective effects of enalapril in hypertensive NIDDM: role of baseline albuminuria. *Kidney Int Suppl* **45**, S150-155

207. Bakris, G. L., Mangrum, A., Copley, J. B., Vicknair, N., and Sadler, R. (1997) Effect of calcium channel or beta-blockade on the progression of diabetic nephropathy in African Americans. *Hypertension* **29**, 744-750
208. (1997) Randomised placebo-controlled trial of effect of ramipril on decline in glomerular filtration rate and risk of terminal renal failure in proteinuric, non-diabetic nephropathy. The GISEN Group (Gruppo Italiano di Studi Epidemiologici in Nefrologia). *Lancet* **349**, 1857-1863
209. Bakris, G. L., Weir, M. R., Shanifar, S., Zhang, Z., Douglas, J., van Dijk, D. J., Brenner, B. M., and Group, R. S. (2003) Effects of blood pressure level on progression of diabetic nephropathy: results from the RENAAL study. *Arch Intern Med* **163**, 1555-1565
210. Remuzzi, G., Benigni, A., and Remuzzi, A. (2006) Mechanisms of progression and regression of renal lesions of chronic nephropathies and diabetes. *J Clin Invest* **116**, 288-296
211. Wolf, G., Butzmann, U., and Wenzel, U. O. (2003) The renin-angiotensin system and progression of renal disease: from hemodynamics to cell biology. *Nephron Physiol* **93**, P3-13
212. Wolf, G., and Neilson, E. G. (1996) From converting enzyme inhibition to angiotensin II receptor blockade: new insight on angiotensin II receptor subtypes in the kidney. *Exp Nephrol* **4 Suppl 1**, 8-19
213. Ruster, C., and Wolf, G. (2006) Renin-angiotensin-aldosterone system and progression of renal disease. *J Am Soc Nephrol* **17**, 2985-2991
214. Lautrette, A., Li, S., Alili, R., Sunnarborg, S. W., Burtin, M., Lee, D. C., Friedlander, G., and Terzi, F. (2005) Angiotensin II and EGF receptor cross-talk in chronic kidney diseases: a new therapeutic approach. *Nat Med* **11**, 867-874
215. Crowley, S. D., Gurley, S. B., Herrera, M. J., Ruiz, P., Griffiths, R., Kumar, A. P., Kim, H. S., Smithies, O., Le, T. H., and Coffman, T. M. (2006) Angiotensin II causes hypertension and cardiac hypertrophy through its receptors in the kidney. *Proc Natl Acad Sci U S A* **103**, 17985-17990
216. Heuser, J. E., and Anderson, R. G. (1989) Hypertonic media inhibit receptor-mediated endocytosis by blocking clathrin-coated pit formation. *J Cell Biol* **108**, 389-400
217. Brenner, B. M., Cooper, M. E., de Zeeuw, D., Keane, W. F., Mitch, W. E., Parving, H. H., Remuzzi, G., Snapinn, S. M., Zhang, Z., and Shahinfar, S. (2001) Effects of losartan on renal and cardiovascular outcomes in

- patients with type 2 diabetes and nephropathy. *N Engl J Med* **345**, 861-869
218. Lewis, E. J., Hunsicker, L. G., Bain, R. P., and Rohde, R. D. (1993) The effect of angiotensin-converting-enzyme inhibition on diabetic nephropathy. The Collaborative Study Group. *N Engl J Med* **329**, 1456-1462
 219. Linz, W., Wiemer, G., Schaper, J., Zimmermann, R., Nagasawa, K., Gohlke, P., Unger, T., and Schölkens, B. A. (1995) Angiotensin converting enzyme inhibitors, left ventricular hypertrophy and fibrosis. *Mol Cell Biochem* **147**, 89-97
 220. Schölkens, B. A., Linz, W., and Martorana, P. A. (1991) Experimental cardiovascular benefits of angiotensin-converting enzyme inhibitors: beyond blood pressure reduction. *J Cardiovasc Pharmacol* **18 Suppl 2**, S26-30
 221. Okamoto, K., Ohta, Y., Chikugo, T., Shiokawa, H., and Morita, N. (1991) Chronic treatment with captopril, SQ29,852, hydralazine and a 33% fish meal diet in malignant stroke-prone spontaneously hypertensive rats. *J Hypertens* **9**, 1105-1117
 222. Hilleman, D. E., and Lucas, B. D. (2004) Angiotensin-converting enzyme inhibitors and stroke risk: benefit beyond blood pressure reduction? *Pharmacotherapy* **24**, 1064-1076
 223. Qi, Z., Whitt, I., Mehta, A., Jin, J., Zhao, M., Harris, R. C., Fogo, A. B., and Breyer, M. D. (2004) Serial determination of glomerular filtration rate in conscious mice using FITC-inulin clearance. *Am J Physiol Renal Physiol* **286**, F590-596
 224. Bartlett, C.S., Boyd, K.L., Harris, R.C., Zent, R., and Breyer, R.M. (2012) EP1 disruption attenuates end-organ damage in a mouse model of hypertension. *Hypertension*. Nov 2012.
 225. Sherwin, R. S., Anderson, R. M., Buse, J. B., Chin, M. H., Eddy, D., Fradkin, J., Ganiats, T. G., Ginsberg, H. N., Kahn, R., Nwankwo, R., Rewers, M., Schlessinger, L., Stern, M., Vinicor, F., Zinman, B., Association, A. D., and Diseases, N. I. o. D. a. D. a. K. (2004) Prevention or delay of type 2 diabetes. *Diabetes Care* **27 Suppl 1**, S47-54
 226. Pyram, R., Kansara, A., Banerji, M. A., and Loney-Hutchinson, L. (2012) Chronic kidney disease and diabetes. *Maturitas* **71**, 94-103

227. Breyer, M. D., Bottinger, E., Brosius, F. C., 3rd, Coffman, T. M., Harris, R. C., Heilig, C. W., and Sharma, K. (2005) Mouse models of diabetic nephropathy. *J Am Soc Nephrol* **16**, 27-45
228. McGowan, T., McCue, P., and Sharma, K. (2001) Diabetic nephropathy. *Clin Lab Med* **21**, 111-146
229. Arauz-Pacheco, C., Parrott, M. A., Raskin, P., and Association, A. D. (2004) Hypertension management in adults with diabetes. *Diabetes Care* **27 Suppl 1**, S65-67
230. Komers, R., Lindsley, J. N., Oyama, T. T., Schutzer, W. E., Reed, J. F., Mader, S. L., and Anderson, S. (2001) Immunohistochemical and functional correlations of renal cyclooxygenase- 2 in experimental diabetes. *J Clin Invest* **107**, 889-898.
231. Cheng, H. F., Wang, S. W., Zhang, M. Z., McKanna, J. A., Breyer, R., and Harris, R. C. (2002) Prostaglandins that increase renin production in response to ACE inhibition are not derived from cyclooxygenase-1. *Am J Physiol Regul Integr Comp Physiol* **283**, R638-646
232. Dey, A., Williams, R. S., Pollock, D. M., Stepp, D. W., Newman, J. W., Hammock, B. D., and Imig, J. D. (2004) Altered kidney CYP2C and cyclooxygenase-2 levels are associated with obesity-related albuminuria. *Obes Res* **12**, 1278-1289
233. Komers, R., Zdychová, J., Cahová, M., Kazdová, L., Lindsley, J. N., and Anderson, S. (2005) Renal cyclooxygenase-2 in obese Zucker (fatty) rats. *Kidney Int* **67**, 2151-2158
234. Khan, K. N., Stanfield, K. M., Harris, R. K., and Baron, D. A. (2001) Expression of cyclooxygenase-2 in the macula densa of human kidney in hypertension, congestive heart failure, and diabetic nephropathy. *Ren Fail* **23**, 321-330
235. Esmatjes, E., Levy, I., Gaya, J., and Rivera, F. (1987) Renal excretion of prostaglandin E2 and plasma renin activity in type I diabetes mellitus: relationship to normoglycemia achieved with artificial pancreas. *Diabetes Care* **10**, 428-431
236. Nasrallah, R., Xiong, H., and Hebert, R. L. (2006) Renal prostaglandin E2 receptor (EP) expression profile is altered in streptozotocin and B6-Ins2Akita type I-diabetic mice. *Am J Physiol Renal Physiol*
237. Nakagawa, T., Sato, W., Glushakova, O., Heinig, M., Clarke, T., Campbell-Thompson, M., Yuzawa, Y., Atkinson, M. A., Johnson, R. J.,

and Croker, B. (2007) Diabetic endothelial nitric oxide synthase knockout mice develop advanced diabetic nephropathy. *J Am Soc Nephrol* **18**, 539-550

238. Cao, X., Peterson, J. R., Wang, G., Anrather, J., Young, C. N., Guraju, M. R., Burmeister, M. A., Iadecola, C., and Davisson, R. L. (2012) Angiotensin II-dependent hypertension requires cyclooxygenase 1-derived prostaglandin E2 and EP1 receptor signaling in the subfornical organ of the brain. *Hypertension* **59**, 869-876