

OXIDATION OF CYTOSINE IN DNA

By

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Dissertation

Submitted to the Faculty of the
Graduate School of Vanderbilt University
In partial fulfillment of the requirements

for the degree of

DOCTOR OF PHILOSOPHY

in

Chemistry

May 2014

Nashville, TN

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To my family,
lovely and wonderful Mom, Dad, Brother
and to my beloved Sunshine

ACKNOWLEDGEMENTS

I would like to thankfully acknowledge my advisor Dr. Michael P. Stone, for giving me the greatest opportunity to perform research in his laboratory. For his guidance, help and the patience during this journey. He made me grow through this voyage to the scientist I am today. Thank you so much for encouragement, support and many great words of advice.

I am also very grateful to members of my PhD committee. Dr. Martin Egli, Dr. Carmelo Rizzo and Dr. Terry Lybrand you were so generous with time and advice both during and outside our formal committee meetings, and I greatly appreciate the different perspectives you have brought to this work and my scientific development.

Throughout my graduate career I had the great pleasure to work with a number of great collaborators. My NMR guru – Dr. Markus Voehler, Director of the Biomolecular NMR Facility, who not only taught me almost everything I know today about the biomolecular NMR, but always had the time and patience to help me with the experimental setup. Markus, I so appreciate your advice, expertise, and all discussions we had, I could not have done a huge part of this work without your unending help. I would also like to thank Dr. Don Stec for help and guidance in the NMR area.

I would like to thank Dr. Pradeep Pallan for guiding me in crystallization setup, data collection and refinement. Special thanks go to the Structural Biology Center team under the supervision of Dr. Andrzej Joachimiak at Argonne National Laboratory: Dr. Jurek Osipiuk, Dr. Robcio Jedrzejczak and special thanks to Dr. Bogi Nocek

for the help and mentoring during X-ray data collection and crystal structure refinement, as well as long and always exciting scientific brain-storming discussions.

This work would not have been accomplished without the tremendous help and engagement of Dr. Surajit Banerjee. I would like to acknowledge Dr. Barry Gold, Dr. Luis A. Marky, and members of their laboratories for providing samples and additional data results and analysis. I would like to thank Dr. Brandt Eichman and Sonja Brooks for helpful discussions during the development of this project. I would also like to thank Dr. Carmelo Rizzo for the opportunity to work in his laboratory and learn oligo synthesis techniques from his group members: Edward Hawkins, and Dr. Plamen Christov.

It has been a tremendous pleasure and fun to work with the Stone Lab during the past five years. A special thank goes to Ewa Kowal, who has been a great friend and wonderful colleague. I am so grateful to past and current members of the Stone group Kyle Brown, Surajit Banerjee, Hai Huang, Sarah Musser, Liang Li, Kallie Stavros, Drew Kellum for their help, advice and friendship. I also had the privilege to work with very talented undergraduate student – Patrick S. Donahue, thank you so much for your help and understanding. I would also like to acknowledge Ms. Sandra Ford for all the help, unending patience, support and friendship.

This work would not have been possible without the financial support of the National Institutes of Health, the Center in Molecular Toxicology, Center for Structural Biology, and Department of Chemistry at Vanderbilt University. I would like to thank the Vanderbilt Graduate School for providing travel grants to attend scientific conferences.

To all my friends, old and new, especially wszyscy Wolscy, Beierowie, pani Bożenka, ciocia Ania z Wiednia z Hansem, Agą oraz Alą, Klawisz, Ewa Karolak, Marta Szelązek, Ola Jankowiak, Agnieszka Bodzioch, Inka Krzyżanowska, Jacek Pecyna, Kasia z Hubertem, Ewa z Michałem, Becky and Kuba for all your friendships, greatest support and cheering me up constantly. Thank you so much for being such great friends.

I would not be here if it were not for the support of my family. Thank you Dad. Thank you Mom, I am so grateful for being so inspiring to me, for your love, support and encouragement. You have always been there for me. Marcin, you are the best brother I could have and anyone could hope for. Thank you so much for being so wonderful, I am so proud to be your sister. Thank you my in-laws, Mom Renia and Dad Antek, Piotr, for believing in me and support.

Most importantly, special thanks and love go to my Sunshine, to my husband. No one has been more important to me in this journey. Słonko, thank you so so much for your love, understanding, patience and support. For giving me all the strength. It would not be possible without you. You are the best husband, partner and friend ever.

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LIST OF ABBREVIATIONS

1D	One dimensional
2D	Two-dimensional
3D	Three-dimensional
5caC	5-carboxyl-2'-deoxycytidine
5fC	5-formyl-2'-deoxycytidine
5hmC	5-hydroxymethyl-2'-deoxycytidine
5mC	5-methyl-2'-deoxycytidine
Å	Angstrom
A	Adenine
AE-HPLC	Anionic-exchange high performance liquid chromatography
AID	Activation-induced deaminase
AlkB	Oxidative demethylase of N1-methyladenine or N3-methylcytosine DNA lesions
AP	Apurinic/apyrimidinic/abasic site
APOBEC	Apolipoprotein B mRNA editing enzyme, catalytic polypeptide-like
BER	Base excision repair
C	Cytosine
C	Celsius
CH ₃ CN	Acetonitrile
COSY	Correlation spectroscopy
D ₂ O	Deuterated/heavy water
dA	Deoxyadenosine

dC	Deoxycytidine
DDD	Dickerson Drew Dodecamer
DFT	Density functional theory
dG	Deoxyguanosine
DMSO	Dimethyl sulfoxide
DNA	Deoxyribonucleic acid
DNMT	DNA methyl transferase
DQF-COSY	Double-quantum filter correlation spectroscopy
DSC	Differential scanning calorimetry
dT	Deoxythymidine
eq.	Equation
EM	Electromagnetic
ES	Embryonic stem
EX1	Exchange regime 1
EX2	Exchange regime 2
G	Guanine
H ₂ O	Water
K	Kelvin
k_0	Rate of exchange from the open state in the absence of ammonia
k_B	Rate constant for exchange catalysis
kcal	kilocalorie
k_{cl}	Rate for base-pair closing

k_D	Bimolecular collision rate between the imino group and proton acceptor in the open state
k_{ex}	Chemical Exchange rate
k_{op}	Rate for base-pair opening
LSQ	Least squares quadratic
MAD	Multiple anomalous wavelength dispersion
MALDI-TOF	Matrix assisted laser desorption ionization time-of-flight mass spectrometry
MeCP2	Methyl CpG binding protein 2
mg	Miligram
MHz	Megahertz
min	Minute
mL	Mililiter
mM	Milimolar
mol	Mole
MPD	2-methyl-2,4-pentadienol
MR	Molecular replacement
mRNA	Messenger ribonucleic acid
Na ₂ EDTA	Sodium ethylenediaminetetraacetate
Na ₂ HPO ₄	Sodium phosphate dibasic
NaCl	Sodium chloride
NaH ₂ PO ₄	Sodium phosphate monobasic
NaN ₃	Sodium azide
NaPi	Sodium phosphate buffer

NE-CAT	Northeastern Collaborative Access Team
NMR	Nuclear Magnetic Resonance
NOE	Nuclear Overhauser Effect
NOESY	Nuclear Overhauser Effect Spectroscopy
PDB	Protein Data Bank
pK _a	Acid dissociation constant
Pol	Polymerase
ppm	Parts per million
rMD	Restrained molecular dynamic
rmsd	Root-mean-square deviation
RNA	Ribonucleic Acid
RP-HPLC	Reversed-phase high performance liquid chromatography
s	Second
SAD	Single anomalous wavelength dispersion
SAM	S-adenosylmethionine
s.d.	Standard deviation
T	Thymine
T ₁	Longitudinal relaxation
TDG	Thymine DNA glycosylase
TEOA	Triethanolamine
TET	Ten-eleven translocation enzyme
T _M	Melting temperature
TOCSY	Total correlation spectroscopy

U	Uracil
UNG	Uracil DNA glycosylase
UV	Ultraviolet
VIS	Visible
αK_{op}	Equilibrium constant for base pair opening
ΔC_p	Heat capacity
ΔG	Gibbs free energy
ΔH	Enthalpy
ΔS	Entropy
μg	Microgram
μL	Microliter
μM	Micromolar
σ_m	Electronic substituent constant

CHAPTER 1

INTRODUCTION

DNA structure

DNA, deoxyribonucleic acid, is the most important biomacromolecule as it carries genetic information for the development and functioning of all living organisms. It was first observed by the Swiss chemist Friedrich Miescher in the late 1800s (1). But nearly a century passed from that discovery until researchers unraveled the structure of the DNA molecule and realized its central importance to biology. In the decades following Miescher's discovery, other scientists in 1944, Avery, MacLeod and McCarty, published their discovery that DNA is the “fundamental unit” of genetic information, based on the results from the experiments on the pathogenicity of pneumococci (2). Soon after that, Phoebus Levene and Erwin Chargaff carried out a series of research efforts that revealed additional details about the DNA molecule, including its primary chemical components and the ways in which they are joined with one another (3-5). For many years, there was a debate among scientists, about the molecule that carries life's biological instructions. Most neglected DNA as too simple a molecule to play such a critical role. Instead, they argued that proteins were more likely to carry out this vital function because of their greater complexity and wider variety of forms. The importance

of DNA became clear in 1953 when James Watson and Francis Crick revealed the double helix structure of DNA (6), after studying Maurice Wilkins's and Rosalind Franklin's X-ray diffraction patterns of DNA fibers (5, 7, 8). A few years later, in an presentation in 1957, Francis Crick laid out the central dogma of molecular biology, which predicted the relationship between DNA, RNA, and proteins, and articulated the "adaptor hypothesis" (9). Final confirmation of the replication mechanism that was implied by the double-helical structure followed in 1958 through the Meselson–Stahl experiment (10). Further work by Crick and coworkers showed that the genetic code was based on non-overlapping triplets of bases, called codons, allowing Har Gobind Khorana, Robert W. Holley and Marshall Warren Nirenberg to decode the genetic code (11-13) These findings represent the birth of molecular biology.

Watson and Crick revealed the structure of DNA based on x-ray diffraction data of DNA fibers collected by Rosalind Franklin and Maurice Wilkins (4). This turned out to be the B type structure, and Watson and Crick proposed that a complete turn of the B-DNA molecule occurs once every ten bases. However, it was not until 1980 when a first crystal structure of this complete DNA turn was published (3). The sequence studied was a self-complementary dodecamer [5'-(CGCGAATTCGCG)-3']₂ and it seem to crystallize very well with "unusual ease and rapidity" compared to other DNA oligonucleotides.

Chemically, DNA is a polyanion at neutral pH. It usually appears as a double-stranded helix composed of two biopolymers that are made of nucleotides. Nucleotides comprise: purines Guanine (G) and Adenine (A), and pyrimidines Cytosine (C) and Thymine (T). The nitrogen bases are connected via deoxyribose ring and phosphate

backbone. The two strands are held together by hydrogen bonding between the bases as shown in Figure 1.1.

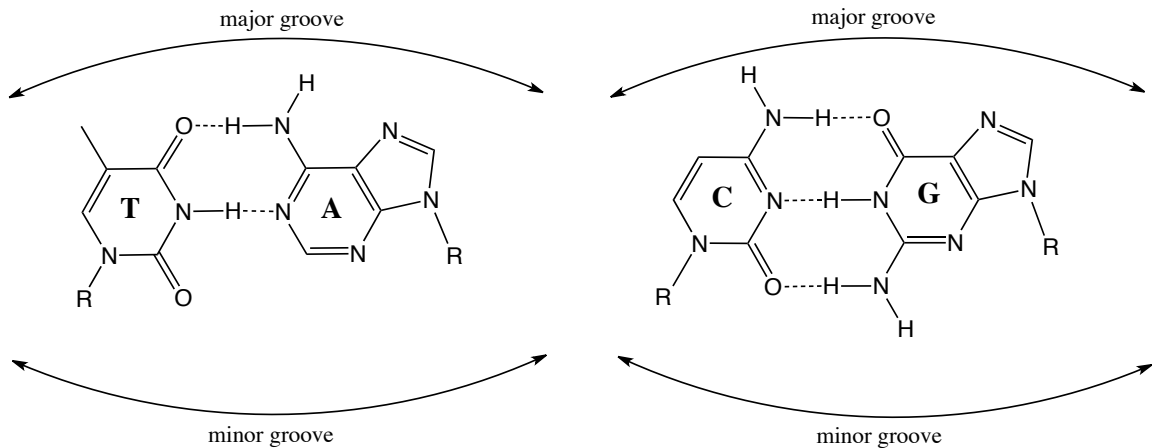


Figure 1.1. Watson-Crick pairing of A:T base pair (left) and a G:C base pair (right).

Bases fit into the double helical model when pyrimidines on one strand are always paired with purine on the other and vice versa. Chargaff's rule has its origin in the specific pairing of A with T and G with C. This pairs a keto base (G or T) with an amino base (C or A), and a purine with a pyrimidine. Two H-bonds can form between an A and T pair, and three can form between a G and C pair. These are the complementary base pairs. The base-pairing scheme immediately suggests a way to replicate and copy the genetic information.

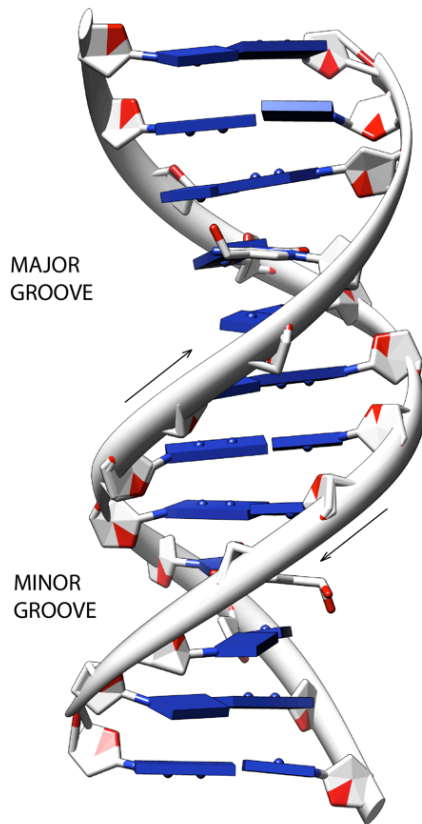


Figure 1.2. Antiparallel plectonemically coiled DNA strands. The arrows are pointed 5' to 3', and they illustrate the antiparallel nature of the duplex. The nucleotides arrayed in a 5' to 3' orientation on one strand align with complementary nucleotides in the 3' to 5' orientation of the opposite strand.

The two DNA strands are coiled around the same helical axis and are intertwined with themselves (which is referred to as a plectonemic coil). One consequence of this intertwining is that the two strands cannot be separated without the DNA rotating, one turn of the DNA for every "untwisting" of the two strands. The intertwined strands make two grooves of different widths, referred to as the major groove and the minor groove. The major groove is wider than the minor groove in DNA (Figure 1.2), and many sequence specific proteins interact in the major groove. The N7 and C6 groups of purines and the C4 and C5 groups of pyrimidines face into the major groove, where they can

make specific contacts with amino acids in DNA-binding proteins. Thus specific amino acids serve as H-bond donors and acceptors to form H-bonds with specific nucleotides in the DNA. H-bond donors and acceptors are also present in the minor groove, and indeed some proteins bind specifically in the minor groove.

Three different duplex have been described (14) (Figure 1.3). The most common form present in most DNA at neutral pH and physiological salt concentrations is B-form. That is the classic, right-handed double helical structure with a turn every 3.4 nm, such that the distance between two neighboring base pairs is 0.34 nm. Hence, there are about 10 pairs per turn. In a solution with higher salt concentrations or with alcohol added, the DNA structure may change to an A form, which is still right-handed, but every 2.3 nm makes a turn and there are 11 base pairs per turn. The A-form is adopted by RNA-DNA duplexes and RNA-RNA duplexes. Another DNA structure is called the Z form, because backbone phosphates are arranged in a zigzag pattern. Z DNA is left-handed. One turn spans 4.6 nm, comprising 12 base pairs. DNA molecules with alternating G-C sequences in alcohol or high salt solution tends to have such structure.

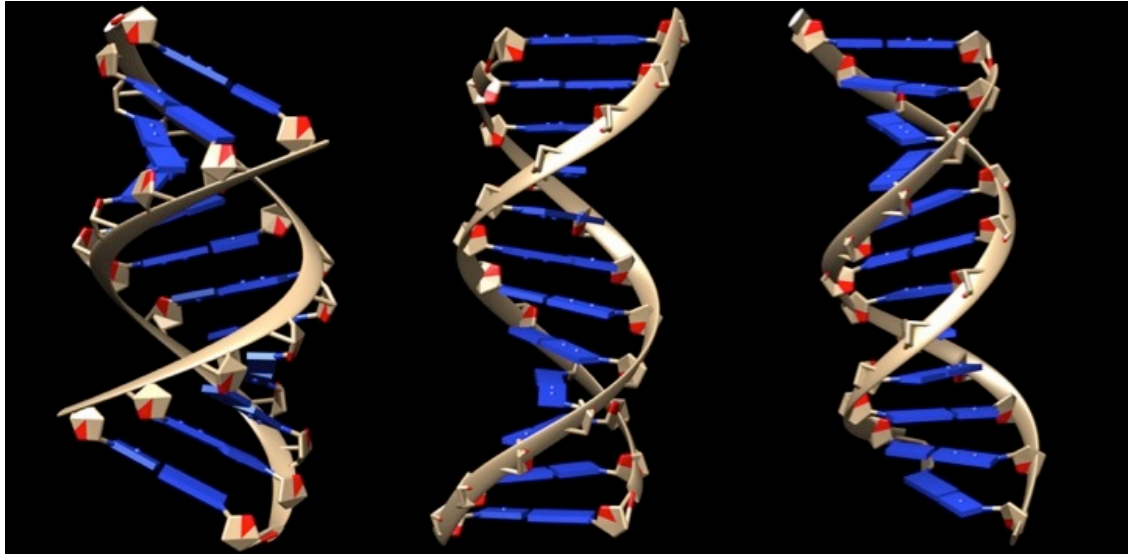


Figure 1.3. Models representing A-form (left), B-form (middle) and Z-form DNA (right).

Even classic B-DNA is not completely uniform in its structure. X-ray diffraction analysis of crystals of double-helical oligonucleotides shows that a given sequence will adopt a unique structure. These variations in B-DNA may differ in the propeller twist (between bases within a pair) to optimize base stacking, or in the three ways that two successive base pairs can be oriented relative to each other: twist, roll, or slide.

The strength of hydrogen bonds between base pairs contributes to the stability of the DNA helix. Appropriate geometrical correspondence of hydrogen bond donors and acceptors allows only for the complementary base to form stable base pairs. DNA with high CG content is more stable than DNA with low CG content, but hydrogen bonds do not stabilize the DNA themselves. Stabilization is mainly due to stacking interactions. Stacking interactions between bases are due to dispersion attractions, short-range exchange repulsions, and electrostatic interactions. Most favorable are GC stacking interactions with adjacent bases (15).

DNA in the form of the duplex is stable at room temperature, but the two strands separate above a melting point that is determined by the length of the molecule and CG content. Higher GC contents result in higher melting temperatures (15).

Epigenetic regulation of the genome

In the genome, DNA is constructed from the four nucleosides: 2'-deoxyadenosine (dA), 2'-deoxyguanosine (dG), thymidine (dT), and 2'-deoxycytidine (dC). These building blocks are assembled inside the DNA double helix (6, 15). In multicellular organisms all cells possess the same genetic content; however, cells might perform very different functions. For example, neuronal cells are designed to conduct electrical signals along the axons, muscle cell are involved in performing contractive motions. The same genetic material manifests itself in different functions of different cells, which is only possible because cells differ in the nature and number of active genes. During cellular development, cells switch unneeded genes on and off on their way to specialization. Chemically, long-term gene silencing is achieved by the methylation of base cytosines (dC) at position C5 in special places in the genome rich in CG repeats, called CpG islands (16, 17).

In duplex DNA, the C5 and C6 positions of cytosine are positioned in the major groove, held by Watson-Crick interactions with complementary guanines. The electrophilic character of the C6 position makes it a key target of modifying enzymes. For example, DNA methyltransferase (DNMTs) transiently modify C6 by attack of an active site cysteine. Methylation results from the concerted addition

of a methyl group derived from S-adenosylmethionine (SAM) to the C5 position (18, 19). The covalent intermediate breaks down, releasing the enzyme and generating genomic 5-methylcytosine (5mC). It was also shown that in the absence of SAM, DNMTs can also catalyze deamination at C4 (20, 21), or the addition of aldehydes to C5 (22), which raises the interesting question about the relevance of these non-classical functions *in vivo*.

The methylation pattern is a crucial part in the epigenetic information and a critical marker that distinguishes cells. The underlying 5-methylcytosine is often considered to be the fifth base of the genome. After fertilization and at certain points of the embryonic development, a large number of the methylation marks are erased (23, 24). This allows embryonic stem cells to differentiate into any possible specialized cell. In certain cases this genome-wide demethylation occurs without cell division and thus without the synthesis of new DNA. The methyl group then has to be actively removed by enzymes. The mechanism underlying this active demethylation is of high interest, because it is speculated that some cells might have the possibility to actively demethylate their genetic material in order to re-differentiate (25).

DNA Demethylation mechanisms

DNA methylation can be passive or active. Passive DNA demethylation occurs in dividing cells. While DNMT is responsible for active methylation during cell replication, its inhibition or malfunction will result in unmethylated cytosines and reduced overall methylation levels. Active DNA demethylation can occur in both dividing and nondividing cells. This process requires enzymatic reactions to revert

the 5mC back to unmodified cytosine (26-29). As of today, there is still no known mechanism in mammalian cells that can cleave the strong covalent C-C bond that connects cytosine to the methyl group. Instead, demethylation appears to involve a series of enzymatic reactions where 5mC is further modified, by either deamination or oxidation to a sequence of modifications that are recognized by the base excision repair (BER) pathway to remove the modified base, and replace it with unmodified cytosine. Several mechanisms of active DNA demethylation have been proposed (Figure 1.4):

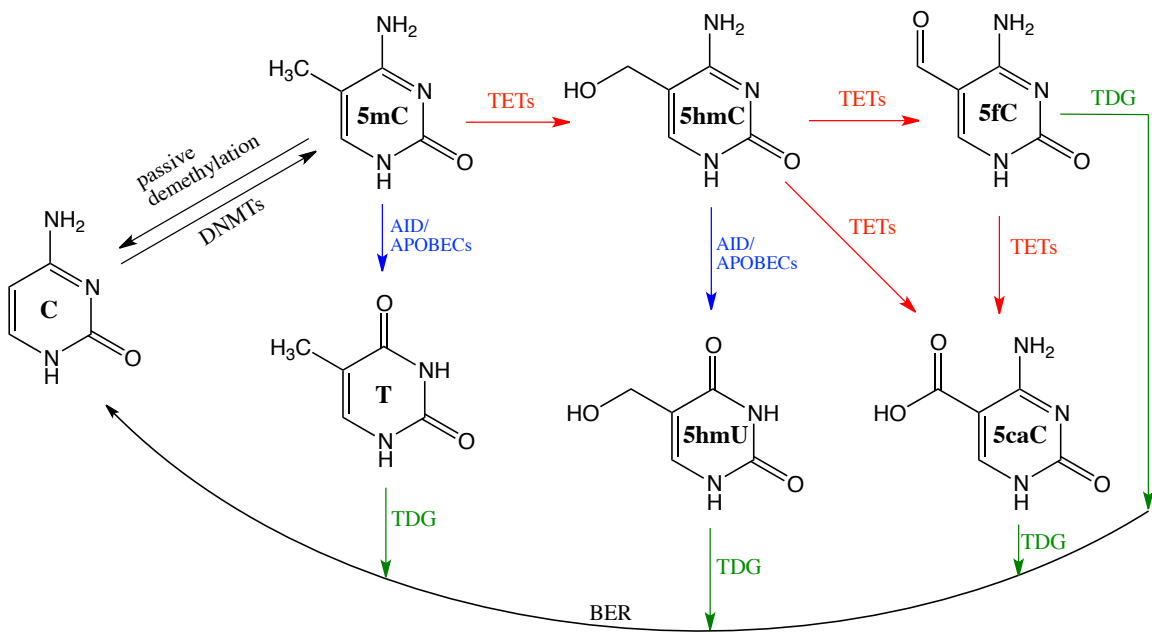


Figure 1.4. DNA demethylation pathways.

5-methylcytosine (5mC) can be chemically modified at two sites. At the C4 amino group of 5mC, which can be deaminated by AID/APOBEC (activation-induced cytidine deaminase/apolipoprotein B mRNA-editing enzyme complex), thereby

converting 5mC into thymine. This results in a G/T mismatch and induces the BER pathway to correct the base to G/C (30-32).

Recently, the search for alternative pathways of active demethylation led to discovery of 5-hydroxymethyl-cytosine (5hmC). Two independent research teams showed that mammalian genomic DNA contains not only 5mC, but also 5hmC, which is now considered to be the sixth base of the genome (33, 34). The 5hmC modified base is formed post-replicatively by enzymatic oxidation (35). The oxidation of 5mC is facilitated by the TET (ten eleven translocation) family of enzymes, which belong to the Fe(II)/ α -ketoglutarate-dependent oxygenase family that includes histone demethylases and the DNA damage repair enzyme AlkB (34, 36). TET enzymes were discovered based on homology to a trypanosome enzyme, which catalyzes oxidation of the exocyclic methyl group of thymine. The oxidation product of 5mC, 5hmC, was found in developed brains at the levels of 0.3-0.7%, which is approximately ten times lower than the natural abundance of 5mC (33, 37). Once 5hmC is formed, two separate mechanisms can convert 5hmC back to unmodified cytosine. In the first, AID/APOBEC can deaminate 5hmC to 5-(hydroxymethyl)-uracil (5hmU) which is a substrate for thymine DNA glycosylase (TDG) and excised via a base excision mechanism (BER) (38). In another pathway for 5hmC, iterative oxidation by TETs further oxidizes 5hmC to form 5-formylcytosine (5fC) and 5-carboxycytosine (5caC) (39). Both modifications are stably detectable intermediates in genomic DNA from embryonic stem (ES) cells (39, 40).

The function of 5hmC is still unclear. On the one hand, it may serve as an intermediate in DNA demethylation, but on the other it may also regulate gene

expression. In support of this theory, the conversion of 5mC to 5hmC impairs the binding of the repressive methyl-binding protein MeCP2 (41). Moreover, 5hmC was found *in vivo* in mammalian tissue, is tissue specific, and may play an important role in regulating DNA demethylation and gene expression.

In all the mentioned pathways of active DNA demethylation, the base excision repair pathway uses thymine DNA glycosylase (TDG) to excise the modified residue – thymine, 5hmU, 5fC, 5caC and replace them with an unmodified cytosine (42, 43). TDG is essential for DNA demethylation and is required for normal development.

DNA repair

Specific base pairing is essential for preserving the information content of the genome. However, the structural properties of DNA bases, and their pairing properties are often modified by reactions with toxins or endogenous metabolic products. As a result mismatches can be formed, and some of them may lead to certain types of cancer (44). Spontaneous deamination of 5mC at CpG sites in human DNA leads to C/T transition and therefore G/T mismatches. Mutations such as G/T mismatches can be very harmful to the genetic code, in terms of the information that is stored in the DNA, and epigenetic methylation patterns. If these mismatches are not repaired by the cellular repair mechanism known as base excision repair (BER), they can lead to DNA abnormalities and carcinogenesis (44). Base excision repair is the main mechanism by which cells correct various types of damaged DNA bases.

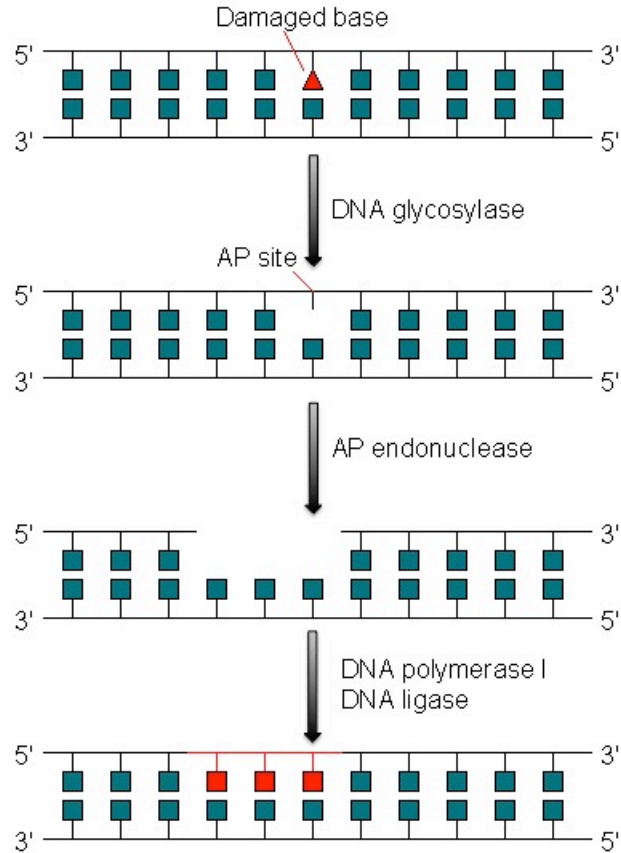


Figure 1.5. Base excision repair of DNA.

The first step in BER is the recognition of a wrong base by a glycosylase enzyme, which then excises the damaged base. This results in the formation of an apurinic or an apyrimidinic (AP) site. These AP sites are then removed by an AP endonuclease that causes nicks in the DNA as a result of the hydrolysis of the phosphodiester bond that is 5' to the AP site. These free ends are degraded by endonucleases, and the gap is then filled by a DNA polymerase enzyme (44, 45). DNA repair is then completed by ligation in the presence of ligases (Figure 1.5).

The recognition and detection of damage or modified bases in DNA by glycosylase is an essential step during DNA repair in all cells. This initial step

involves a specific repair enzyme finding its substrate in the midst of a vast excess of canonical bases that in many cases are structurally similar to the damaged base. A base-flipping repair protein may have to search through millions of base pairs to find a single point of damage. How these proteins efficiently locate base lesions has long been an interesting question. Assuming that repair protein first binds to DNA nonspecifically and then moves in one dimension, there are three potential damage-locating mechanisms: in the first, every base is actively flipped and inserted into the active site by the protein until the damage is located; in the second, repair protein selectively detects an unstable or non-Watson-Crick base pair that contains the damaged base; and in the third, a transiently extrahelical base lesion is captured by the repair protein (46). In the process of enzymatic base flipping, the DNA undergoes deformation, which is unfavorable in terms of energy. One of the most dramatic substrate conformational changes in site-specific nucleic acid recognition is the complete rotation of a base and its attached sugar from the base stack through either the major or minor groove. The unfavorable energetic events during the reaction include: nucleotide rotation, breaking of the Watson-Crick hydrogen bonds, and disruption of the base stacking and DNA bending. The binding repair enzyme has to pay for these costs through use of a favorable bending energy, which would increase in this case (Figure 1.6) (47).

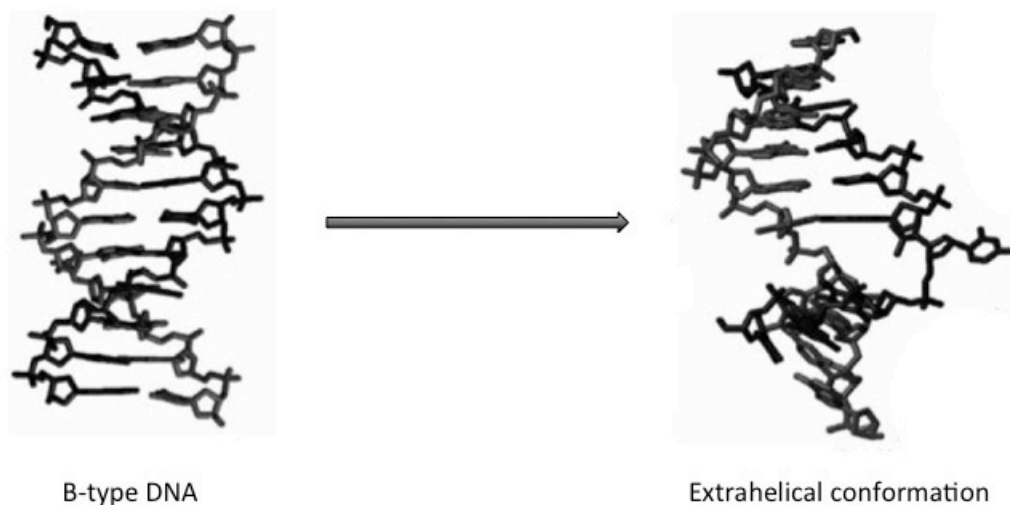


Figure 1.6. Base flipping occurring on the DNA molecule during the process of enzymatic flipping of the damage DNA base. (Figure adopted from (47)).

The most common experimental method to measure the flipping or base opening is imino exchange by nuclear magnetic resonance (NMR) spectroscopy. This technique indirectly measures the rates and the equilibrium for base pair opening and closing, by monitoring the exchange of the imino protons on G or T with solvent protons that have been magnetically labeled by selective inversion (48-52). The key assumption is that exchange occurs only in the open state. However, when the exchange of the imino proton is much faster than the opening of the base, then the latter becomes the rate-limiting step, and the observed exchange rate is equal to base pair opening; in other words, exchange occurs at each opening event. Since the exchanging imino protons are on either T or G, it is assumed, that within the whole base pair, these two bases will be flipping out from the stack in order to exchange their imino proton. However, as proposed based on computational studies, it is possible that for T exchange, the opposite adenine flips out, leading to exposure of the T imino proton without requiring T to leave the base stack. Likewise, imino proton exchange on G could occur

by flipping the opposite cytosine base (53). The increased flexibility or dynamic of a damaged or modified base pair could guide the repair enzyme to the site. The studies with uracil DNA glycosylase (UDG) have provided a precedent for this mechanism (54-56). In these studies, the DNA duplexes containing a single U:X or T:X base pair were used, where X was an adenine analog able to form a single hydrogen bond, double or triple hydrogen bonds with the opposite base. Progressive destabilization of the studied base pair by removal of a hydrogen bond would be expected to favor base pair opening and enzyme binding, because less binding energy would be required to open the destabilized base pair. The results with a U:X base pair indicated that base pairs with stronger hydrogen bonds inhibits uracil flipping into the enzyme binding site. The second result with a T:X base pair showed that a base pairs with larger equilibrium constants for opening lead to enhanced binding. The energetic correlations between the intrinsic stability and dynamics of the base pair and the enzyme binding to both uracil and thymine base pairs revealed that the initial damage recognition might rely on the intrinsic dynamic and physical properties of the base pair (54-56).

Another example of a damage-recognition mechanism involves selective detection of a non-Watson-Crick base pair that contains the damaged bases by the repair enzyme. An example of this type of mispairing is G pairing either with T or U in the Wobble base pair configuration (Figure 1.7). In this unique base pair G, which is normally capable of forming three hydrogen bonds, forms only two hydrogen bonds with the opposite T or U, due to a slight movement of G into the minor groove. This unusual structural feature is recognized by thymine DNA glycosylase and allows for efficient removal of T or U from the mismatched DNA (57).

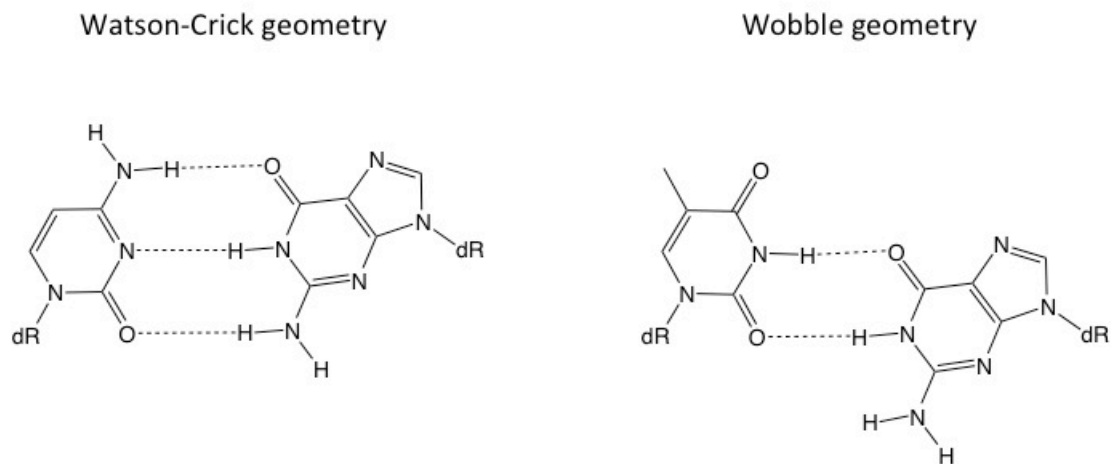


Figure 1.7. Different base pairing geometries.

Experimental methods for structural and dynamic analysis

Biological macromolecules play an important role in all cellular processes. Structural biology studies the relationship between the structure of biological macromolecules and their intrinsic function in order to understand chemical reactions, that are crucial to life. Two techniques are widely used for the structure determination of biological macromolecules at the level of distinguishing individual atoms: X-ray crystallography and nuclear magnetic resonance spectroscopy (NMR).

Nuclear Magnetic Resonance Spectroscopy

Nuclear Magnetic Resonance (NMR) spectroscopy has become a prominent tool in structural biology, as it has the ability to determine the atomic resolution structures of biomacromolecules in a native solution environment. NMR provides the means

for studying critical biological phenomena including nucleic acid and protein structure, and dynamics and it serves as a practical approach to drug design (8). The most basic physical principles that describe this biophysical technique as well as NMR experiments that are most relevant to structural biology are described below.

The NMR era started in the late 1930s when Rabi *et al.* measured the intrinsic magnetic moment of each spin (58). Bloch (59) and Purcell (60) then worked on transition effects caused by the resonance during the radiofrequency sweep in the static magnetic field, for which they were awarded with the Nobel Prize in 1962. Only three years later, Ernst and Anderson introduced pulsed excitation and signal averaging with the application of Fourier transformation of the time domain data. This allowed for averaging of multiple scans resulting in improved signals (61). That was the time when detection in two dimensions started, with pioneering contributions by Jeener (62), Earnst and studied by others (63). Kurt Wüthrich was first to apply these techniques to protein analysis (64), when he published the first NMR derived assignments of a protein leading to structure determination by NMR in 1982. Since then the analysis of biomacromolecules by NMR (65) at the 3D structural level has expanded, and it continues to be one of the major techniques in structural biology.

Nuclear magnetic resonance relies upon manipulation of the fundamental quantum mechanical properties of atomic nuclei to provide unique information on their electronic environment (9). The quantum mechanical property of interest is the nuclear spin angular momentum. Placement of the nuclei with the magnetically active spins (where the spin quantum number is $I = \frac{1}{2}$) within a static magnetic field of sufficient strength induces the spin angular momentum vector to rotate about an axis parallel

to the magnetic field (Figure 1.8) (10-12). The frequency of rotational motion, known as the resonance frequency, depends upon the electronic environment surrounding a given nucleus, i.e. whether it forms part of a covalent or non-covalent bond, or is in the proximity to other atoms. Subsequent excitation of these processing spins by an oscillating electromagnetic (EM) field, which transmits energies corresponding to their resonance frequencies, rotates the spins to the plane perpendicular to the static field (Figure 1.8). As they continue their precession in the transverse plane, a time-dependent signal that encodes the collective rotational frequencies of the spins is collected (Figure 1.8). Once processed by Fourier transformation, this signal provides a ‘frequency map’ that reflects the unique electronic surroundings of each spins (Figure 1.8).

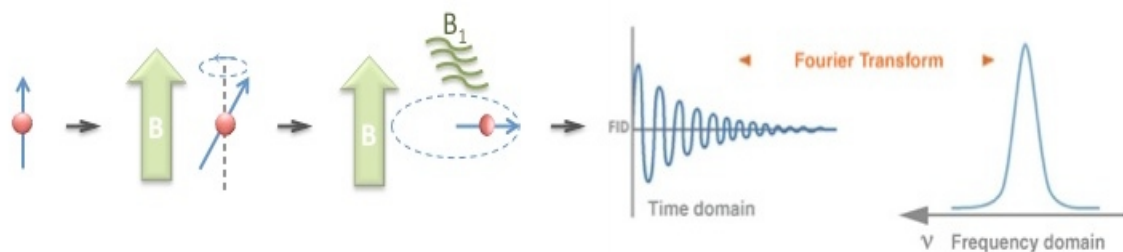


Figure 1.8. Nuclear magnetic resonance spectroscopy, and a basic outline of the NMR experiment. Nuclear spins (red sphere and blue arrow) undergo precessive motion at a characteristic resonance frequency when placed in a static magnetic field B (green thick arrow). Application of an oscillating magnetic pulse (B_1) rotates the spin vector to the perpendicular plane, where it continues its precessional motion and induces an oscillating voltage signal. Fourier transform of this time-domain signal generates a frequency spectrum with the higher intensity peak centered at the spin’s resonance frequency.

Without the presence of the excitatory EM field, transverse spin precession gradually returns to the original parallel precession extant prior to excitation. This return of excited spins to their ground energy state is known as ‘relaxation’, and provides a powerful

means for accessing global and local molecular motions, as the timescales of motion influence the rapidity with which spins relax (12, 13, 66, 67).

In terms of NMR, biomacromolecules are complex assemblies of nuclear spins. Although one dimensional ^1H NMR exploits the high natural abundance of the ^1H isotope in macromolecules, the large number of protons present in a macromolecule (as compared to a small molecule) often leads to extensive overlap within the resulting 1D ^1H NMR spectrum. Therefore, 2D NMR has been developed to overcome this problem.

One of the most powerful 2D NMR experiments for the solution structure determination of oligodeoxynucleotides is ^1H - ^1H Nuclear Overhauser Effect Spectroscopy (NOESY). This experiment uses the dipolar interaction of spins for correlation of protons located within approximately 5 Å apart in space. With the application of different mixing times ranging from 30 to 300 ms NOESY experiments can address spin diffusion effects. For the detection of non-exchangeable protons, deuterated solvent is used for NOESY in order to avoid excessive solvent signals. For the assignment of exchangeable protons, such as amino and imino protons, which relates to Watson-Crick base pairing, the NOESY spectrum can be obtained with the sample in H_2O solution.

The assignment of non-exchangeable protons in a DNA duplex can be performed on adjacent bases throughout the DNA strand (68). In the 5' to 3' direction, the intranucleotide, ^1H - ^1H distance between the aromatic H6 or H8 proton on the aromatic base purine or pyrimidine, respectively, and its own H1' deoxyribose sugar anomeric proton

allows for a cross-peak. This is followed by a cross peak of the H1' sugar proton to the H6 or H8 proton of its 3' neighboring base (Figure 1.9).

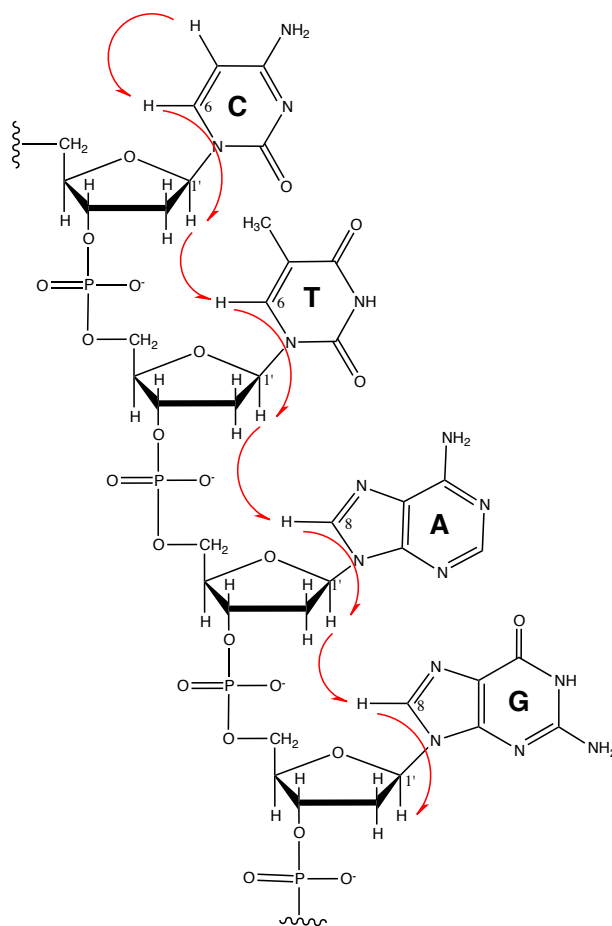


Figure 1.9. Sequential walk between aromatic H6/H8 aromatic base protons and anomeric H1' sugar protons.

In the same way, the inter-nucleotide distance between the H6 or H8 proton and its own H3' sugar proton allows for a cross peak, followed by an H3' sugar interaction with the 3'-neighboring H6 or H8 proton. This method of sequential assignment accomplished by “walking” through the DNA duplex in a 3' to 5' direction is known as a NOESY walk (Figure 1.9).

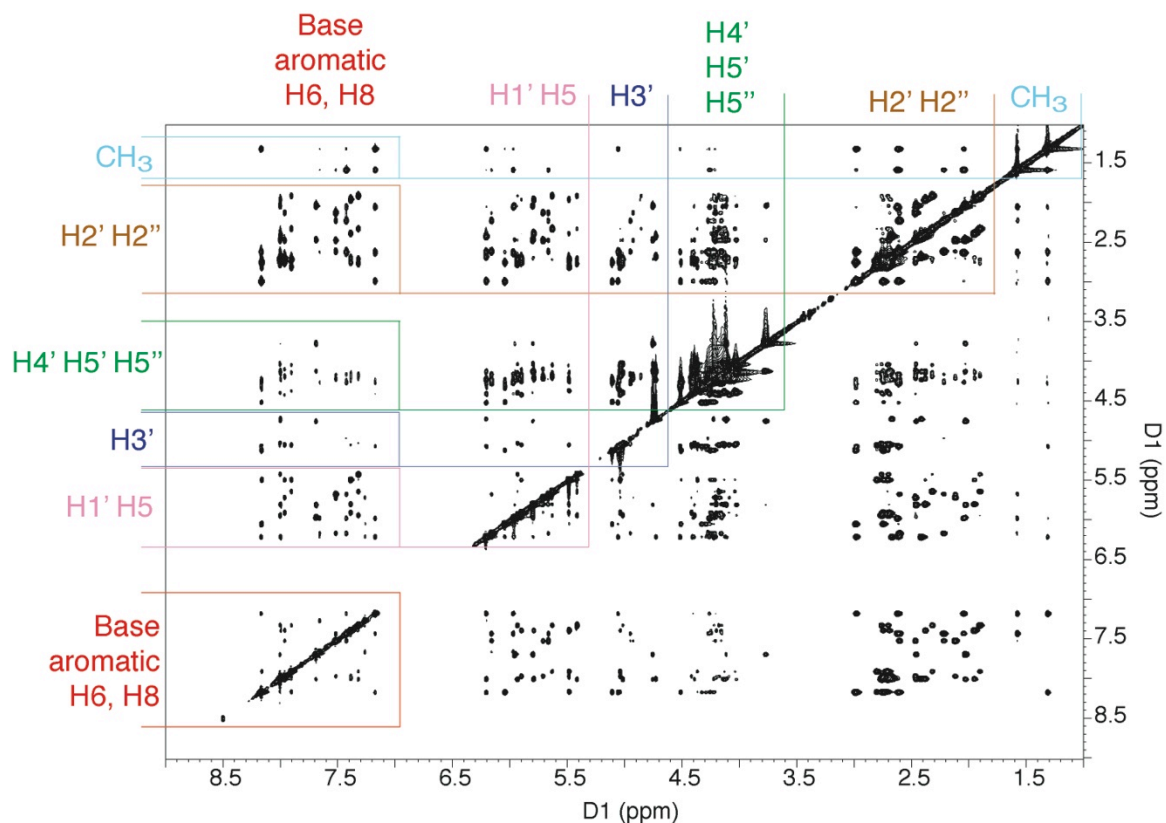


Figure 1.10. A typical ^1H - ^1H NOESY spectrum of an oligodeoxynucleotide. Interactions of NOE's are observed between peaks at the specific regions.

The NOESY spectrum (Figure 1.10) is very useful for observing DNA damage and modifications—induced disruption of the sequential connectivity. It also provides chemical shift information, as well as insight into the orientation of the modified base. A NOESY spectrum measured for a sample in H_2O solvent as the allows for detection of exchangeable protons. In this way one can detect imino protons, and their sequential intra-nucleotide connectivities between guanines and thymines throughout the whole DNA duplex (69). Interactions between imino protons as well as imino and amino protons provide information about Watson-Crick base pairing and stacking interaction in DNA.

Another widely used 2D experiment for determining molecular structures is correlation spectroscopy (COSY). It provides additional information by correlating the chemical shift of ^1H nuclei which are J -coupled to one another (70). With ^1H - ^1H COSY one can detect signals from geminal or vicinal hydrogens. In this way cross-peaks between cytosine H5 and H6 can be located, as well as the methyl protons of thymine. The Double Quantum Filtered COSY (DQF-COSY) (66, 67, 71) allows for partial cancellation of the diagonal peaks and elimination of strong signals coming from solvent protons that do not experience homonuclear J -coupling, as compared with magnitude COSY. In the latter, signals with small J -coupling constants and broad lines will show enormous diagonal signals, but only very small or vanishing cross-peaks. However, only DQF-COSY, where both cross and diagonal peak intensities depend on the size of the coupling constant, allows for detection of cross-peaks of the protons close in chemical shift and calculation of J -coupling constants. These values are useful in generating structural restraints such as the pucker of the deoxyribose sugar ring in DNA structural studies (72).

With Total Correlation Spectroscopy (TOCSY) one can detect the interactions of all protons with a spin system that are not directly connected by chemical bonds (73). As a result scalar coupling of a complete spin system can be determined. The scalar coupling range depends on the mixing time. For the short mixing time period, short range intraproton interactions are observed, and once the mixing time gets longer, then the correlation with longer distance protons can be detected. With the complete set of NMR 2D data a solution structure of DNA duplex can be obtained by applying restraints molecular dynamics (rMD) calculations.

X-ray crystallography

The era of 3D crystal structure determination of biomacromolecules started in the late 1950s, with the first crystal structure of sperm whale myoglobin by Kendrew and Perutz (74, 75), for which they were awarded the Nobel Prize in Chemistry in 1962. Thirty years later Richard Wing and co-authors reported the first single-crystal structure of more than a complete turn of right-handed B-DNA, based on the self-complementary dodecamer sequence [5'-(CGCGAATTCGCG)-3']₂ (76). The sequence crystallized in the $P2_12_12_1$ space group and the duplex was extensively characterized by Dickerson and Drew in 1981 (5-7). It was found to have an overall bend of 19° due to crystal packing forces, indicating intrusive flexibility of the DNA molecule. The duplex exhibited a rise of 3.4 Å per base pair, and there were 10.1 base pairs per DNA turn. Since then the Dickerson Drew Dodecamer (DDD) sequence has been extensively studied by crystallography, due to its ease of crystallization, and also by NMR. The crystal packing allows for a stable conformation of the DNA and well diffracting crystals. The DDD duplex has specific cation binding sites where Mg²⁺ or Ca²⁺ interact in the grooves of DNA. In addition a characteristic inner and outer spines of hydration are observed in the minor groove of the DDD (77). Cations and water molecules play an important role in crystal formation and stabilization of DNA duplex.

X-ray crystallography is used to determine the arrangement of atoms in a crystal. To analyze DNA by X-ray crystallography, it is necessary to grow crystals of a purified DNA sample. Crystals are formed as the conditions in a supersaturated solution slowly change. One of the methods for growing biomacromolecule crystals is the hanging drop vapor diffusion method, where a drop of protein solution is suspended over a reservoir

containing buffer and precipitant. Water diffuses from the drop to the solution leaving the drop with optimal crystal growth conditions. Crystallization conditions have to be optimized to obtain the desired crystals, the size of which is typically between 10-300 μm along each edge. Once crystals of good quality have been grown, of a good quality, they are looped out and flash frozen in liquid nitrogen and ready for X-ray analysis.

X-rays represent high-energy electromagnetic radiation and can be generated from accelerated electrons. Electrons are produced by a cathode and then accelerated to 99.9999% of the speed of light in a linear accelerator. The electrons are then injected into the booster synchrotron, where they are sent around an oval racetrack of electromagnets, providing further acceleration. Upon reaching the 99.9999% speed of light the electrons are injected into a storage ring. Once there, the electrons are focused into a beam and collimated with the sets of adjustable slits to ensure a parallel beam that is available for use in experimentation. X-rays are directed at the crystal when electrons diffract the X-rays, thus resulting in a diffraction pattern. Using the Fourier transformation these patterns can be converted into electron density maps. These maps show contour lines of electron density. Since electrons more or less surround atoms uniformly, it is possible to determine where atoms are located. Unfortunately since hydrogen has only one electron, it is difficult to map hydrogens. To get a three dimensional picture, the crystal is rotated while a computerized detector produces two-dimensional electron density maps for each angle of rotation. The third dimension comes from comparing the rotation of the crystal with the series of images. Computer programs use this method to come up with three dimensional spatial coordinates.

Using a synchrotron sources ensures a high intensity X-ray beam and allows shorter exposure times and a higher signal to noise ratios of the diffraction spots, which is required for crystallographic studies of biomacromolecules. With the application of CCD or Pilates detectors, the time of data collection of a complete data set from a single crystal is now rather short. As the diffraction spots become weaker at higher resolution, a compromise between increased resolution and decreased diffraction quality has to be made. When sufficiently high-resolution data are collected and processed, the unit cell dimensions, the crystal system, and the space group of the crystal can be determined.

A key factor in a crystal structure determination is the quantity of the collected data. A specific strategies have to be followed, that is dependent on the properties of the unit cell and the symmetry of the crystal. While exposed to the X-ray beam, the crystal is rotated and a series of diffraction patterns is collected. The spacing and intensities of the spots are the most significant information contained in the diffraction pattern. The spacing of the spots is dependent on the size and shape of the unit cell, and the intensity is determined by the amplitude of the diffracted waves and by the phase difference. Since the amplitudes can be recalculated, the information about the phase is lost; therefore it is referred as the phase problem. There are two methods most frequently used to overcome the phase problem: molecular replacement (MR), and multiple wavelength anomalous dispersion (MAD). The most common technique used is MR. When a homologous model is available, then coordinates and phases can be used with the experimental diffraction data to calculate an electron density map of the new molecule. In the next step the map is used to determine

the structure, which forms the basis for the refined set of structures. The MAD technique requires a single crystal to contain a heavy atom, which will cause anomalous scattering. The most common heavy metals used here are strontium, barium or selenium. With the MAD technique, the wavelengths of X-rays will be adjusted to strengthen the anomalous scattering. Thus, the heavy atoms can act as a reference marker and alter the intensity of spots in the diffraction pattern, which allows for the position of the heavy atoms to be determined and phases to be assigned.

With the solved phases and observed intensities of diffracted spots, Fourier transformation can be applied to obtain electron density map. This map forms a 3D contour into which the model can be built. Once the starting model is obtained, the phases can then be recalculated. In the next cycle, the calculated phase together with the original spot intensities can be used to rebuild and improve the electron density map and thus the new model. The improvement of the model from each cycle is called refinement, and is verified by comparing observed with calculated wave amplitudes, which is given by the R-factor. For DNA dodecamers, a satisfactory R-factor is between 0.15-0.25. Another validating factor, which evaluates the quality of the refinement is R-free, and it represents the difference between randomly selected 5-10% of the original data that are not used in the refinement cycle and the calculated data derived from the refined structure. The R-free should be below 20% for DNA structures.

Scope of this work

The main topic of this thesis concerns oxidation products of 5-methylcytosine (5mC), such as 5-hydroxymethylcytosine (5hmC), 5-formylcytosine (5fC) and 5-carboxylcytosine (5caC) with a specific focus on their structural and dynamic effects on the DNA duplex. Maiti and Drohat (78) showed that active demethylation of 5mC requires further oxidation of 5hmC to the downstream products 5fC and 5caC, allowing TDG to restore the epigenetically unmodified G:C pair. In their work they showed that both 5fC and 5caC are substrates for thymine DNA glycosylase (TDG), but not 5hmC. There are two major hypotheses in this research project. From studies with thymine DNA glycosylase (TDG), it was proposed that the enzyme recognizes a mismatched base of G/T or G/U mispairs in the DNA duplex, whereby the mismatch base pair forms a Wobble geometry (57). This unique structural geometry is recognized extrahelically by the enzyme, and results in excision of the mispaired base. Mechanistically, TDG employs an extrahelical base flipping mechanism (79-81) to position substrates into its active site for catalysis.

According to studies done by Stivers (47, 82), DNA stability and dynamics play a crucial role in the detection of damaged bases by a repair enzyme. The assumption is that the base pair with lower stability will have kinetically higher opening rates. In order to characterize the kinetics of base pair opening in nucleic acid duplexes, high-resolution proton exchange NMR spectroscopy has been used. The rates of exchange of imino protons with solvent protons were measured by magnetization transfer from

water protons for each DNA duplex as a function of the concentration of exchange catalyst.

The first goal of this research project is to determine the structures of 5hmC, 5fC and 5caC in the DDD sequence and to investigate the conformational perturbations of DNA induced by the presence of the modified bases, with a specific interest in base pairing and base stacking in the DNA double helix. The second goal is to study how the presence of the modifying groups in the major groove of DNA affects the stability of the double helix. The third and last goal is to determine the kinetics of base pair opening in the modified DNA double helix in order to understand dynamics around the modification site.

Chapter 2 describes the materials and methods. Chapter 3 reports the detailed thermodynamic analysis of 5hmC in duplex DNA, high-resolution crystal structure and dynamics of base pair opening. In chapter 4, the crystallographic results of 5fC in DDD are reported, as well as the kinetic characterization of base pair dynamics. Chapter 5 reveals the crystal structure of 5caC in DNA duplex, along with the kinetic results obtained by high-resolution NMR spectroscopy. Finally, chapter 6 provides conclusions driven from the work described in this dissertation and discusses future directions relevant to this project.

CHAPTER 2

MATERIALS AND METHODS

Oligodeoxynucleotide synthesis

Unmodified (DDD) and modified oligodeoxynucleotides with 5mC, 5fC or 5caC (DDD^m, DDD^f, and DDD^{ca}, respectively) were synthesized by Midland Certified Reagent Co. (Midland, TX) and purified by anion-exchange high performance liquid chromatography (AE-HPLC).

The 5hmC oligodeoxynucleotide (DDD^{hm}) was synthesized on an Expedite 8909 DNA synthesizer (PerSeptive Biosystems) on a 1- μ mol scale using the UltraMild line of phosphoramidites (ethylcyanide-protected 5-hydroxymethyl-dC, phenoxyacetyl-protected dA, 4-isopropyl-phenoxyacetyl-protected dG, acetyl-protected dC and dT phosphoramidites) and solid supports from Glen Research (Sterling, VA). The manufacturer's standard synthesis protocol was followed except in the case of the incorporation of the modified phosphoramidite, which was accomplished manually, off-line. At this point, the column was removed from the instrument and sealed with two syringes, one of which contained 250-300 μ L of the manufacturer's 1H-tetrazole activator solution (1.9-4.0% in CH₃CN, v/v) and the other contained 250 μ L of the modified phosphoramidite solution (15 mg in anhydrous CH₃CN). The 1H-tetrazole and the phosphoramidite solutions were sequentially drawn through the column (1H-tetrazole first), and this procedure was repeated periodically over 30 min. After this

time, the column was washed with manufacturer's grade anhydrous CH₃CN and returned to the instrument for the capping, oxidation, and detritylation steps. After the synthesis the deprotection reaction was run with 30% ammonium hydroxide for 17 hours at 75 °C.

Oligodeoxynucleotide purification

The unmodified and modified oligodeoxynucleotides were purified by semi-preparative reversed-phase high performance liquid chromatography (RP-HPLC). The purification was conducted on a Beckman HPLC system (32 Karat software) with a diode array UV detector measuring at 260 nm. The semi-preparative column was used (Atlantis by Waters, C18, 5 μm, 250 mm x 10.0 mm), and was equilibrated either with 30 mM phosphate buffer at pH 7.0 (for DDD, DDD^m, DDD^{hm}, and DDD^{ca}) or 0.1 M ammonium formate at pH 6.5 (for DDD^f) and acetonitrile. The purification method was set up with the following gradient: 1-15% acetonitrile over 20 min., 15-80% acetonitrile over 5 min., and then to 1% acetonitrile over 5 min., with a flow rate of 4.5 mL/min. The collected fractions were incubated at -80 °C followed by lyophilization. Figure 2.1 shows an example of RP-HPLC chromatogram of DDD^{hm} after purification. Dried oligonucleotides were resuspended in 0.5 mL of deionized water and desalted on a G-25 Sephadex column.

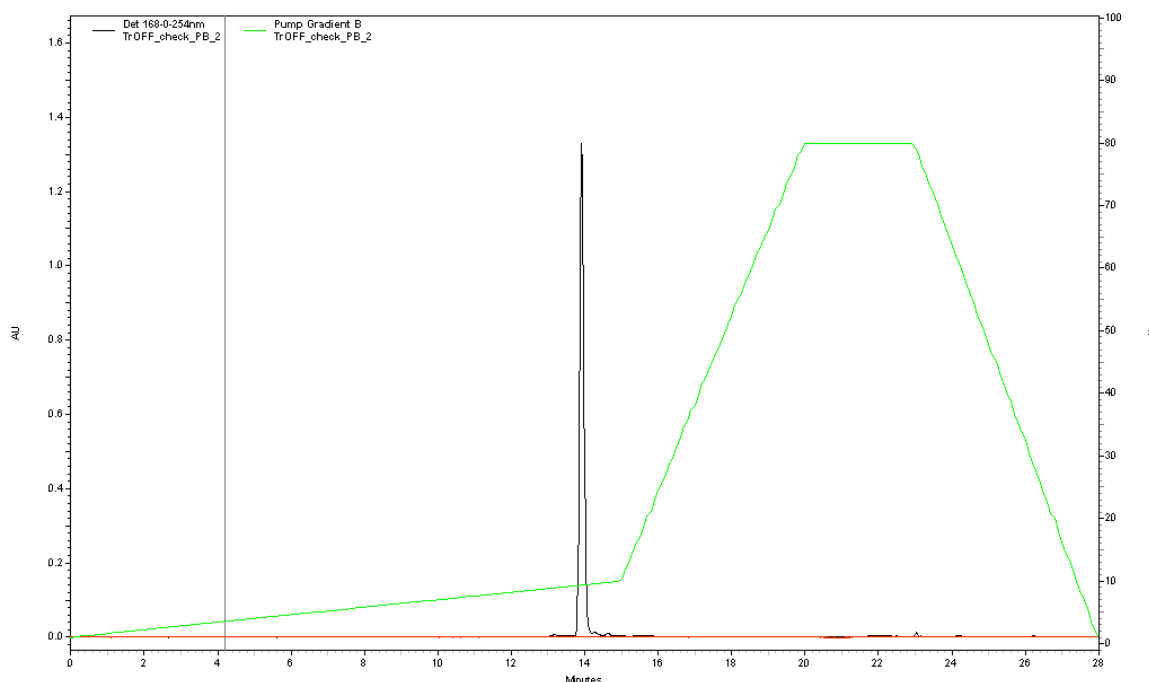


Figure 2.1. Example of an RP-HPLC chromatogram showing pure oligodeoxynucleotide.

Matrix-assisted laser desorption/ionization mass spectrometry

The purity of the oligonucleotides was confirmed by MALDI-TOF mass spectrometry. The mass analysis was performed using a Voyager-DE (PerSeptive Biosystems, Inc., Foster City, CA) spectrometer. Samples were suspended in a matrix containing 0.5 M 3-hydroxypicolinic acid in 1:1 CH₃CN:H₂O, 0.1 M ammonium citrate and spotted onto sample plates. Mass spectra were recorded in a negative reflector and averaged from 512 scans. Calculated mass for DDD [M-H]⁻ *m/z* 3646.4, found *m/z* 3647.8; for DDD^m calculated mass of [M-H]⁻ *m/z* 3660.5, found 3663.4; for DDD^{hm} calculated mass of [M-H]⁻ *m/z* 3675.5, found 3679.7; for DDD^f calculated mass of [M-H]⁻ *m/z* 3674.4, found 3673.2; for DDD^{ca} calculated mass of [M-H]⁻ *m/z* 3690.4, found 3693.1.

Sample preparation

The lyophilized and desalted oligodeoxynucleotides were dissolved in buffer containing 10 mM Na₂HPO₄, 100 mM NaCl, 50 μM Na₂EDTA at pH 7.0, and annealed by heating the solution to 85 °C for 15 min, and allowing it to slowly cool to room temperature. Duplex concentrations were determined by UV absorbance using extinction coefficients determined at 260 nm.

Thermodynamic measurements

Temperature-Dependent UV Spectroscopy (UV Melting Curves).

UV melting curves (absorption versus temperature profiles) for the helix–coil transition of DDD, DDD^f and DDD^{ca} duplexes were obtained with a thermoelectrically controlled Cary 100 Bio UV-VIS spectrophotometer (Varian, Inc. Palo Alto, CA), interfaced to a PC computer with Cary WinUV Thermal Application (v. 2.0) for data acquisition and analysis. The samples containing 0.20 A₂₆₀ unit of duplex were dissolved in 1 mL of buffer containing 10 mM Na₂HPO₄/NaH₂PO₄ at pH 7.0, 100 mM NaCl, 50 μM Na₂EDTA. The UV absorbance measurements were performed for unmodified and modified duplexes, at 260 nm, at 1 min intervals with a 1 °C/min temperature gradient. The temperature was cycled between 10 and 85 °C. The first derivative of the melting curve was used to establish the T_M values.

For the samples of DDD, DDD^m and DDD^{hm} a complete thermodynamic analysis was performed in collaboration with the Marky Lab from the University of Nebraska

Medical Center. The detailed methodology is described below. The UV melting experiments were performed on Lambda-10 Perkin-Elmer or AVIV spectrophotometers, equipped with a thermoelectrically controlled sample holder. The UV absorbances at 260 nm and 275 nm were measured in the temperature range of 0–90 °C with a heating rate of 0.6 °C /min. The melting temperature, T_M , was determined from the midpoints of the UV melting curves (Figure 2.2). The van't Hoff enthalpy, ΔH_{vH} , was determined from the slopes of the linear plots of the experimentally measured values of $1/T_M$ vs. $\ln(C_T)$, according to the relationship:

$$\frac{1}{T_M} = \left(\frac{R}{\Delta H_{vH}} \right) \ln C_T + \left(\frac{\Delta S_{vH}}{\Delta H_{vH}} \right) \quad (1)$$

where C_T is the total concentration of DNA strands. The nature of complex formation of each oligonucleotide was studied by performing UV melts as a function of their total strand concentration (3–330 μ M). To understand the molecular changes accompanying the unfolding transitions, the differential binding of counterions (Δn_{Na^+}) was determined by performing UV melts in the presence of salt (10–200 mM NaCl). This parameter is calculated using the following thermodynamic relationship (83, 84):

$$\Delta n_{Na^+} = 0.483 \left(\frac{\Delta H_{cal}}{RT_M^2} \right) \left(\frac{\partial T_M}{\partial \log [Na^+]} \right) \quad (2)$$

where R is the molar gas constant. The first bracketed term in is experimentally determined from differential scanning calorimetric experiments, while the second bracketed term is determined by measuring the T_M at different NaCl concentrations (83).

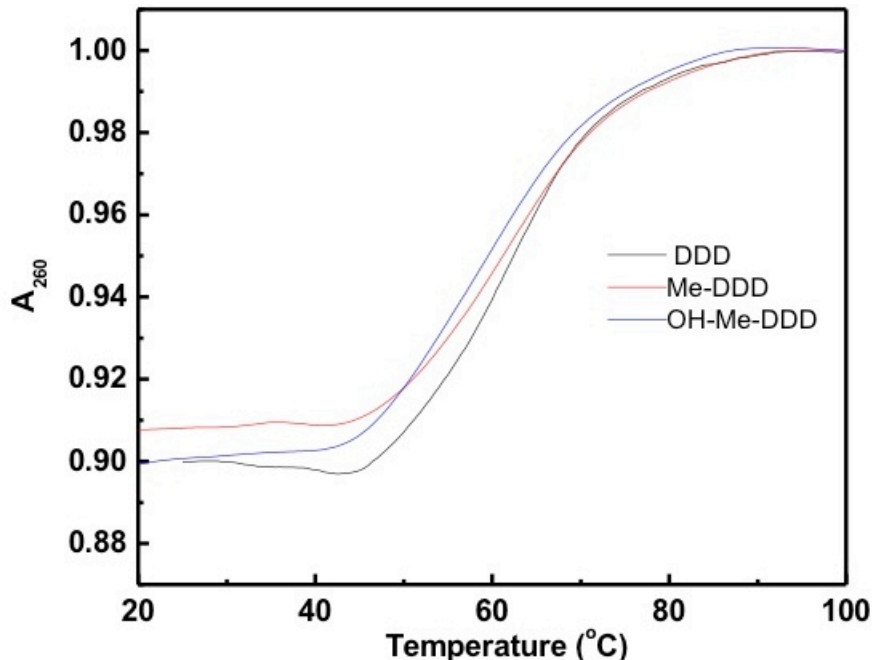


Figure 2.2. The example of the UV melting curve.

Differential Scanning Calorimetry (DSC).

The changes in heat capacity (ΔC_p) as a function of temperature were determined with a VP-DSC differential scanning calorimeter (MicroCal, LLC, Northampton, MA, USA). In a typical DSC experiment, ~ 200 μM on total strand concentration of the oligonucleotide solution in the sample cell (0.506 mL cell volume) was scanned from 0–90 °C at a rate of 45 °C/h with buffer in the reference cell. A buffer versus buffer scan was also done under similar conditions and subtracted from the sample runs. The resulting thermograms were plotted as ΔC_p versus T profiles using the Origin 7.0 software provided with the instrument. Analysis of the integrated plots of the anomalous ΔC_p versus temperature curves ($\int \Delta C_p dT$) and normalization for the number of moles, yields the molar calorimetric enthalpy (ΔH_{cal}). The molar calorimetric entropy (ΔS_{cal})

was calculated from integration of the $\Delta C_p/T$ versus T curves [$\int(\Delta C_p/T)dT$]. The Gibbs equation was used to calculate the Gibbs free energy at 20 °C:

$$\Delta G_{(20)} = \Delta H_{\text{cal}} - T\Delta S_{\text{cal}} \quad (3)$$

Furthermore, heat capacity measurements for the unfolding of each duplex were measured indirectly in DSC experiments at different salt concentrations, this information is obtained from the slope of the ΔH_{cal} versus T_M plots.

Circular dichroism (CD) Spectroscopy.

CD measurements were conducted on an Aviv (model 202SF) CD spectrometer (Lakewood, NJ, USA). The spectrum of each duplex was obtained using a strain-free 1 cm quartz cell at low temperatures to ensure 100% duplex formation. Typically, 1 OD of a duplex sample was dissolved in 1 mL of 10 mM sodium phosphate buffer at pH 7.0. The reported spectra correspond to an average of three scans from 220 to 350 nm with a wavelength step of 1 nm.

Nuclear Magnetic Resonance Spectroscopy

The modified and unmodified samples were diluted to a duplex concentration of 0.25 mM in 180 μL of 10 mM NaH_2PO_4 , 100 mM NaCl , 0.011 M NaN_3 , 50 μM Na_2EDTA buffer. The samples were exchanged with D_2O and dissolved in 180 μL of 99.99% D_2O to observe nonexchangeable protons in the spectra. For the observation

of exchangeable protons, the samples were dissolved in 180 μL of 9:1 $\text{H}_2\text{O}:\text{D}_2\text{O}$. The NOESY and DQF-COSY spectra of samples in D_2O were collected at 15 $^\circ\text{C}$ on a Bruker Avance spectrometer operating at 900 MHz with 5 mm CPTCI cryoprobe. For assignment of exchangeable protons, NOESY experiments with mixing times of 150, 200 and 250 ms and TPPI quadrature detection was conducted. These data were recorded with 2048 real data points in the t_2 dimension and 1024 data points in the t_1 dimension. The relaxation delay was 2.0 s. The data in the t_1 and t_2 dimension were zero-filled to give the matrix of $2\text{K}\times 2\text{K}$ real points. The NMR spectra for the exchangeable protons were recorded at 5, 15, 25, 35, 45, 55 and 65 $^\circ\text{C}$. The NOESY spectra of unmodified and modified samples in H_2O were collected at 5 $^\circ\text{C}$ with 70 and 250 ms mixing times and relaxation delay of 2.0 s. Water suppression was achieved by a gradient Watergate pulse sequence. Chemical shifts were referenced to water. NMR data were processed using TOPSPIN software (2.0.b.6, Bruker Biospin Inc., Billerica, MA).

Imino Exchange Measurements

Characteristic of base pair opening processes in DNA relies upon the exchange properties of imino protons, i.e. N1H in guanines and N3H in thymines. The opening of individual base pairs in DNA is generally characterized from the exchange of imino protons (Figure 2.3) with solvent protons. In the native DNA double helix, the imino protons are not accessible to solvent due to their location in the center of the structure and their participation in hydrogen bonds. For the exchange to occur, the base pairs must open up. In this opening reaction, the hydrogen bond holding the imino proton

is transiently broken and the proton is moved into a solvent accessible state, where it can be transferred to acceptors present in solution (48).

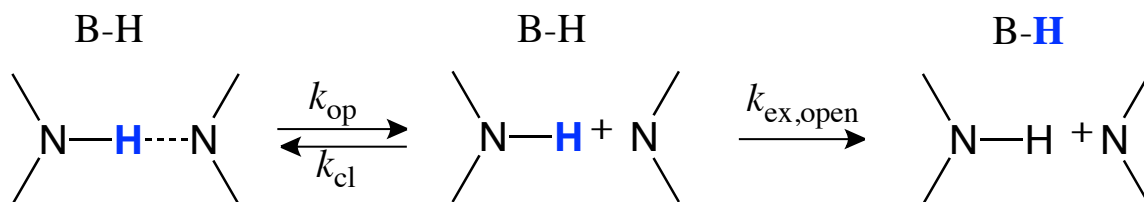


Figure 2.3. Base pair opening process and imino proton exchange.

The exchange rate observed experimentally depends upon the kinetic parameters of the opening reaction as:

$$k_{ex} = \frac{k_{op} \cdot k_{ex,open}}{k_{cl} + k_{ex,open}} \quad (4)$$

where k_{op} and k_{cl} are the rates of opening and closing of the base pair, and $k_{ex,open}$ is the rate of exchange from the open state. To determine the opening and closing rates, one varies the rate of exchange from the open state, $k_{ex,open}$, by adding to the DNA solution increasing concentrations of a proton acceptor. In the present work ammonia was used as a proton acceptor due to its small size and lack of charge, and to minimize catalysis due to presence of OH^- ions (85). Increasing concentrations of ammonia base catalyst were obtained by adding to the sample small aliquots of an ammonia stock solution at the desired pH. The DNA samples contained 1mM TEOA (triethanolamine), which was used to monitor the pH of the sample directly in the NMR tube during ammonia titration. This was done by measuring the difference between the chemical

shifts of the resonances of the two methylene groups of TEOA, according to previous reports (50). The pH was measured after each ammonia titration, and its experimental range was found to be 8.8-9.0. A pK_a value 9.218 for ammonia at 15 °C was used. The final concentration of the active form of the ammonia base catalyst was calculated from the total ammonia concentration (c_0) and pH as follows:

$$[B] = c_0 \cdot \frac{10^{-pK}}{10^{-pH} + 10^{-pK}} \quad (5)$$

The rate of exchange of the imino proton from the open state of the base pair depends on the concentration of proton acceptor B:

$$k_{ex,open} = k_0 + \alpha k_B [B] \quad (6)$$

where k_0 is the rate of exchange from the open state in the absence of ammonia, k_B is the rate constant for transfer of the imino proton to base catalyst in isolated nucleotides, and α is a factor that accounts for any differences in the rate of proton transfer between isolated nucleotides and open DNA base pairs, e.g., restricted accessibility of the proton acceptor to the imino proton in the open base. Previous investigations have shown that, for ammonia base, α is close to unity (85). k_B is the rate constant for exchange catalysis and is calculated from (50):

$$k_B = \frac{k_D}{1 + 10^{pK_a^{Nu} - pK_a^B}} \quad (7)$$

in which k_D is the bimolecular collision rate between the imino group and proton acceptor in the open state ($1.0 \times 10^9 \text{ M}^{-1}\text{s}^{-1}$ for ammonia at 15°C), pK_a^{Nu} and pK_a^{B} are the pK_a values for the imino proton of interest and the ammonia base catalyst, respectively. The final equation for the exchange rate is obtained by inserting eq 6 into eq 4 (with $\alpha = 1$):

$$k_{ex} = \frac{k_{op}(k_0 + k_B[B])}{k_{cl} + k_0 + k_B[B]} \quad (8)$$

Two kinetic regimes can be distinguished depending on how the rate of exchange from the open state compares with the rate of closing of the base pair (50). The EX2 regime occurs when the concentration of proton acceptor is low such that $k_{ex,open} \ll k_{cl}$. In this case, the observed rate of exchange is proportional to the rate of exchange from the open state (eq 4), and thus to the concentration of proton acceptor B:

$$k_{ex} = \frac{k_{op}}{k_{cl}} \cdot k_{ex,open} = K_{op} \cdot (k_0 + k_B[B]) \quad (9)$$

where $K_{op} = k_{op}/k_{cl}$ is the equilibrium constant for opening of the base that contains the imino proton. The other regime, called EX1 regime, occurs at high concentrations of proton acceptor, when $k_{ex,open} \gg k_{cl}$. In this regime, the exchange occurs in each opening event and $k_{ex} = k_{op}$. For the determination of the base pair opening (k_{op}), imino exchange rates are measured as a function of titrated ammonia base catalyst and are fit to equation 8.

For the imino exchange experiments NMR data were collected at 15 °C at 500 MHz using a Bruker AV-III spectrometer equipped with 5 mm CPQCI probe. The samples were dissolved in 180 mL of 90% H₂O, 10% D₂O solution containing 10 mM phosphate buffer, 100 mM NaCl, 0.05 mM EDTA, 0.011 M NaN₃, 1 mM TEOA (pH 8.9). Magnetization transfer from water to the imino protons was followed by observation of the imino protons after a variable mixing time. For selective spin inversion of the water protons, a 2 ms, a 180° Sinc pulse with 1000 points was used. To minimize effects of radiation damping during the mixing time, a 0.1 G·cm⁻¹ gradient was used. Water suppression was achieved by a binominal 1-1 echo sequence, jump and return, with flanking 1 ms smooth square shape gradients, 15 G·cm⁻¹. Sixteen values for the variable delay ranging from 1 ms to 15 s were used for each experiment. All data were processed and analyzed in TOPSPIN.

The exchange rates were calculated from the following equation:

$$\frac{I_z(t_{\text{mix}})}{I_{z,\text{eq}}} = 1 + Ek_{\text{ex}}(e^{-R_{1i}t_{\text{mix}}} - e^{-R_{1w}t_{\text{mix}}}) \quad (10)$$

in which $I_z(t_{\text{mix}})$ and $I_{z,\text{eq}}$ are intensities of the imino proton peaks at a given value of t_{mix} and at equilibrium, respectively; k_{ex} is the chemical exchange rate, R_{1w} is the longitudinal relaxation rate of water (3.15 sec as determined separately under the same conditions), R_{1i} is the sum of the imino proton relaxation rate and k_{ex} , and E is the efficiency of water inversion (value of -2 as given elsewhere (86, 87)).

The data analysis and fits were performed using PRISM software (v. 6.0b, GraphPad Software, Inc.).

Gaussian calculations

GAUSSIAN09 (88) was used for the calculation of bond length, angle and torsion angle values for non-standard residues (i.e. 5hmC, 5fC, 5caC). The restrained electrostatic potential charges were calculated using the B3LYP Density Functional Theory (DFT) method with a 6-31G* basis set.

X-ray Crystallography

Crystallization of DNA

Crystals of the DDD^{hm}, or DDD^f or DDD^{ca}, were grown at 18 °C by the hanging-drop vapor-diffusion method, using the Nucleic Acid Mini-screen (Hampton Research, Aliso Viejo, CA). Droplets of 2 µL volume containing 1.2 mM modified DNA duplex, and precipitating solution, were equilibrated against 0.75 µL of 35% MPD. Crystals grew within 8-16 days. Precise crystallization conditions and solution compositions of particular oligodeoxynucleotides are summarized in Table 2.1. Single crystals from each DNA sample were mounted in nylon loops and flash-frozen in liquid nitrogen.

Condition	DDD^{hm}	DDD^f	DDD^{ca}
pH	7.0	6.0	6.0
Buffer	40 mM Na Cacodylate	40 mM Na Cacodylate	40 mM Na Cacodylate
Salts	80 mM NaCl, 20 mM MgCl ₂	80 mM NaCl	80 mM SrCl ₂
Additives	12 mM spermine 4HCl	12 mM spermine 4HCl	12 mM spermine 4HCl
2-methyl-2,4-pentadienol (MPD)	10 % (v/v)	10 % (v/v)	10 % (v/v)

Table 2.1. Crystallization conditions.

X-ray Diffraction Data Collection and Processing

For the DDD^{hm} crystal diffraction data were collected on the 19-ID beamline of the Structural Biology Center at the Advanced Photon Source of Argonne National Laboratory (Argonne, IL) (89). The wavelength was 0.9794 Å. Initial indexing and scaling of recorded diffraction images, together with further reflection merging, were done using HKL3000 (90). In order to ensure high completeness of data two separate passes were collected.

For the DDD^f and DDD^{ca} crystals, data were collected on the 24-IDE and 24-IDC beamlines, respectively, of the Northeastern Collaborative Access Team (NE-CAT) at the Advanced Photon Source of Argonne National Laboratory (Argonne, IL). The wavelength was 0.97920 Å. Initial indexing and scaling of recorded diffraction images, together with further reflection merging, were done using XDS (91, 92), and SCALA (93) in the CCP4 (94) suite as part of the RAPD data-collection strategy at NE-CAT. Data collection details are shown in Table 2.2:

Parameter	DDD ^{hm}	DDD ^f	DDD ^{ca}
Crystal data			
Space group	<i>P</i> 2 ₁ 2 ₁ 2 ₁	<i>P</i> 2 ₁ 2 ₁ 2 ₁	<i>P</i> 2 ₁ 2 ₁ 2 ₁
Unit cell			
<i>a</i> [Å]	25.6	25.3	24.2
<i>b</i> [Å]	41.3	41.5	41.3
<i>c</i> [Å]	64.3	65.6	66.4
Data collection			
Resolution range [Å]	40-1.00	35-1.74	26-1.95
Reflections			
Observed	338726	8305	20462
unique	37637	3296	5113
Completeness (%)	99.6	99.4	99.5
In the outer shell (%)	98.5	94.8	97.5
R _{merge} ^a	0.044	0.034	0.045
In the outer shell ^b	0.979	0.747	0.619
I/σI	52	20	16
In the outer shell ^b	1.65	2.64	2.82
Structure refinement			
Resolution range [Å]	40-1.02	35-1.74	26-1.09
R-work	0.15	0.23	0.22
R-free	0.17	0.29	0.26
RMS deviation			
Bond lengths [Å]	0.018	0.009	0.008
Angle distances [Å]	1.015	1.747	2.209
Number of ions	1 Mg ²⁺	-	-
Number of ligands	3	-	-
Number of water molecules	178	56	13

Table 2.2. Crystal Data, Data Collection, and Refinement Statistics for DDD^{hm}, DDD^f and DDD^{ca}.

^a $R_{\text{merge}} = \frac{\sum_{hkl} \sum_i |I_i - \langle I \rangle|}{\sum_{hkl} \sum_l \langle I \rangle}$, where I_i is the intensity for the i_{th} measurement of an equivalent reflection with indices h , k , and l .

^b Numbers in parentheses are values for the highest-resolution bin.

Crystal Structure Determination and Refinement

The structures of the modified DDDs were determined by molecular replacement using the Dickerson-Drew dodecamer as the search model (PDB code 436D (77)). Molecular replacement searches were completed with program MOLREP of the CCP4 suite (94-96). An initial model was subsequently manually checked and rebuilt in program COOT (97). The final model was further rebuilt and refined using REFMAC 5.6 (98, 99). The final models were refined against all reflections, except for 5% randomly selected reflections that were used for monitoring R_{free} . The final refinement statistics are presented in Table 2.2.

Helicoidal Analysis

The calculations of helicoidal parameters, including backbone torsion angles, intra- and interbase translations and rotations, was performed using Curves+ (100). For crystal structures, the last refined structure was used, prior to validation.

CHAPTER 3

CHARACTERIZATION OF 5-HYDROXYMETHYLCYTOSINE IN DNA: COMPARISON WITH 5-METHYLCYTOSINE

Introduction

The methylation of cytosine by DNA methyltransferases (*101, 102*) to form 5-methylcytosine (5mC), plays an important role in the epigenetic regulation of the eukaryotic genome (*103, 104*). The reverse process of active demethylation has been of considerable interest (*105, 106*) and has led to the important discovery of 5-hydroxymethylcytosine (5hmC) in mammalian DNA (*107, 108*). The 5hmC base is generated from 5mC through the action of ten-eleven translocation (TET) deoxygenases (*109-111*). It is also formed in response to oxidative stress as a consequence of UV radiation (*112*). It is thought that 5hmC is itself an important epigenetic marker and transcriptional regulator. Support for this hypothesis comes from the observation that altered levels of 5mC and 5hmC are observed in early embryonic development, embryonic stem cell differentiation, and tumors (*109, 113*). Cellular levels of 5hmC are tissue-specific, with the highest levels found in the central nervous system (*114*). 5hmC values increase during brain development, suggesting a role of this modification in brain maturation and neuronal development (*108*). The balance between 5mC and 5hmC at gene promoters and CpG islands appears to be linked to pluripotency of the cell (*110*).

Further oxidation of 5hmC by TET deoxygenases leads to the formation of 5-formylcytosine (5fC) and 5-carboxycytosine (5caC) (*109-111*). However, mass

spectrometric analyses have led to estimates that some hundreds of pmols of 5hmC are present in mammalian tissues, while the levels of 5fC and 5caC are much lower (114) suggesting that the latter oxidation products are more efficiently removed from DNA than is 5hmC. Indeed, Maiti and Drohat (115) showed that both 5fC and 5caC are substrates for thymine DNA glycosylase (TDG), but not 5hmC. This suggested that active demethylation of 5mC requires further oxidation of 5hmC to the downstream products 5fC and 5caC, allowing TDG to restore the epigenetically unmodified G:C pair.

Mechanistically, TDG employs the familiar extrahelical base flipping mechanism (79-81) to position its substrates into its active site for catalysis. Recently, Renciuik *et al.* (116) obtained a structure of the Dickerson dodecamer, in which either a 5mC or 5hmC modification was placed site-specifically either at the C⁹:G¹⁶ base pair or the C³:G²² base pair. They concluded that the presence of either 5mC or 5hmC did not influence the overall DNA structure. The hydroxyl group of the 5hmC base was oriented toward the 3' end of the duplex, away from the phosphate backbone. They also reported the thermodynamic effects of these modified bases using a combination of CD and UV spectroscopy. The presence of the 5hmC modification resulted in a slight destabilization of the duplex. They proposed that the cytosine C5 carbon provides an ideal location to encode epigenetic information. Since polymerases may not be able to distinguish 5hmC from 5mC and unmodified cytosine, these modifications might be anticipated to be non-mutagenic.

Maiti and Drohat obtained the structure of the hTDG catalytic domain complexed with DNA containing an abasic site, (117) which revealed interactions promoting the specificity requirement of guanine vs. adenine as the pairing partner of the target base and additional protein-DNA interactions associated with the specificity for CpG sites. Further studies showed that the conserved Asn¹⁴⁰ residue was implicated in the chemical step, whereas the conserved Arg²⁷⁵ was implicated in nucleotide flipping into the active site (118). Subsequently, Hashimoto *et al.* (119) determined the structure of a post-

reactive complex of hTDG with a DNA containing a G:5hmU mismatch, showing that the glycosylase had flipped the 5hmU nucleotide from the DNA, and suggesting that TDG allows hydrogen-bonding interactions to both 5hmU and 5caC. They proposed that amino-imino tautomerization of the substrate base may explain how TDG discriminates against 5hmU and 5caC.

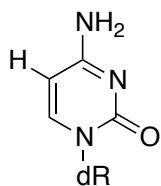
To further characterize the conformational consequences of 5hmC inserted into a DNA duplex, we collected NMR data on the Dickerson-Drew dodecamer (DDD) (120, 121) site-specifically modified with 5hmC, [5'-d(C¹G²C³G⁴A⁵A⁶T⁷T⁸Z⁹G¹⁰C¹¹G¹²)-3']₂, Z⁹=5hmC (Figure 3.1). We used high-resolution 2D NMR data to further characterize the 5hmC-modified duplex, and compared it to the 5mC duplex. We have also characterized the kinetics of base-pair opening in 5hmC-modified DNA by measuring the rates of exchange of imino protons with solvent protons by magnetization transfer from water, as a function of the concentration of exchange catalyst. Our results show that the equilibrium constant for opening of 5hmC is low. We gathered differential scanning calorimetry (DSC), and additional CD and UV spectroscopy data to characterize the thermodynamic effects of these 5-substituted cytosines in DNA. Finally, we determined at 1 Å resolution resolution the structure of the DDD with 5hmC at base pair C⁹:G¹⁶.

Results

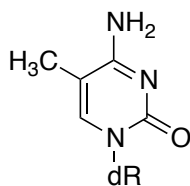
5-hydroxymethyl-cytosine in comparison with 5-methyl-cytosine

The DNA duplexes used in these studies are shown in Figure 3.1. The sequence we chose for these studies is well-established, self complementary Dickerson Drew dodecamer. The ninth residue in this sequence is replaced by either 5mC or 5hmC.

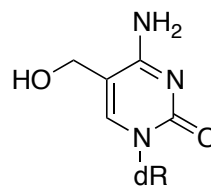
A



Cytosine; C



5-Methyl-cytosine; 5mC



5-Hydroxymethyl-cytosine; 5hmC

B

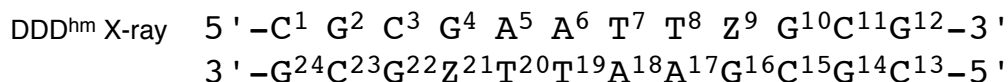
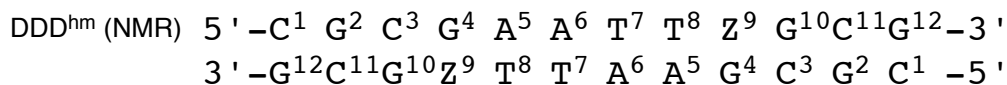
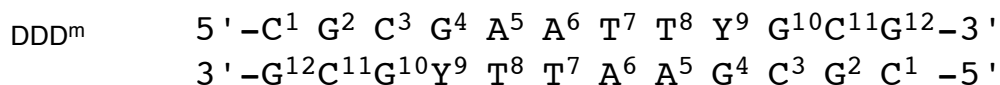
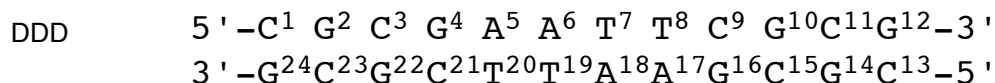


Figure 3.1. A. Structure of dC, 5mC and 5hmC. B. Sequences and numbering of the nucleotides for unmodified DDD, DDD^m, DDD^{hm} (NMR) and DDD^{hm} (X-ray) duplexes. In solution, the two strands of the DDD exhibit pseudo-dyad symmetry and the NMR resonances of symmetry-related nucleotides in the two strands are not individually observed. In the crystal, electron density for symmetry-related nucleotides in the two strands is observed and the nucleotides are numbered individually.

Unfolding Thermodynamic of Oligonucleotide Duplexes.

Typical UV melting curves of each duplex at 260nm and 275nm are compared in Figure 3.2.

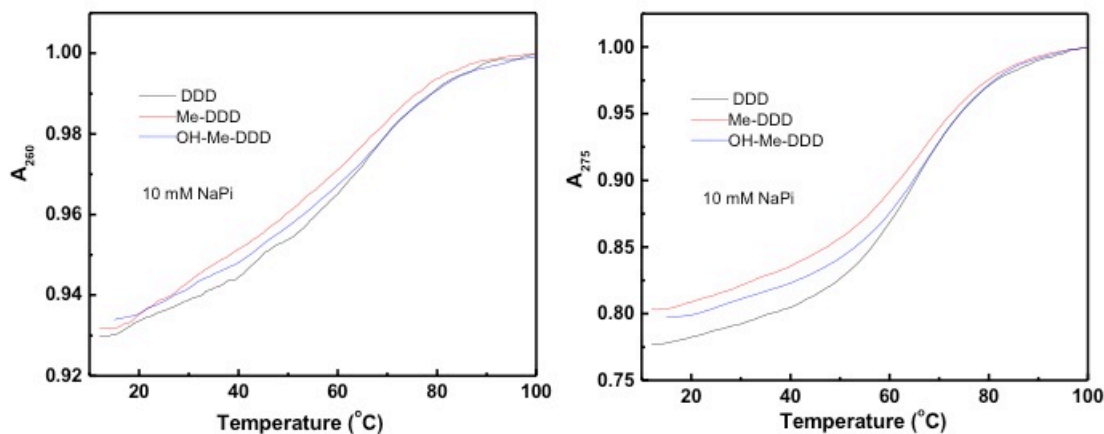


Figure 3.2. UV melting curves of DDD, DDD^m and DDD^{hm} at 260 nm and 275 nm, in 10 mM NaP_i buffer.

These curves show that at the lowest temperatures, below ~ 20 °C, each molecule is in the duplex state, while at higher temperatures, all curves are sigmoidal in shape which is characteristic of the unfolding of DNA duplexes. The inclusion of a methyl or hydroxymethyl groups into cytosine causes a weak and minimal thermal destabilization of the DNA. Their thermal stability is in the order: DDD^{hm} < DDD^m < DDD. Analysis of the melting curves as a function of C_T shows that the T_M values increase with increasing DNA concentration, Figure 3.3, as expected for the unfolding of self-complementary duplexes into two single strands.

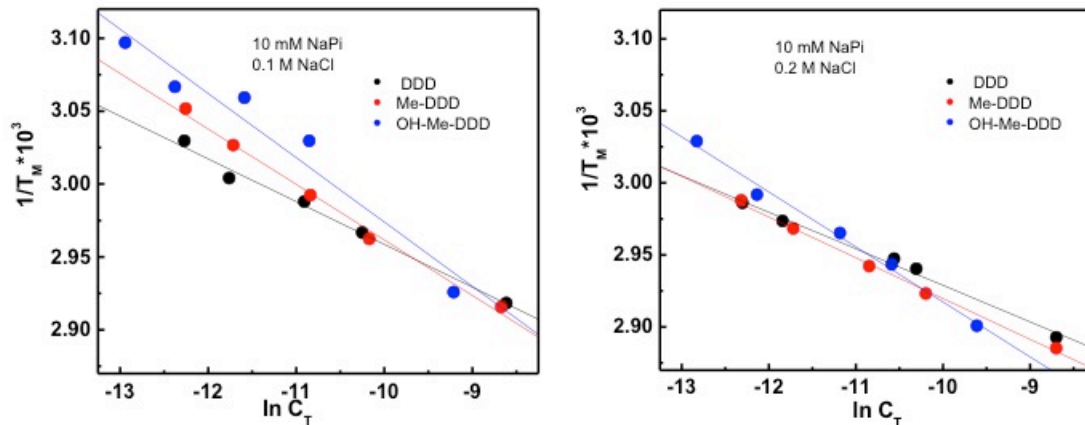


Figure 3.3. T_M -dependences on strand concentration of DDD, DDD^m and DDD^{hm}.

The CD spectra of each duplex at 20 °C show a positive Cotton effect at 280 nm and a strong negative Cotton effect around 250 nm, which is characteristic of a right-handed helix in the B-conformation. The intensities of both positive and negative bands are similar in magnitude, the intensity of the negative band correlates with the extent of base-pair stacking contributions. Thus, the incorporation of 5mC and 5hmC contributes equally to the extent of base-pair stacking interactions among these three duplexes. The CD spectra at 90 °C correspond to the characteristic spectra of DNA random coils.

The DSC melting curves for all three duplexes in 0.1 M and 0.2 M NaCl buffered solution are compared in Figure 3.4.

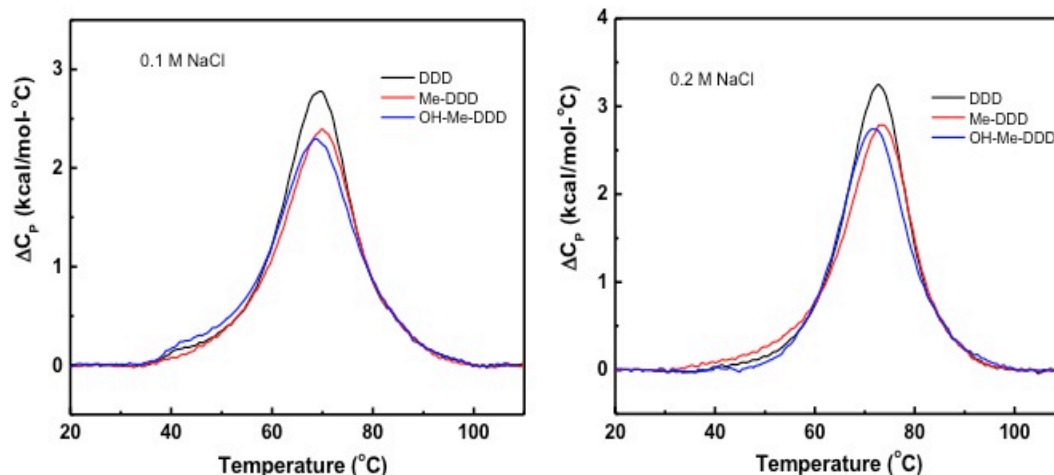


Figure 3.4. DSC melting curves of DDD, DDD^m and DDD^{hm} in 10 mM NaPi buffer at pH 7 and 0.1 M NaCl (left) or 0.2 M NaCl (right).

The thermodynamic profiles at 0.1 M and 0.2 M NaCl are summarized in Table 3.1. The unfolding of each duplex is monophasic, the increased in salt shifts the duplex transition curves to higher temperatures, and this is due to a higher screening by salt of the duplex phosphates. Analysis of these curves revealed a decreased average endothermic enthalpy of 5.5 kcal/mol for the modified duplexes relative to DDD in the 0.1-0.2 M NaCl range (Table 3.1). This decrease in enthalpy, due to the incorporation of modified cytosines, is within the experimental error of the enthalpy determination. The lowering of the enthalpies at lowered salts suggests the presence of heat capacity effects. The heat capacity values were 0.6 kcal/K·mol (DDD), 1.7 kcal/K·mol (DDD^m) and 1.1 kcal/K·mol (DDD^{hm}). The presence of heat capacity effects has been attributed to the exposure of non-polar groups to the solvent and/or to changes in structural hydration between the unfolded and folded states (83).

Sample	T_M (°C)	ΔH_{cal} (kcal/mol)	$\Delta G^\circ_{(20)}$ (kcal/mol)	$T\Delta S$ (kcal/mol)	ΔH_{vH}^* (kcal/mol)	Δn_{Na^+} (mol _{Na⁺} / mol)	ΔC_p (kcal/mol-°C)
DDD	69.6	-108.4	-15.7	-92.7	68	-2.8	-0.6
	(72.6)	(-110.2)	(-16.8)	(-93.4)	(78)		
DDD ^m	69.9	-99.6	-14.5	-85.1	52	-2.6	-1.7
	(73.5)	(-105.8)	(-16.3)	(-89.5)	(70)		
DDD ^{hm}	68.7	-102.2	-15.7	-89.5	45	-2.5	-1.1
	(71.6)	(-105.3)	(-15.0)	(-85.0)	(52)		

Table 3.1. Folding thermodynamic profiles for DDD, DDD^m and DDD^{hm}. All experiments were performed in 10 mM NaP_i buffer at pH 7.0 and 0.1 M NaCl or 0.2 M NaCl in parenthesis. *The ΔH_{vH} is calculated from the dependences of the T_M on strand concentration.

Furthermore, the average values of the $\Delta H_{vH} / \Delta H_{cal}$ ratios at these salt concentrations are: 0.67 (DDD), 0.59 (DDD^m) and 0.46 (DDD^{hm}), indicating all three duplexes unfolds in non two-state transitions. The incorporation of the modified bases actually lowers the size of the cooperative melting unit.

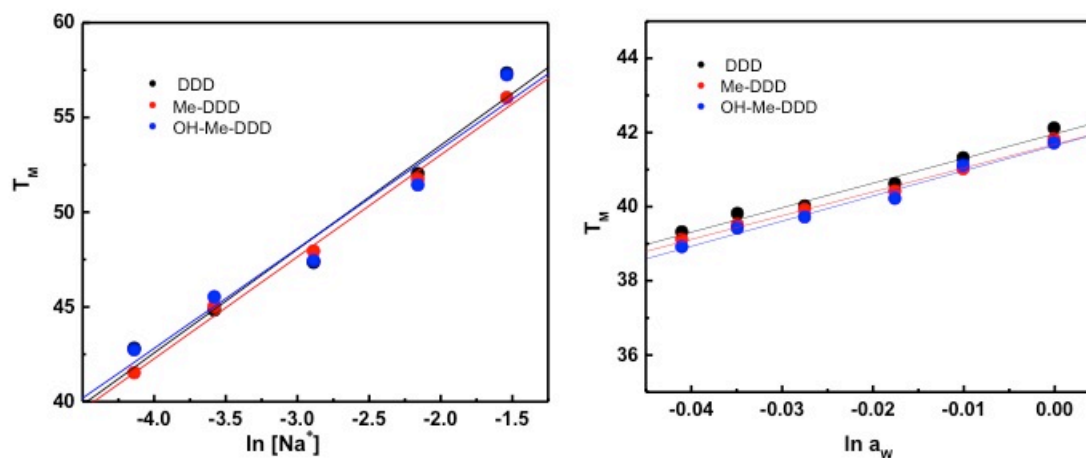


Figure 3.5. T_M -dependences of DDD, DDD^m and DDD^{hm} on salt concentration and water activity.

UV melting curves were measured at different salt concentrations, 16-216 mM NaCl, to determine the differential binding of counterions, Δn_{Na^+} . The T_M values increase linearly as a function of $\ln [\text{Na}^+]$ as shown in Figure 3.5., consistent with the observation that salt favors the duplex state with a higher charge density parameter. We obtained Δn_{Na^+} values (Table 3.1.) of 2.8 mol Na^+ /mol DNA (DDD), 2.6 kcal/K·mol (DDD^m) and 2.5 kcal/K·mol (DDD^{hm}).

The thermodynamic profiles for the folding of each duplex are listed in Table 3.1. Analysis of the data shows that a favorable Gibbs free energy accompanies the stable formation of each duplex, which results from compensation of favorable enthalpy-unfavorable entropy contributions. The favorable enthalpies arise from the formation of base-pairs and base pair stacks. Unfavorable entropy terms includes the ordering of two strands to form a duplex and condensation of counterions.

Relative to the unmodified dodecamer duplex, the DDD^m and DDD^{hm} modified oligomers were destabilized at 0.1 M and 0.2 M NaCl concentrations. Specifically, the inclusion of two 5mC or two 5hmC modifications in DDD yielded an average decrease in $\Delta G^\circ_{(20)}$ of 0.9 kcal/mol, which is enthalpy driven ($\Delta\Delta H = 6.1$ kcal/mol).

NMR melting studies

A series of 1D ¹H NMR spectra for the exchangeable protons were recorded at 5, 15, 25, 35, 45, and 55 °C and are shown in Figure 3.6. The data show that the G²N1H imino proton resonance of the G²:C¹¹ base pair in the DDD^m was sharp and detectable only at 5 °C. The same imino peak in the DDD^{hm} began to broaden at 25 °C, and at higher temperatures this resonance disappeared. In the unmodified duplex the G²

N1H imino proton of the G²:C¹¹ base pair was sharp up to 20 °C, above which it started to broaden. The G¹⁰ N1H imino proton of the 5'-neighbor base pair, C³:G¹⁰ was significantly broadened at 35 °C in the DDD^m, while still sharp even at 45 °C for the DDD, and at 35 °C in DDD^{hm} this imino resonance started to broaden. The G⁴ N1H imino proton of the G⁴:C⁹ base pair in DDD and the G⁴:Z⁹ proton of the DDD^{hm} duplex remained sharp at as high temperature as 45 °C, and at 55 °C started to broaden; however the same resonance in the DDD^m at 45 °C was already broadened. The N3H imino resonances of T⁷ and T⁸ were the sharpest in all three duplexes; they started to broaden at 55 °C in the DDD and DDD^{hm} duplexes, while in the DDD^m these peaks were not observed.

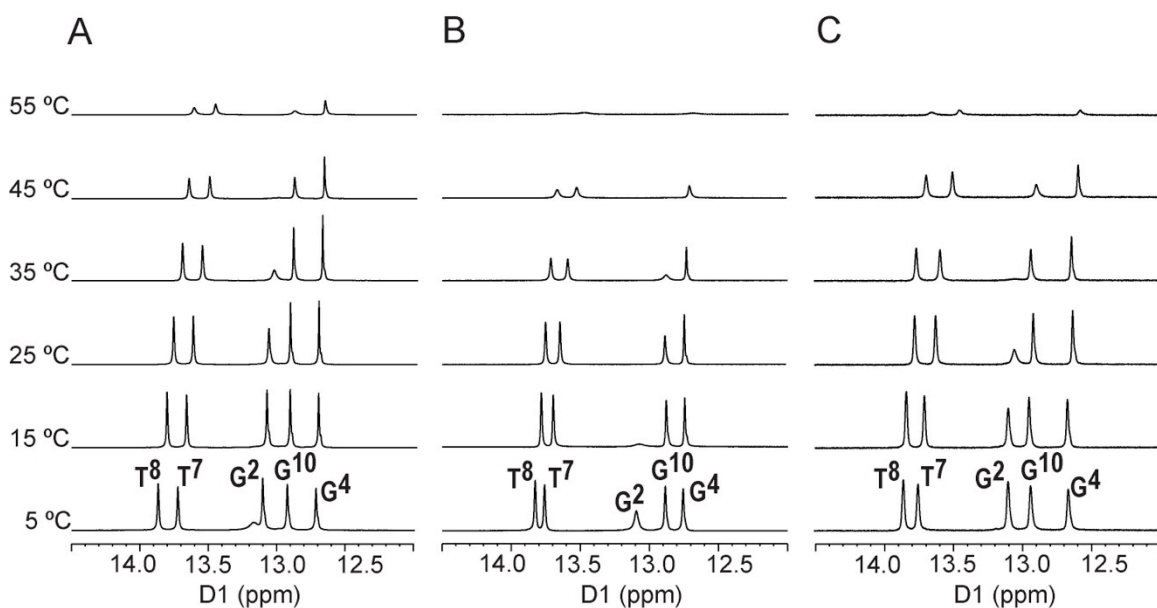


Figure 3.6. ¹H-NMR of imino proton resonances as a function of temperature for the unmodified DDD duplex (A), modified DDD^m (B), and modified DDD^{hm} (C) duplexes.

Exchangeable protons

Figure 3.7 shows the NOE connectivity of the purine N1H and pyrimidine N3H imino protons, for the DDD, DDD^m, and DDD^{hm} duplexes. The base imino protons were assigned based on their sequential connectivities in NOESY spectra, and these assignments were supported by their NOE cross-peaks to Watson-Crick base-paired amino protons (69). The sequential connectivities were obtained from base pairs G²:C¹¹ → C³:G¹⁰ → G⁴:C⁹ → A⁵:T⁸ → A⁶:T⁷. For the DDD and DDD^m duplexes the imino-proton resonances of the terminal base pairs C¹:G¹² were not observed, presumably due to fast exchange with water. In the region of the NOESY spectrum showing the NOEs between the imino and amino protons cross-peaks from modified base C⁹H41, C⁹H42 to the complementary base G⁴H1 were observed, as well as interactions to neighbor bases T⁸H3 and G¹⁰H1 (Figure 3.7).

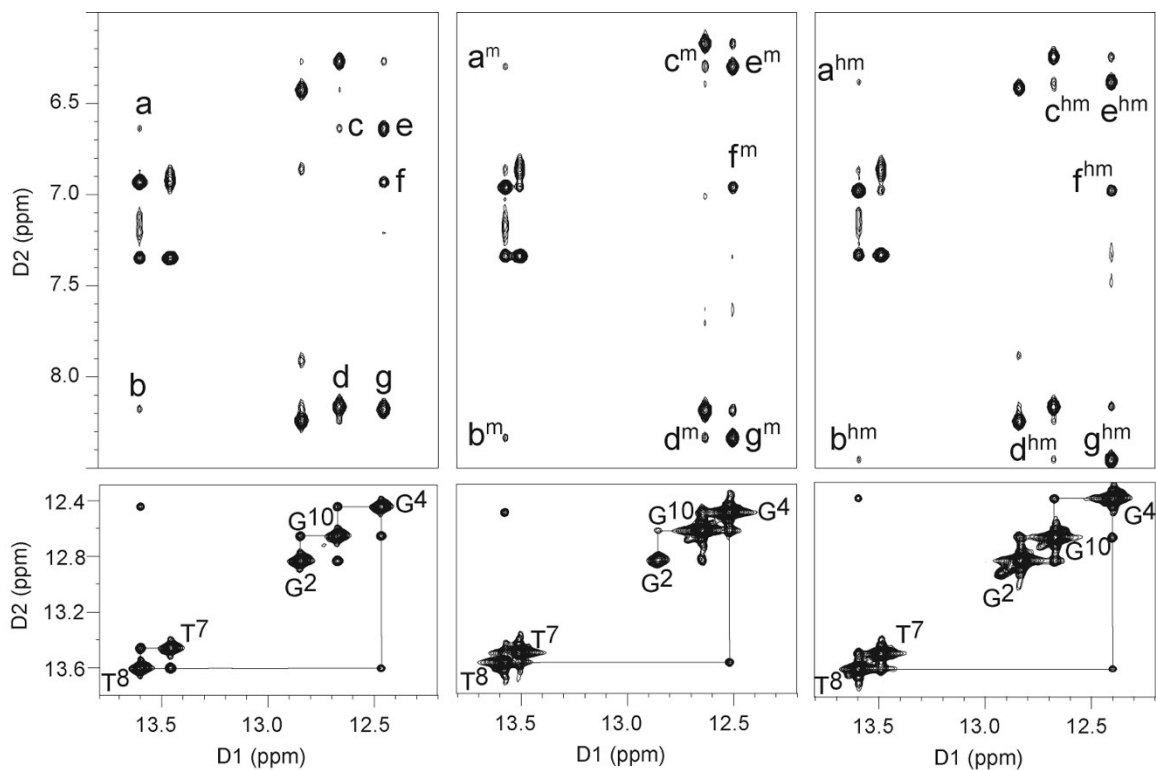


Figure 3.7. ^1H - ^1H NMR NOESY spectrum showing resonances for the thymine and guanine imino protons and sequential NOE connectivity for the imino protons of the base pairs $\text{G}^2:\text{C}^{11}$ to $\text{A}^6:\text{T}^7$ for the unmodified DDD (A), modified DDD^m (B), and modified DDD^{hm} (C) duplexes. Expansion of the ^1H - ^1H NOESY spectra for the DDD (A), DDD^m (B) and DDD^{hm} (C) showing the conservation of Watson-Crick base pairing and base stacking at the modification site. a, $\text{C}^9\text{H}41 \rightarrow \text{T}^8\text{H}3$; b, $\text{C}^9\text{H}42 \rightarrow \text{T}^8\text{H}3$; c, $\text{C}^9\text{H}41 \rightarrow \text{G}^{10}\text{H}1$; d, $\text{C}^9\text{H}42 \rightarrow \text{G}^{10}\text{H}1$; e, $\text{C}^9\text{H}41 \rightarrow \text{G}^4\text{H}1$; f, $\text{A}^5\text{H}2 \rightarrow \text{G}^4\text{H}1$; g, $\text{C}^9\text{H}42 \rightarrow \text{G}^4\text{H}1$. (Index ^m or ^{hm} refers to the base pairs in the modified duplexes, DDD^m and DDD^{hm}, respectively).

Nonexchangeable protons

For each of the three duplexes, the sequential assignment of non-exchangeable protons was accomplished using standard protocols (122, 123). In each instance, the anticipated pattern of sequential base aromatic \rightarrow deoxyribose anomeric NOEs was identified from $\text{C}^1 \rightarrow \text{G}^{12}$ (Figure 3.8). Three strong NOEs accounted for H5-H6 cross-peaks of cytosines C^1 , C^3 , and C^{11} .

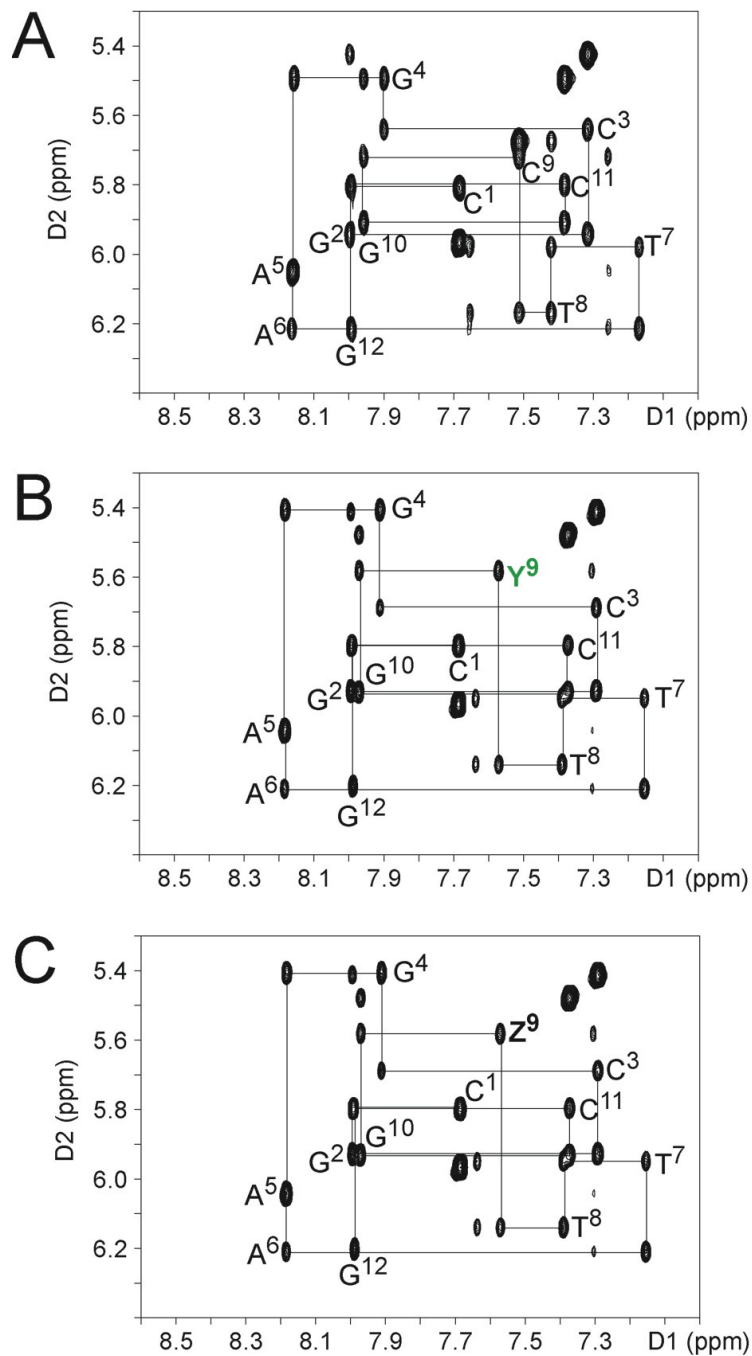


Figure 3.8. Expanded plot from the aromatic-anomeric region of the NOESY spectrum, showing sequential NOE connectivities of the unmodified DDD (A), modified DDD^m (B), and modified DDD^{hm} (C) duplexes.

Cytosine assignments were confirmed by a DQF-COSY spectrum recorded at identical conditions (data not shown). As compared to the DDD, in the DDD^m and DDD^{hm}

duplexes the cytosine H5 protons at nucleotides Y⁹ or Z⁹ were absent due to presence of either the methyl or hydroxymethyl substitution at the cytosine C5 position, respectively; thus, no cytosine H5-H6 cross-peaks were observed in this region of the spectrum for nucleotides Y⁹ and Z⁹. In all instances, for all three duplexes, the NOE cross-peaks intensities between the base protons and the deoxyribose H1' protons were of the same relative magnitudes as those between other bases in the sequence, indicating that the glycosyl bonds maintained the anticipated anti conformations associated with B-type DNA.

Dynamics of base pair opening

The imino proton exchange rates for the three duplexes were obtained by measuring magnetization transfer from water as a function of added ammonia base catalyst. The dependence of the exchange rates on the catalyst concentration is illustrated in Figure 3.9. for the C³:G¹⁰, G⁴:C⁹, A⁵:T⁸ and A⁶:T⁷ base pairs of the DDD, DDD^m and DDD^{hm} duplexes. This figure also shows the fits of the exchange rate as a function of increasing ammonia base concentration to equation 8.

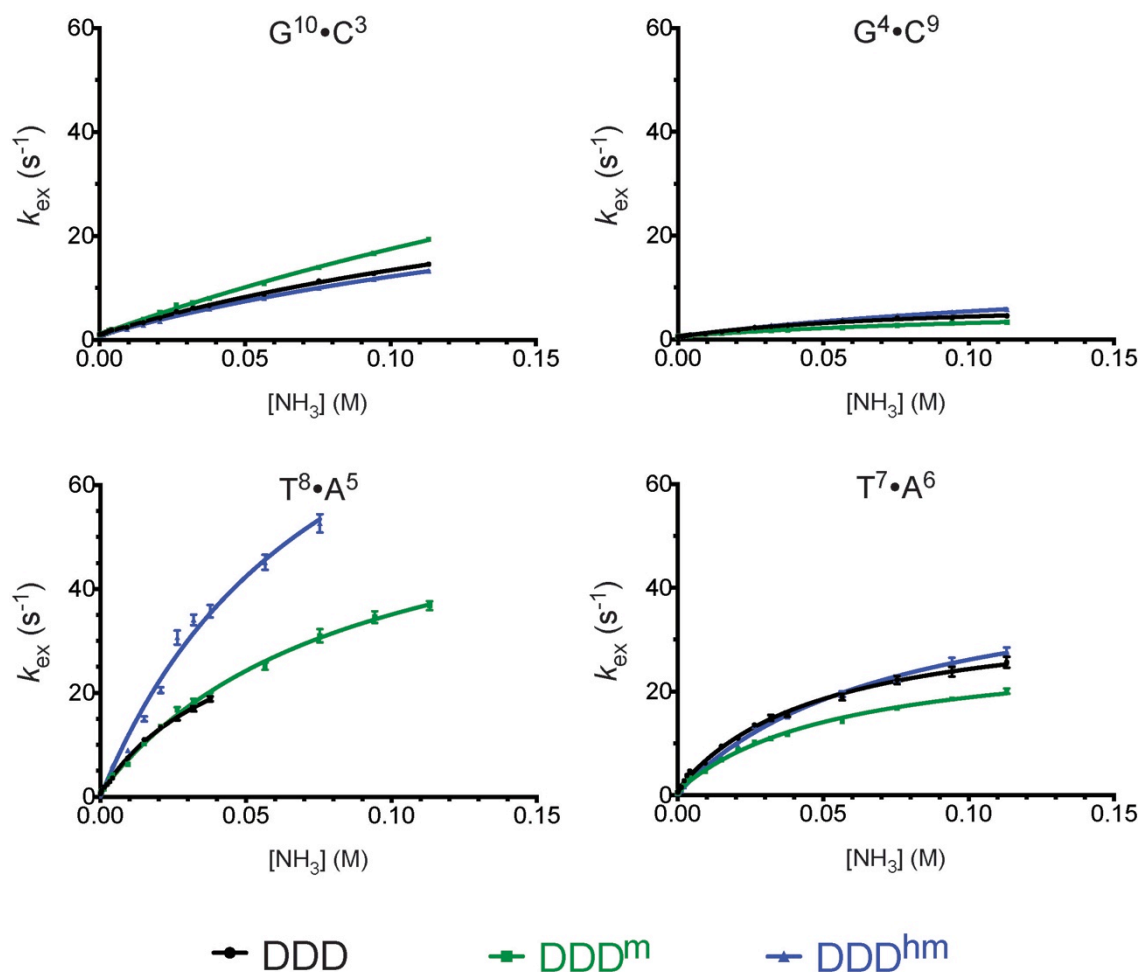


Figure 3.9. Plots showing imino proton exchange rates obtained by monitoring magnetization from water as a function of ammonia base catalyst. A. $G^{10}:C^3$. B. $G^4:C^9$. C. $T^8:A^5$. D. $T^7:A^6$ in DDD (black), DDD^m (green) and DDD^{hm} (blue).

These fits were used to determine the base pair opening (k_{op}) and closing (k_{cl}) rates. To obtain the opening equilibrium constant (K_{op}) data were fitted to the eq. 9 in the lower range of catalyst concentration where the dependence between exchange rate and concentration of base catalyst is linear. The values of k_{op} , k_{cl} and K_{op} in the three duplexes are summarized in Table 3.2.

	$k_0 (\text{s}^{-1})^a$			$K_{\text{op}} \times 10^7$		
	DDD	DDD ^m	DDD ^{hm}	DDD	DDD ^m	DDD ^{hm}
C ³ :G ¹⁰	1.1 ± 0.06	1.0 ± 0.07	0.95 ± 0.05	2.9 ± 0.1	3.6 ± 0.1	2.5 ± 0.1
G ⁴ :C ⁹	0.57 ± 0.03	0.56 ± 0.1	0.68 ± 0.05	1.2 ± 0.04	0.75 ± 0.04	1.2 ± 0.06
A ⁵ :T ⁸	0.59 ± 0.03	0.73 ± 0.07	0.77 ± 0.03	41 ± 0.08	35 ± 2	73 ± 6
A ⁶ :T ⁷	0.61 ± 0.03	0.43 ± 0.04	0.36 ± 0.01	37 ± 5	26 ± 3	29 ± 3

	$k_{\text{op}} (\text{s}^{-1})$			$k_{\text{cl}} (\times 10^{-7} \text{ s}^{-1})$		
	DDD	DDD ^m	DDD ^{hm}	DDD	DDD ^m	DDD ^{hm}
C ³ :G ¹⁰	45 ± 3	90 ± 12	42 ± 5	15 ± 1	25 ± 2	17 ± 1
G ⁴ :C ⁹	8 ± 0.5	7 ± 0.6	16 ± 2	6.7 ± 0.2	9.4 ± 1	13 ± 0.8
A ⁵ :T ⁸	40 ± 2	65 ± 2	110 ± 13	0.97 ± 0.04	1.9 ± 0.04	1.5 ± 0.05
A ⁶ :T ⁷	36 ± 1	29 ± 1	45 ± 2	0.98 ± 0.09	1.1 ± 0.08	1.6 ± 0.08

Table 3.2. Rate and Equilibrium Constants for DNA Base Pair Opening. ^aThe observed exchange rate without an ammonia catalyst.

For each of the observed imino protons, in all three duplexes, as the concentration of ammonia increased, the exchange rate increased until a plateau was reached, which indicated a change for the rate-limiting step from chemical exchange to base pair opening ($k_{\text{ex}} = k_{\text{op}}$; exchange occurs at each opening event). For the G⁴ N1H imino proton, the exchange rate had only a weak dependence on the concentration of ammonia. This reflected the small equilibrium constant for the opening the G⁴:C⁹ base pair, and resulted in low and similar K_{op} values in the DDD, DDD^m and DDD^{hm} duplexes (1.2×10^7 , 0.75×10^7 , and 1.2×10^7 , respectively). Also, opening rates decreased more for base pair G⁴:C⁹ than for the other base pairs (Table 3.2), suggesting that the opening

dynamics of this base pair are lower. The opening rate for the G⁴:C⁹ base pair was ~2 times greater in DDD^{hm} ($k_{op} = 16 \text{ s}^{-1}$), than in DDD and DDD^m ($k_{op} = 8$ and 7 s^{-1} , respectively), but the open lifetime expressed by $1/k_{cl}$ was ~2 times less in DDD^{hm} and ~1.5 in DDD^m than in DDD (Table 3.2). For the C³:G¹⁰ base pair, the opening rate was greater than for the other C:G base pairs. However, it was comparable for the DDD and DDD^{hm} duplexes (42 and 45 s^{-1} , respectively). This was confirmed by the opening equilibrium constant values (Table 3.2). The greatest values for the equilibrium constants for base pair opening as well as the opening rates were observed for the T⁸ N3H imino proton, which is located in the 5' neighbor base pair from the modification sites. This applied to all three duplexes. The dynamics of this base pair were different for each duplex. As compared to base pair A⁵:T⁸ in the DDD, in the DDD^m and DDD^{hm} duplexes this base pair opened ~1.5 and ~3 times faster, respectively. For the terminal C¹:G¹² and penultimate G²:C¹¹ base pairs the exchange rates could not be measured because of the exchange with the solvent protons.

The atomic resolution crystal structure of modified DDD

Crystals belonged to the primitive orthorhombic space group $P2_12_12_1$ with unit cell parameters $a=25.3 \text{ \AA}$, $b=40.2 \text{ \AA}$, $c=65.5 \text{ \AA}$. The crystal structure of DDD^{hm} was refined anisotropically to 1.02 \AA resolution, and is shown in Figure 3.10. The crystallographic asymmetric unit consists of two chemically equivalent self-complementary strands of the DDD^{hm} antiparallel duplex numbered (C¹ to G¹² and C¹³ to G²⁴), one magnesium ion and 2 molecules of spermine and 221 water molecules. The electron density maps obtained were of high quality and allowed building

of a detailed model as well as modeling alternative conformations. The final model exhibited very good crystallographic statistics and was refined to a final R_{work} of 15.7 and R_{free} of 17.8 (Table 2.2).

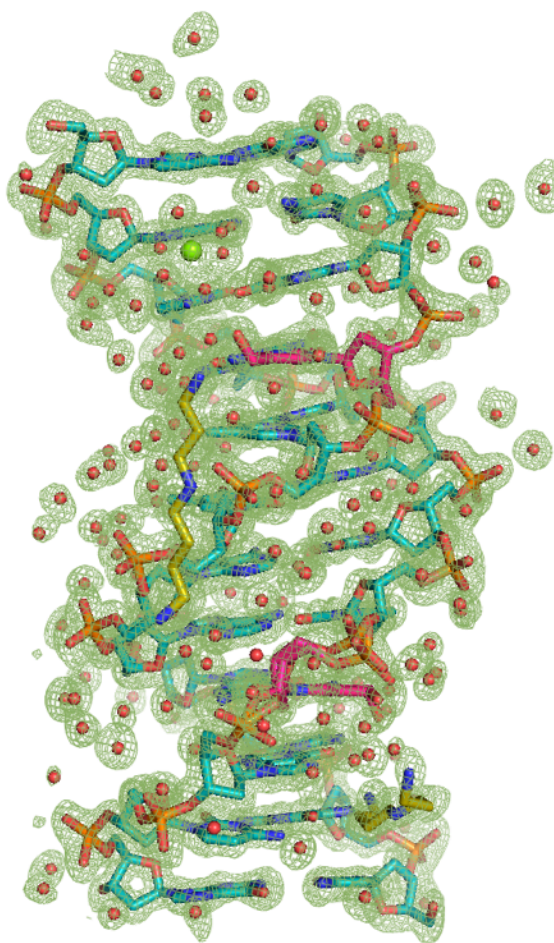


Figure 3.10. Fourier (2Fo-Fc) sum electron density contoured at the 1.0σ level (green meshwork surrounding the DDD^{hm} duplex. 5hmC modified base shown in pink.

The DDD^{hm} structure is consistent with the other DDD structures. It overlays well with the canonical structure of DDD (PDB code: 1BNA) with rmsd of 0.7 \AA (least squares (LSQ) superposition using all atoms fit in COOT), and with the high resolution structure of unmodified DDD (PDB code: 355D) with rmsd of 0.7 \AA . The base-stacking

pattern in the modified DDD^{hm} is very similar to the one observed in unmodified B-type DNA and helical parameters obtained by program Curves+ are comparable (Figures 3.13 and 3.14). As anticipated presence of the hydroxymethyl side chain at the cytidine nucleobase, did not significantly alter base packing between $Z^{21}:\text{G}^4$, nor did it disrupt Watson-Crick base pairing or changed the conformation of these residues (Figure 3.11 and 3.12). It also did not alter Mg^{2+} binding in the crystal structure. The single octahedrally hydrated magnesium cation is located in the major groove and it found to interact via waters with G^2 (chain A), $\text{O}6$ of G^{22} (chain B) and with $\text{O}2\text{P}$ ($\text{P}^{7'}$) and $\text{O}1\text{P}$ ($\text{P}^{6'}$) of an adjacent symmetry related molecule, as it was described previously by Drew-Dickerson *et al.* (14), and Tereshko *et al.* (77). It is noteworthy that an additional hydrogen bond interaction is formed between the hydroxyl group of modified cytosine Z^{21} (chain B), and the axially coordinated water (HOH 12) (2.7 Å distance), whereby it interacts further with N^7 of the G^{22} residue (2.8 Å distance). An additional H-bond interaction is observed between hydroxyl group on Z^{21} and $\text{O}6$ of G^{22} via water molecule HOH 11 (3.0 Å distance from Z^{21} to HOH11, and 2.7 Å distance from HOH11 to $\text{O}6$ of G^{22}) (Figure 3.11).

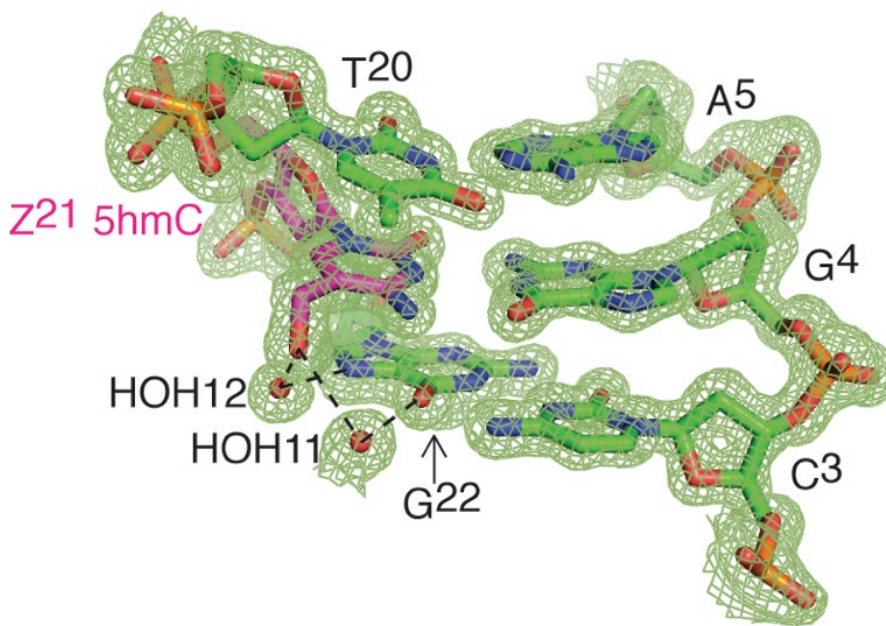


Figure 3.11. Sum electron density contoured at the 1.0σ level (green meshwork) surrounding the DDD^{hm} duplex at the modification site.

In the second modified base Z^9 (chain A) the hydroxyl moiety is observed to be in two conformations (a major conformation refined with occupancy of 0.8 and a minor with occupancy of 0.2). In the major conformation the 5-hydroxyl moiety forms a H-bond with the neighboring N^1 of spermine I residue and the $O6$ of G^{10} via HOH 200 water molecule. In the minor conformation, the 5-hydroxyl moiety is turned towards backbone phosphate moiety and interacts with the neighboring waters (HOH 155). In addition, dual conformations (with occupancies 0.8/0.2) were also modeled for residues G^{10} of chain A and the complementary counterparts in chain B (the residues A^{18} and T^{19}), such that significant differences in the phosphate backbone positions are observed. Analysis of the waters forming the “spine of hydration” shows that waters involved in the minor groove hydration are conserved

and superimposable between the structures. Analysis of the alternative conformations of DDD^{hm} did not reveal significant changes in the helical parameters (Figures 3.13 and 3.14).

The differences between the two conformations primarily entail changes in torsion angles and phosphate backbone positions and might arise as a result of the association with the spermine molecules (which seem to slightly alter crystal-packing interactions. Spermine I, that is fully visible in the electron density map (all 14 non-hydrogen atoms) is located in at the end of the duplex, and extends between the bases of G¹⁰ and G¹⁴ towards the phosphate residue of the Z⁹. The spermine was modeled in two conformations, which mainly differ in the position of the terminal ammonium groups (with occupancy 0.5 and 0.3). The interactions between spermine and DNA appear to be limited to direct interactions via N¹, N¹¹, and N¹⁴.

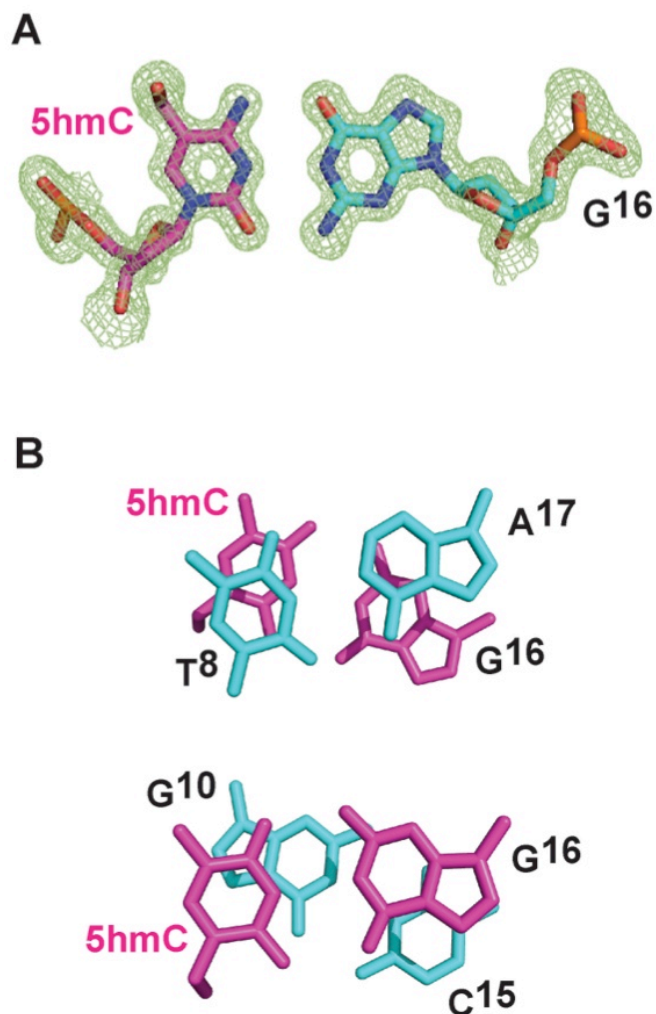


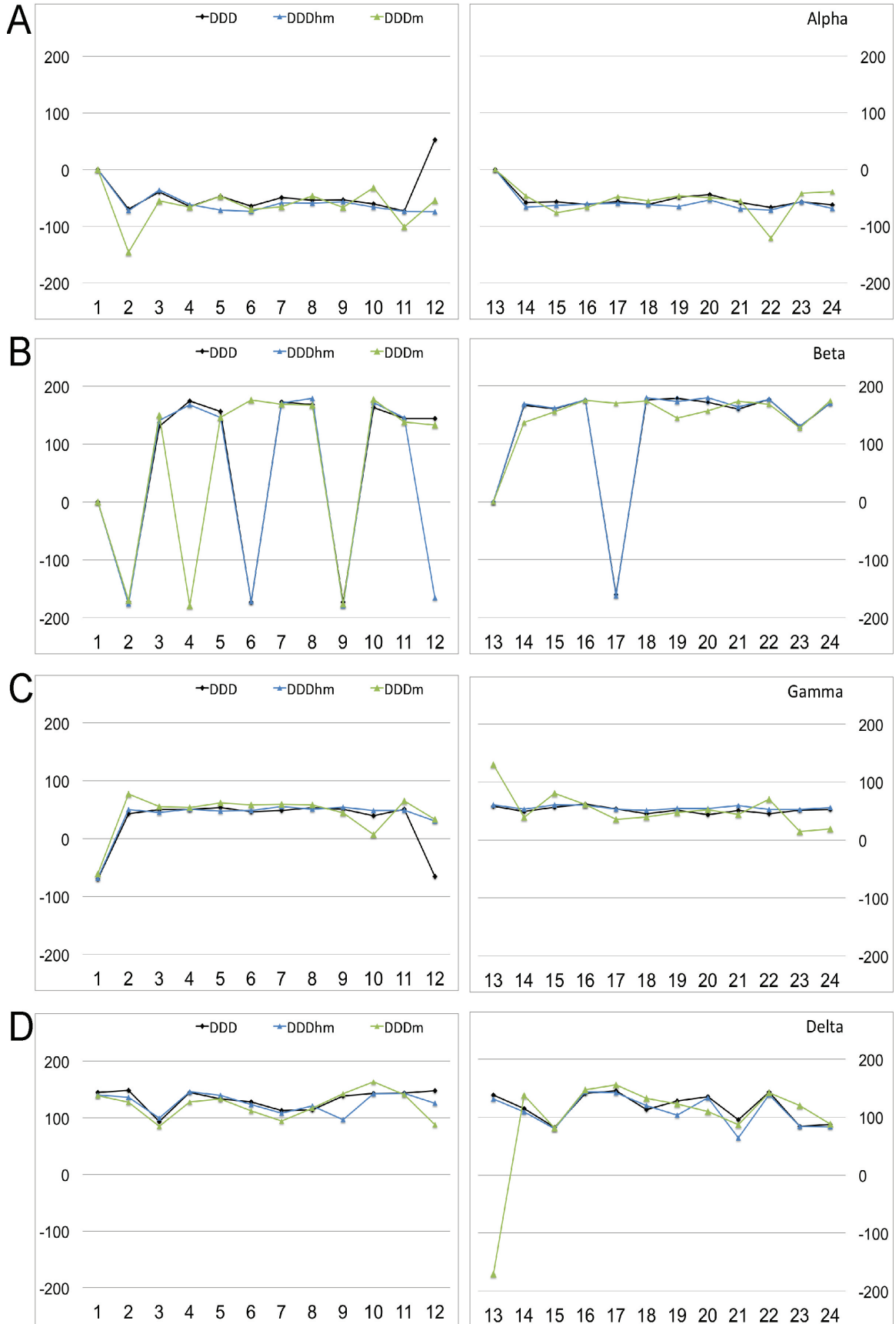
Figure 3.12. Sum electron density contoured at the 1.0σ level (green meshwork) around the 5hmC:dG modified base pair showing Watson-Crick interactions (A), and base stacking of 5hmC·dG modified base pair with T⁸:A¹⁷ and G¹⁰:C¹⁵ (B).

Spermine II is partially disordered in the density maps and only 11 residues were modeled. This molecule is positioned at the center of the duplex and is wedged between the sugar phosphate backbones of strand B (residues T¹⁹, T²⁰) and the strands of the symmetry mate (residues G¹², chain A', and A¹⁸, chain B'). The N⁵ amino group of spermine II interacts with *OIP* (3.00 Å) and *O2P* (3.32 Å), whereas N¹⁰ interacts with *OIP* of T¹⁹ (2.91 Å) and likely triggers the dual conformation of the phosphate

moiety of the neighboring nucleotide. The phosphate moiety of T¹⁹ adopts two conformations such that the positions of phosphate groups differ by ~0.8 Å. We suspect that there might be a third spermine molecule, which is positioned next to the phosphates of the residues Z²¹ and G²². However, it was not modeled due to the disordered density features, and instead was modeled with water molecules.

Helicoidal analysis

An analysis of the helical parameters of the crystal structure was performed in program Curves+ (100). The results of the analysis of torsion angles are shown in Figure 3.13. There were no significant changes in the alpha, gamma and delta torsion angles. For torsion angle beta, the DDD^{hm} and unmodified DDD are similar (77), however, peaks at the G², A⁶ and A¹⁷ reflect changes induced by the interacting Mg²⁺ ion that is observed in both structures. The effect of ion binding is also reflected in the changes in the epsilon torsion angle. Significant changes in the zeta torsion angle between DDD^{hm} and the unmodified DDD for residue G⁴ are due to the presence of the modification on the opposite base Z⁹. The analysis of interbase parameters shows no substantial changes in helical rise, roll and twist, as shown in Figure 3.14.



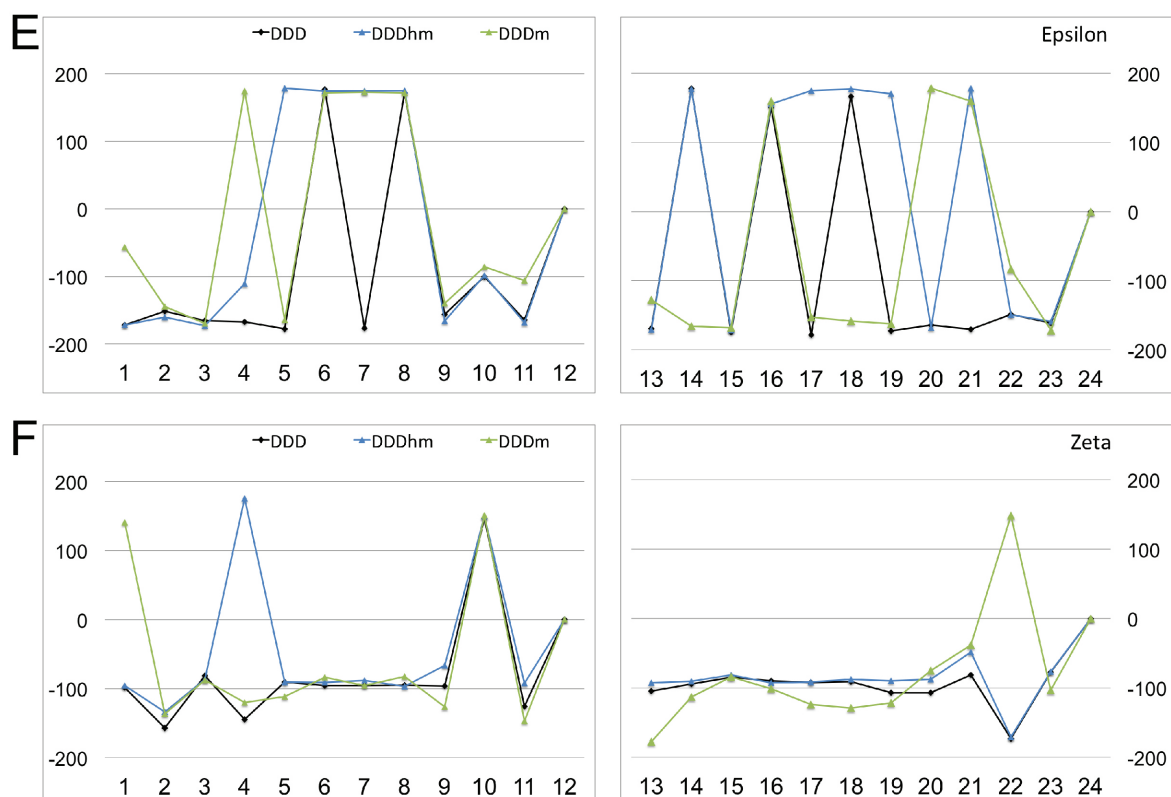


Figure 3.13. Comparison of backbone torsion angles (a) alpha, (b) beta, (c) gamma, (d) delta, (e) epsilon, and (f) zeta in the structure of the DDD^{hm} in blue, unmodified DDD (PDB entry 436D (*124*)) in black, DDD^m (PDB entry 265D) in green.

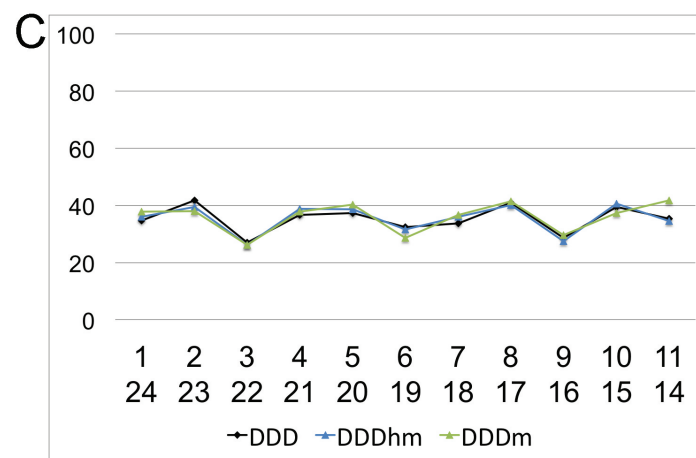
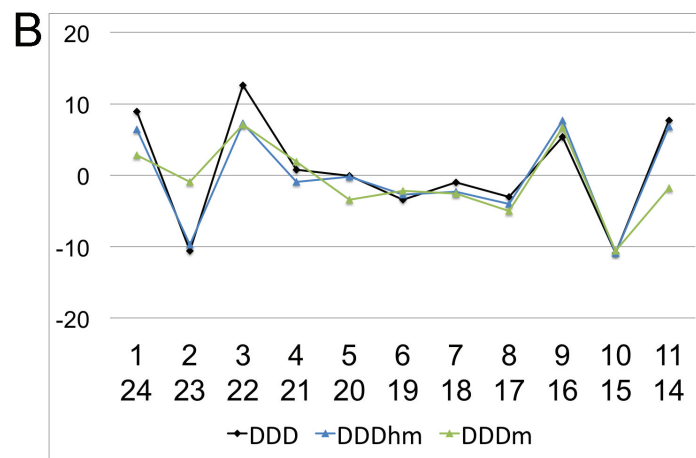
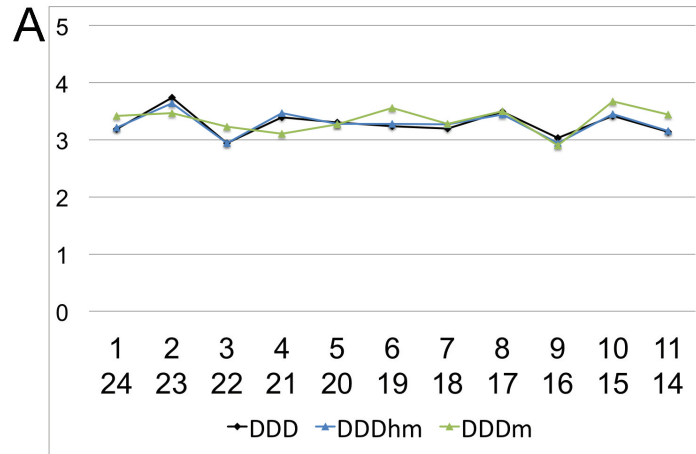


Figure 3.14. Interbase pair parameters: (a) helical rise, (b) roll, and (c) twist for the structure of DDDhm in blue, unmodified DDD (PDB entry 436D (124)) in black, DDDm (PDB entry 265D) in green.

Discussion

It has been shown that active demethylation involves oxidation by TET enzymes, which oxidize 5mC to 5hmC and further to 5fC and 5caC. These oxidation products provide substrates for BER, that would reinsert cytosines into the DNA (104, 112, 125, 126).

Thermodynamic Analysis of DNA with 5mC or 5hmC

DSC and UV spectroscopy experiments revealed, that with respect to the free energy of duplex formation, the presence of either 5mC or 5hmC in the DDD is similarly favorable for the DDD sequence; $\Delta G = -14.5$ kcal/mol and -15.7 kcal/mol for DDD^m and DDD^{hm}, respectively; compared to $\Delta G = -15.7$ kcal/mol in DDD (Table 3.1). All three duplexes possess similar T_M values, even at higher salt conditions (Figure 3.2 and Table 3.1). These results are in agreement with previously reported UV spectroscopic data (116).

Imino Proton Exchange and Base Pair Opening Kinetics

NMR spectroscopy as a function of temperature (Figure 3.6) confirms the thermodynamic results (Table 3.1). All of the imino resonances are sharp and stable at 45 °C. The G⁴ N1H imino proton of G⁴:Z⁹, which represents the modified base pair, exhibits the sharpest resonance in the spectrum. This reflects a lower exchange rate with water as shown in the kinetic analysis. For the G⁴N1H imino proton, the exchange rate has only a weak dependence on the concentration of ammonia base (Figure 3.9).

This is reflected in the corresponding small equilibrium constant for opening of the G⁴:C⁹ base pair ($K_{\text{op}} = 1.2 \times 10^7$ for DDD^{hm} and DDD, 0.75×10^7 for DDD^m), and reflects a short open lifetime (Table 3.2). From the 3'- site, flanking C³:G²² base pair interacts via two water molecules to the hydroxymethyl group of the modified 5hmC base (Figure 3.11). This could suggest stabilization effects and lower dynamic properties. However the open lifetime expressed by $1/k_{\text{cl}}$ stays at similar levels for both base pairs ($k_{\text{cl}} = 17 \times 10^{-7} \text{ s}^{-1}$ for C³:G¹⁰ in DDD^{hm}, and $13 \times 10^{-7} \text{ s}^{-1}$ for G⁴:Z⁹ in DDD^{hm}), even though the overall opening rates for C³:G¹⁰ in DDD^{hm} are somewhat higher than for the modified G⁴:C⁹ ($k_{\text{op}} = 42 \text{ s}^{-1}$ for C³:G¹⁰ in DDD^{hm} and $k_{\text{op}} = 16 \text{ s}^{-1}$ for G⁴:Z⁹ in DDD^{hm}) (Table 3.2). Finally, the rates for G⁴:C⁹ base pair opening have similar values in DDD^{hm} and DDD, which is also reflected by comparable opening equilibrium constants. A comparison of aK_{op} for the A⁵:T⁸ base pair showed an increased opening equilibrium for DDD^{hm} ($aK_{\text{op}} = 73 \times 10^7$), whereas the other imino proton resonances showed only a small or no increase. Compared with the unmodified base pair A⁵:T⁸ in DDD^{hm} spends twice as much time in the open state toward the solution. A key finding is that the apparent closing rate for the A⁵:T⁸ base pair in all three duplexes is small, while it opens 1.5 and 3 times faster in the DDD^m and DDD^{hm}, respectively, as compared with the DDD control duplex (Table 3.2). The higher exchange and base pair opening rates for thymine relative to guanine base pairs was expected (85). However, the 3-fold increase in opening rate and double in equilibrium constant for the 5' neighbor A⁵:T⁸ base pair in DDD^{hm} is significant. From the other site there is an interaction of the hydroxyl group with 3'-flanking G²², which might explain the stabilizing effect of 5hmC on the overall DNA structure.

Effect of 5hmC on DNA Structure

Renciuk *et al.* (116) showed that the presence either 5mC or 5hmC does not influence the overall B-DNA structure of the DDD. Our solution NMR data corroborates the crystallographic data (116), as can be seen from a comparison of NOESY spectra of the modified and unmodified duplexes. For example, the sequential pattern of imino-imino and imino-amino NOEs is conserved in the presence of the 5hmC:G base pair, confirming that the base pairing and base stacking is maintained at the lesion sites (Figure 3.7). In addition, the pattern of sequential base aromatic→deoxyribose anomeric NOEs is also conserved (Figure 3.8), with no significant changes in chemical shifts, which indicates the presence of a normal undisrupted B-type helix. Our crystal structure confirms that the 5hmC:G base pair imparts a minimal effect on the conformation of the DDD duplex (Figures 3.10 and 3.11) and does not significantly impact Watson-Crick base pairing and base stacking (Figure 3.12) as compared to the unmodified C:G base pair. Our crystallographic data indicate that the hydroxymethyl moieties on the modified cytosines are in the major groove, oriented toward the 3' end of each strand. The hydroxyl groups interact with *O6* and *N7* of the 3'-flanking G^{22} via two water molecules (Figure 3.11). One hydrogen bonding interaction was observed between the hydroxyl group at the modified cytosine Z^{21} and axially coordinated water (HOH 12) within 2.7 Å, with a further interaction to $G^{22} N^7$ (2.8 Å). A second hydrogen bonding interaction was observed between the Z^{21} hydroxyl group and $G^{22} O6$ via water HOH 11 (3.0 Å distance from Z^{21} to HOH11, and 2.7 Å from HOH11 to $G^{22} O6$). Thus, the presence of these two water molecules might explain the low base pair opening rates for modified $G^4:Z^{21}$ base pair.

Structure-Activity Relationships

It has been proposed that active demethylation could involve oxidation of 5mC to 5hmC, and then to 5fC and 5caC, facilitated by TET enzymes, where 5caC in the last step undergoes decarboxylation to an unmodified C (104, 125). Maiti and Drohat showed that TDG might be involved in demethylation when 5fC and 5caC, but not 5hmC, are removed via BER (115). While thymine DNA glycosylase (TDG) removes 5fC and 5caC, it is unable to remove 5hmC (115). They proposed that TDG activity is modulated by the electron withdrawing properties of the substituent at the 5-position of the cytosine ring, such that strongly electron withdrawing C5-substituents stabilize the developing negative charge in the base excision transition state (127). They concluded that the negligible electronic effect for the hydroxymethyl group ($\sigma_m = 0$) might explain why 5hmC is not excised by TDG. Similarly, 5mC ($\sigma_m = -0.07$) and cytosine ($\sigma_m = 0$) are not removed via BER (115, 127). Studies using NMR imino proton exchange measurements by Stivers and Song (54) showed that uracil DNA glycosylase (UDG) can substantially increase the equilibrium constant for opening of A:T base pairs relative to free DNA, which could provide a dynamic mode of modification identification by DNA glycosylases that require the lesion to be extrahelical. Our results show that the equilibrium constant for opening of 5hmC is low. Thus, if the Stivers model is correct, 5hmC should not be a good substrate for BER.

A role of 5hmC as an epigenetic modifier and transcriptional regulator has been proposed, with altered levels in early embryonic development, embryonic stem cell

differentiation, and tumors (109, 113). It was reported that the level of methylation depends on cell or tissue type and developmental stage (128). After fertilization, during embryonic development, many of the methylation markers are erased. This allows embryonic stem cells to differentiate into specialized cells. It is now known that 5hmC and TET deoxygenases play a crucial role in epigenetic reprogramming and regulation of tissue-specific gene expression (129-132). However, it was proposed that 5hmC may only be a reaction intermediate in the process of active demethylation (39, 40).

Acknowledgements

This work was supported by NIH grant R01 CA-55678 (M.P.S.). Funding for the NMR instruments was supplied by NIH grants S10 RR-05805, S10 RR-025677 and NSF Grant DBI 0922862, the latter funded by the American Recovery and Reinvestment Act of 2009 (Public Law 111-5). Vanderbilt University assisted with the purchase of in-house crystallographic and NMR instrumentation.

CHAPTER 4

CHARACTERIZATION OF 5-FORMYCYTOSINE IN DNA

Introduction

5-formylcytosine (5fC) is an oxidation derivative of 5-methylcytosine formed by TET enzymes in the process of active demethylation (39, 133). The TET family of proteins has the capacity to convert 5mC to 5fC *in vitro*. The presence of 5fC was found in the genomic DNA of mouse embryonic stem (ES) cells and organs (39). Genome-wide mapping in mouse ES cells showed high levels of 5fC in CpG islands of promoters and exons, which corresponded to transcriptionally active genes. Based on these studies it was proposed that 5fC plays a key role in reprogramming with specific genomic regions, that are controlled by the base excision mechanism and TDG. Excision of 5fC in ES cells is crucial for correct establishment of CpG methylation patterns during differentiation and for appropriate patterns of gene expression during development (134). Recently, the functional effect on transcription of 5fC in the genome has been described, for mammalian and yeast RNA polymerase II. Pol II polymerization rates and specificity constants for GTP incorporation against 5fC were reduced significantly, while there were no changes in GTP incorporation opposite C, 5mC or 5hmC templates. Thus, Pol II can read and distinguish subtle changes at the 5 position of modified cytosines and process them in different ways (135).

In the process of active demethylation of 5mC, 5fC is removed by thymine DNA glycosylase (TDG) via the base excision repair mechanism, resulting in the formation of unmodified cytosine (78). It was shown that TDG can rapidly excise 5fC from DNA in vitro, and this activity was subsequently found in mammalian cells (136-139). In order to understand the mechanisms of active demethylation through 5fC formation, chemical reactivity studies of the newly formed nucleobases were reported (140). In this study, the sensitivity of these modifications to oxidation and deamination was measured, along with C-C bond cleaving reactivity, either in the absence and presence of thiols as biologically relevant (organo) catalysts. These studies revealed that 5hmC is rapidly oxidized to 5fC, in comparison with 5mC, and the deamination reaction occurred only at a minimal level. Moreover 5fC can undergo thiol-mediated and acid-catalyzed C-C bond cleavage reactions to form unmodified dC with the release of formic acid. Thus if the DNA demethylation occurs via 5hmC oxidation to 5fC, then demethylation could take place via alternative active demethylation mechanisms (140). However, recent biochemical and biological studies have established that the pathway for active DNA demethylation involves BER and TDG enzyme. The latest studies of Maiti and Drohat report the investigation of the TDG mechanism excising 5fC from DNA, and the chemical properties indicating the catalytic requirements for the excision (78, 141). They showed that TDG can rapidly remove 5fC, with higher activity than T from G/T mismatches (78). TDG activity is greater for cytosine analogs with an electron-withdrawing C5-substituent ($\sigma_m > 0$, σ_m - electronic substituent constant) that can stabilize negative charge developing on the excised base in the chemical transition state. As the formyl group is strongly electron-withdrawing ($\sigma_m > 0.35$), it implies that TDG

could remove it. The next aspect considered by the authors was the tautomerism of 5fC under physiological conditions. It was proposed that 5fC favors an imino tautomer and adopts a Wobble-like structure, similar to the structure of G/U or G/T mispairs (Figure 4.1) (142, 143).

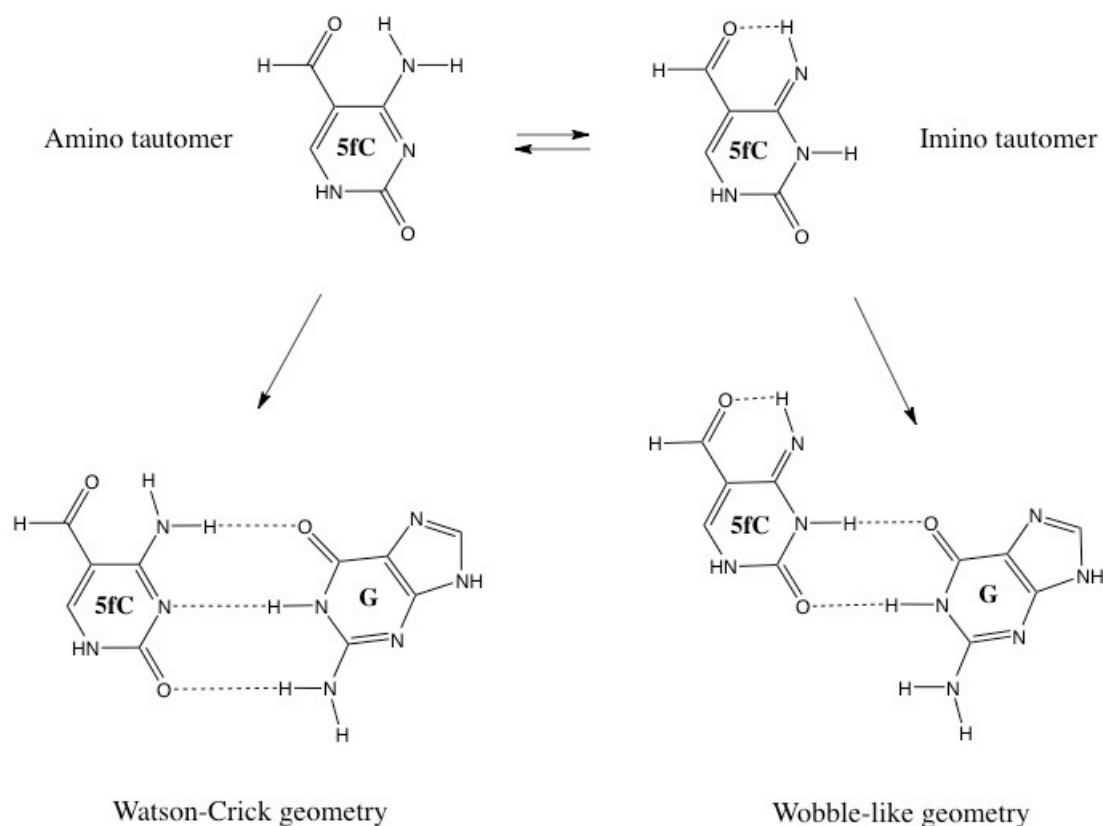


Figure 4.1. Tautomerism of 5fC modified base and base pairing schemes.

In the studies with TDG it was proposed that this unique structural geometry of a mismatched base pair is recognized by the enzyme, which results in excision of the mispaired base by TDG. The hypothesis was tested by Maiti and Drohat recently (141), where they calculated the relative stability of the amino and imino tautomers for 5fC in single base form. In their findings, 5fC as an anionic tautomer

is much more stable than its imino counterpart in the gas phase and in the water phase (141), suggesting that 5fC when paired with G forms a Watson-Crick base pair with normal geometry. In addition, NMR studies in DMSO showed the amino tautomer of 5fC was the predominant form (144). In the present work we will address their hypothesis and investigate duplex DNA containing 5fC modified base paired with G (DDD^f) by X-ray crystallography.

DNA glycosylases use an extrahelical base recognition mechanism, which relies on kinetically enhanced base pair opening rates for destabilized base pairs (82). High-resolution proton exchange NMR spectroscopy was performed to characterize the kinetics of base-pair opening in nucleic acid duplexes. The rates of exchange of imino protons with solvent protons were measured by magnetization transfer from water for each DNA duplex as a function of the concentration of exchange catalyst. In work presented here the dynamics of base pair opening in DDD^f in comparison with canonical DDD is discussed.

Results

The sequence studied in this work is well-characterized Dickerson-Drew Dodecamer, which has C/G rich termini and A/T rich core. This sequence was selected because it is self-complementary, gives well-resolved peaks in NMR spectra, and crystallizes well. The 5fC modification was incorporated into the DDD oligodeoxynucleotide at the ninth position; consequently, after annealing the duplex

were contains two modified bases. The DNA duplexes used in these studies are presented in Figure 4.2.

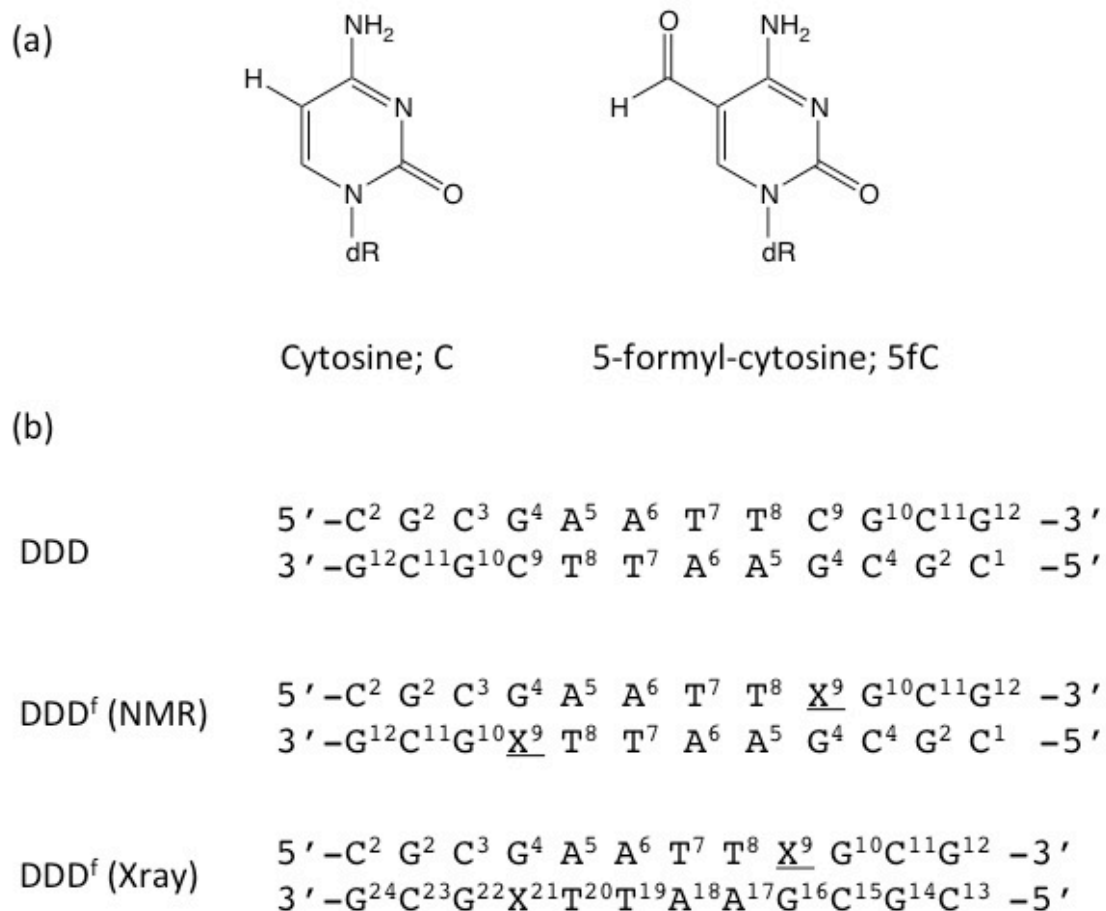


Figure 4.2. (a) Structure of dC, and 5fC. (b) Sequences and numbering of the nucleotides for unmodified DDD, and DDD^f (NMR) and DDD^f (X-ray) duplexes. In solution, the two strands of the DDD exhibit pseudo-dyad symmetry and the NMR resonances of symmetry-related nucleotides in the two strands are not individually observed. In the crystal, electron density for symmetry-related nucleotides in the two strands is observed and the nucleotides are numbered individually.

Thermal denaturation studies

The melting temperature of DDD and DDD^f was examined by temperature-dependent UV spectroscopy, and taking the first derivative of the resulting UC melting curves. The T_M of DDD^f was 46 °C, and for the unmodified DDD was 48 °C, measured at the same conditions.

NMR melting studies

A series of 1D ¹H NMR spectra for the exchangeable protons were recorded at 5, 15, 25, 35, 45, and 55 °C and are shown in Figure 4.3. The data show that the N^1 -imino proton of the G²·C¹¹ base pair in DDD^f was sharp and detectable only at 5 °C, and broadened at 15 °C. The same imino peak in DDD started to broaden at 35 °C. The next base pair, G¹⁰·C³, which is the 5' neighbor of the modification, almost disappeared in 35 °C in DDD^f, while it is still sharp even at 45 °C for the control experiment. The N^1 -imino proton of the X⁹·G⁴ modified base pair in DDD^f remained sharp even at 35 °C, and started to broaden at 45 °C; but the same resonance in control DDD was still sharp and detectable at a temperature as high as 55 °C. The N^3 -imino resonances of T⁷ and T⁸ are sharp in DDD at 55 °C, while these peaks are gone in DDD^m. There was only one observed change in the chemical shift for the G⁴ imino proton in the DDD^f which was upfield by 0.5 ppm when compared to G⁴ in the unmodified DDD.

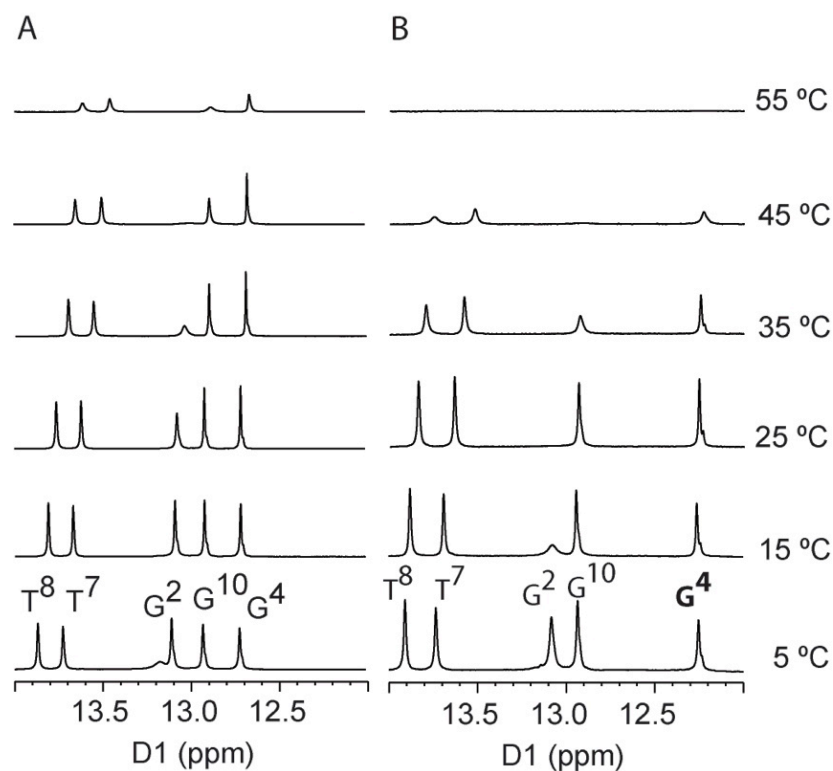


Figure 4.3. ¹H-NMR of imino proton resonances as a function of temperature for the unmodified duplex (A), the 5fC modified duplex (B).

Exchangeable protons.

Figure 4.4 shows the NOE connectivity of the purine N^1 and pyrimidine N^3 imino protons. The base imino protons were assigned based on their sequential connectivities in NOESY spectra, and these assignments were supported by their NOE cross-peaks to their Watson-Crick base-paired amino protons. The sequential connectivities were obtained from base pairs $G^2:C^{11} \rightarrow G^{10}:C^3 \rightarrow G^4:C^9 \rightarrow T^8:A^5 \rightarrow T^7:A^6$. For the DDD and DDD^f duplexes the imino-proton resonances of the terminal base pairs $C^1:G^{12}$ are lost by fast exchange with water. The imino resonance from G^4 , which is base paired with X^9 was as intense as other imino peaks in this region. However, the G^4 imino peak was shifted upfield by 0.5 ppm, reflecting the effect of base pairing opposite X^9 .

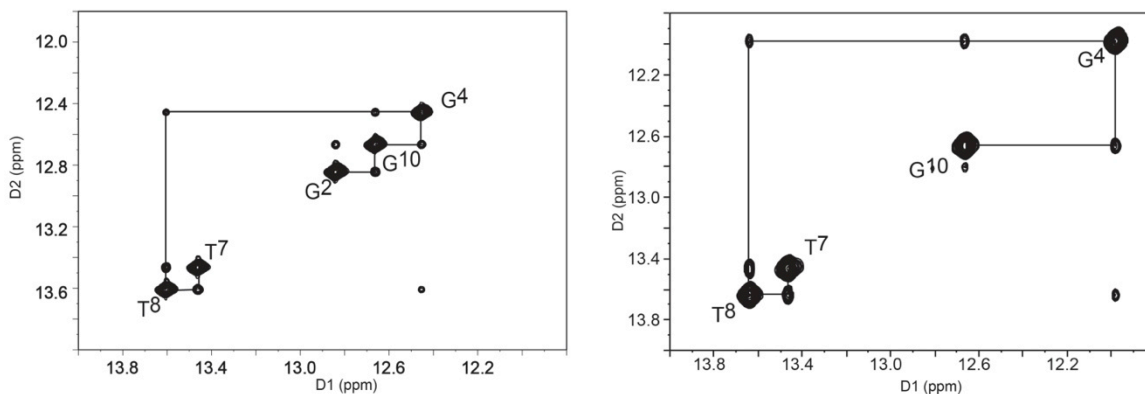


Figure 4.4. ^1H - ^1H NMR NOESY spectrum showing resonances for the thymine and guanine imino protons and sequential NOE connectivity for the imino protons of the base pairs $\text{G}^2:\text{C}^{11}$ to $\text{A}^6:\text{T}^7$ for the unmodified (left), and the 5fC-modified (right) duplexes.

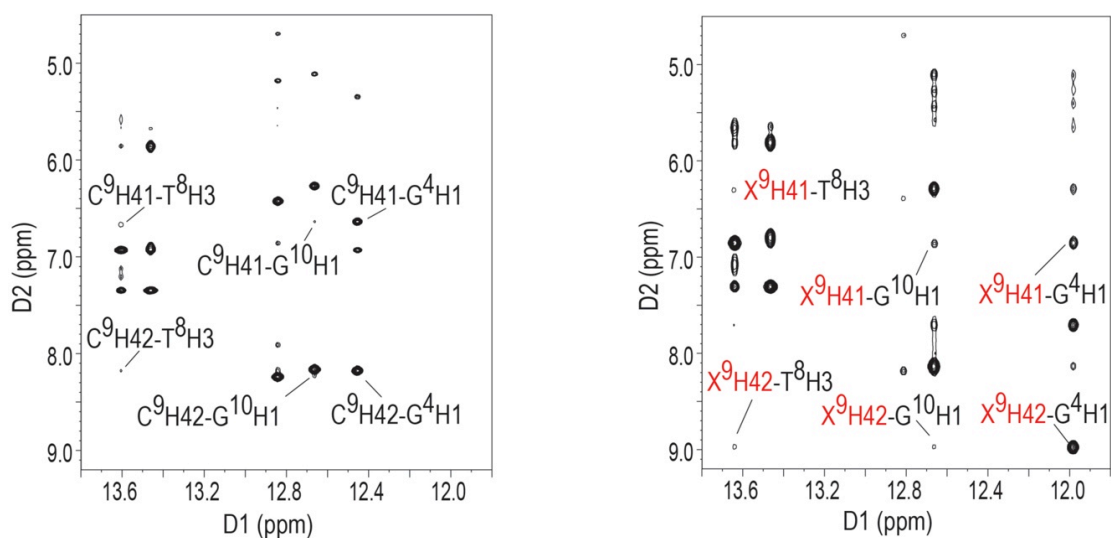


Figure 4.5. Expansion of the ^1H - ^1H NOESY spectra for DDD (left), and DDD^f (right) showing the conservation of Watson-Crick base pairing and base stacking at the modification site.

The upfield region of the NOESY spectrum (Figure 4.5) showed the NOEs between the imino and amino protons. Cross peaks from modified base C⁹H41, C⁹H42 to opposite base G⁴H1 were observed, as well as interactions to neighbor bases T⁸H3 and G¹⁰H1.

Nonexchangeable protons

The sequential assignment of nonexchangeable protons was accomplished using standard protocols. The unmodified duplex was used as a control for NMR assignments for DDD^f. For modified duplexes, the anticipated pattern of sequential base aromatic → deoxyribose anomeric nuclear Overhauser enhancement (NOE) was identified from C¹ → G¹² (Figure 4.6).

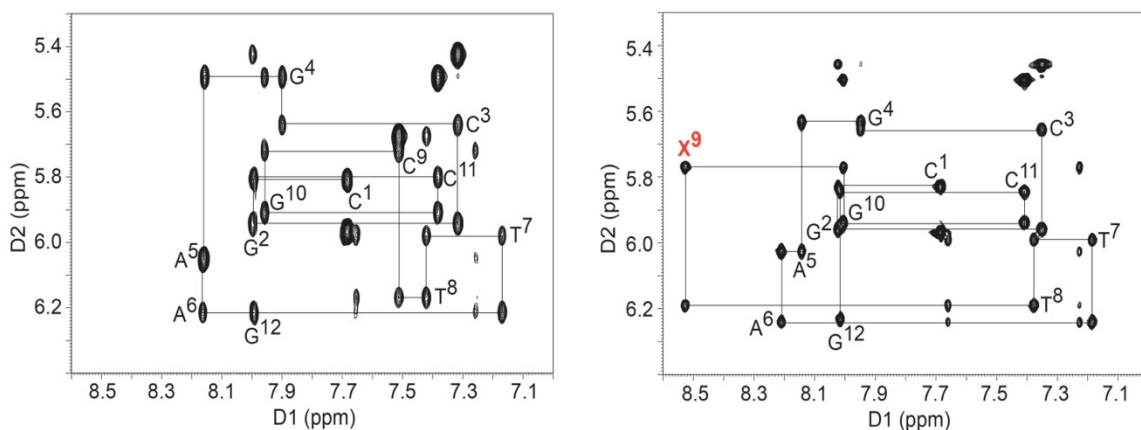


Figure 4.6. Expanded plot from the aromatic-anomeric region of the NOESY spectrum, showing sequential NOE connectivities of unmodified DDD, and modified DDD^f duplexes.

Only one set of resonances was observed because the sequences are self-complementary. All spectra exhibited well resolved cross-peaks. Three strong NOEs accounted for H5-H6 cross-peaks of cytosine residues (C¹, C³, and C¹¹). The H5-H6 resonance from X⁹

is missing due to formyl substitution at 5 position in DDD^f. Cytosine assignments were confirmed by a DQF-COSY spectrum recorded at identical conditions (data not shown). Each base proton exhibited NOE peaks to its own and the 5'-flanking H1'-deoxyribose protons. For T⁸ and G¹⁰ the NOE cross-peaks intensities between the base protons and the sugar H1' of the attached deoxyribose moieties were of the same relative magnitudes as those between other bases in the sequence. The 5fC H6 resonance was observed at 8.5 ppm, shifted downfield by approximately 1 ppm with respect to that of the unmodified oligodeoxynucleotide. This was attributed to the differential electronic density for 5fC as compared to G. Proton resonances from the opposite G⁴ and A⁵ bases exhibited chemical shift changes of <0.1 ppm, compared with those bases in the unmodified DDD duplex.

Crystal structure of DDD with 5fC

Crystals belonged to the orthorhombic $P2_12_12_1$ space group with unit cell parameters $a = 25.32 \text{ \AA}$, $b = 41.47 \text{ \AA}$, $c = 65.66 \text{ \AA}$. The unmodified DDD (PDB entry 436D) was used as a search model for molecular replacement. The crystal structure of DDD^f was refined isotropically to a resolution of 1.74 \AA . The rmsd values for bond lengths were 0.009 \AA , and 1.747 deg for angles. The structure overlaid well with the canonical DDD (PDB entry: 436D (77)) structure with a rmsd of 0.329 \AA and no structural perturbations were observed at the two modification sites (Figures 4.7). At the sites of modification, the 5fC formyl groups were in the major groove (Figure 4.8). They remained in the plane with the modified cytosine bases, thus potentiating formation of hydrogen bonds to the exocyclic amino groups at the C4 position of modified

cytosines. For both modified base pairs, the base stacking pattern in the modified DDD^f was similar to that observed in the unmodified DDD (Figure 4.9). The helical parameters calculated with the program Curves+ (100) were comparable to the unmodified DDD and are presented in the Figures 4.11 – 4.13.



Figure 4.7. Sum electron density contoured at the 1.0σ level (green meshwork) surrounding the DDD^f duplex at the modification site. The 5fC modified base is shown in red, and water molecules are shown in dark red.

The substitution by formyl at the C5 position of cytidine, did not disrupt Watson-Crick base pairing with G or change the conformation of these residues (Figure 4.9).

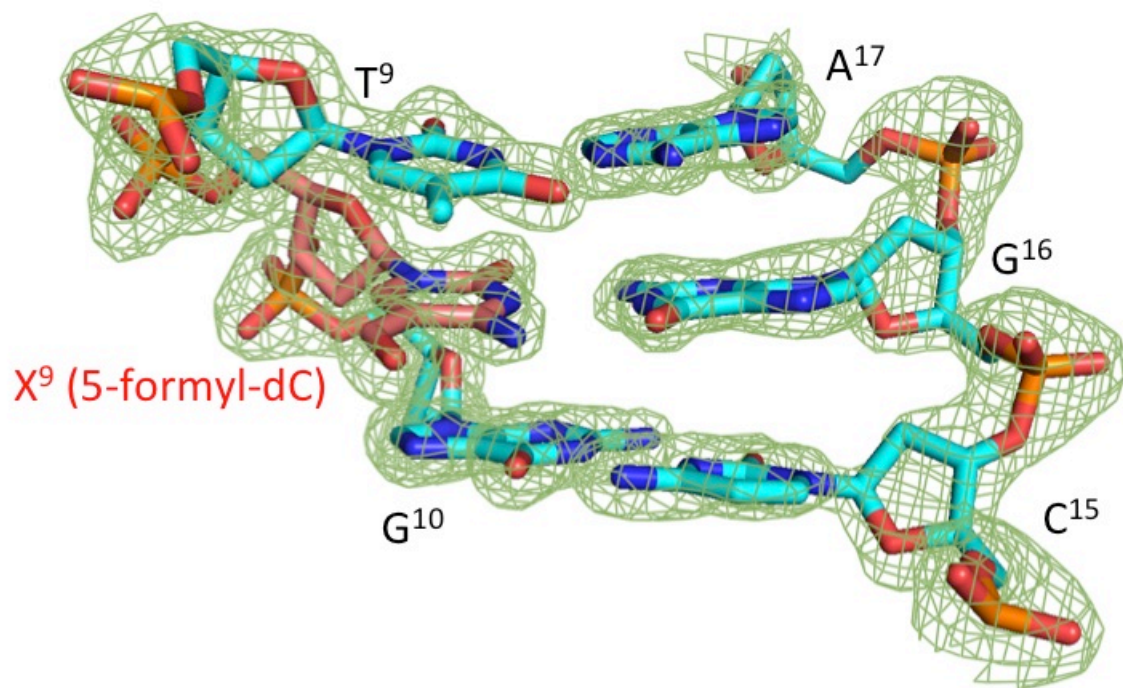


Figure 4.8. Fourier ($2F_o - F_c$) sum electron density contoured at the 1.0σ level (green meshwork) surrounding the DDD^f duplex at the modification site.

Analysis of the waters forming the “spine” of hydration, shows that the waters involved in the minor groove hydration are conserved. Detailed information about data collection and refinement statistics are shown in Table 2.2.

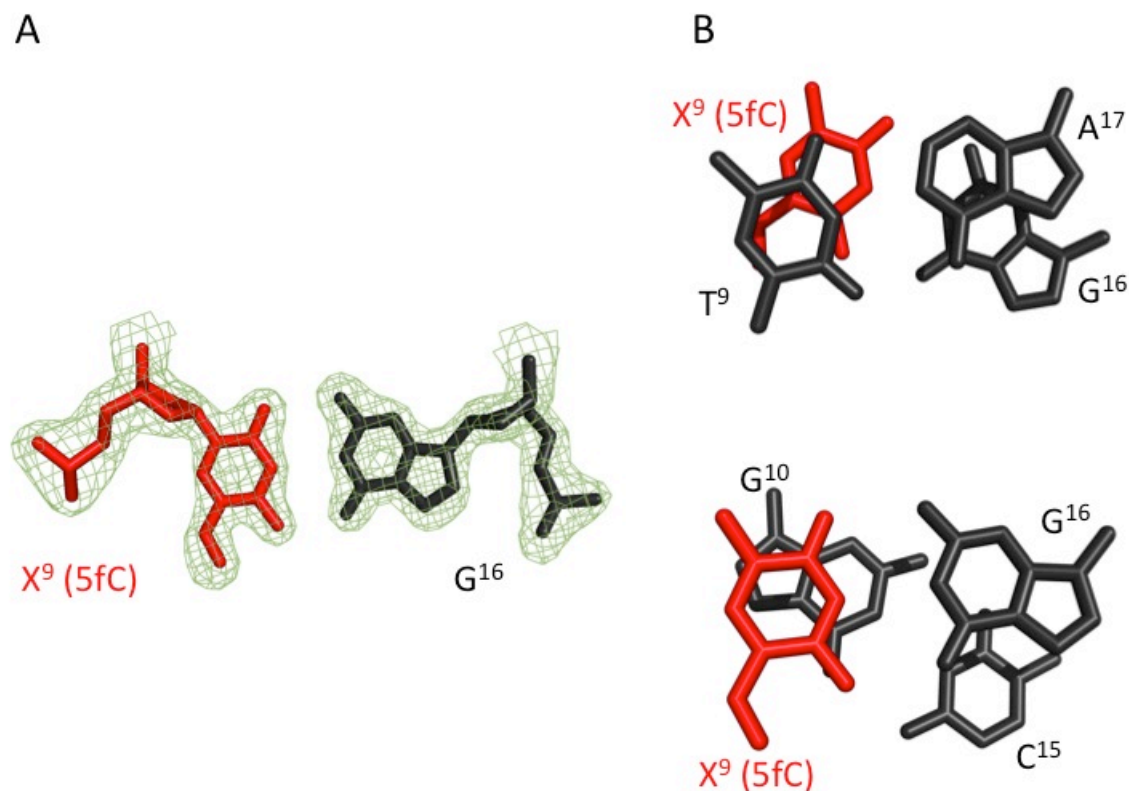


Figure 4.9. (A) Sum electron density contoured at the 1.0σ level (green meshwork) around the 5fC:dG modified base pair showing conserved Watson-Crick base pairing geometry. (B) Expanded view of the DDD^f crystal structure, showing stacking interactions: (top) stacking of base pair T⁸:A¹⁷ above base pair X⁹:G¹⁶, and (bottom) stacking of base pair X⁹:G¹⁶ above base pair G¹⁰:C¹⁵.

Dynamics of base pair opening

To determine the dynamics of base pairs, imino proton exchange rates were measured in the presence of ammonia base catalyst, that was used because of its small size, lack of charge and high accessibility factor. Figure 4.10 shows imino proton exchange rates obtained by measuring magnetization transfer from water as a function of added ammonia base catalyst. The results of the imino proton exchange analysis are summarized in Table 4.1. For the terminal (i.e. C¹:G¹²) and penultimate

(G²·C¹¹) base pairs the exchange rates could not be measured because of the increased exchange with solvent protons.

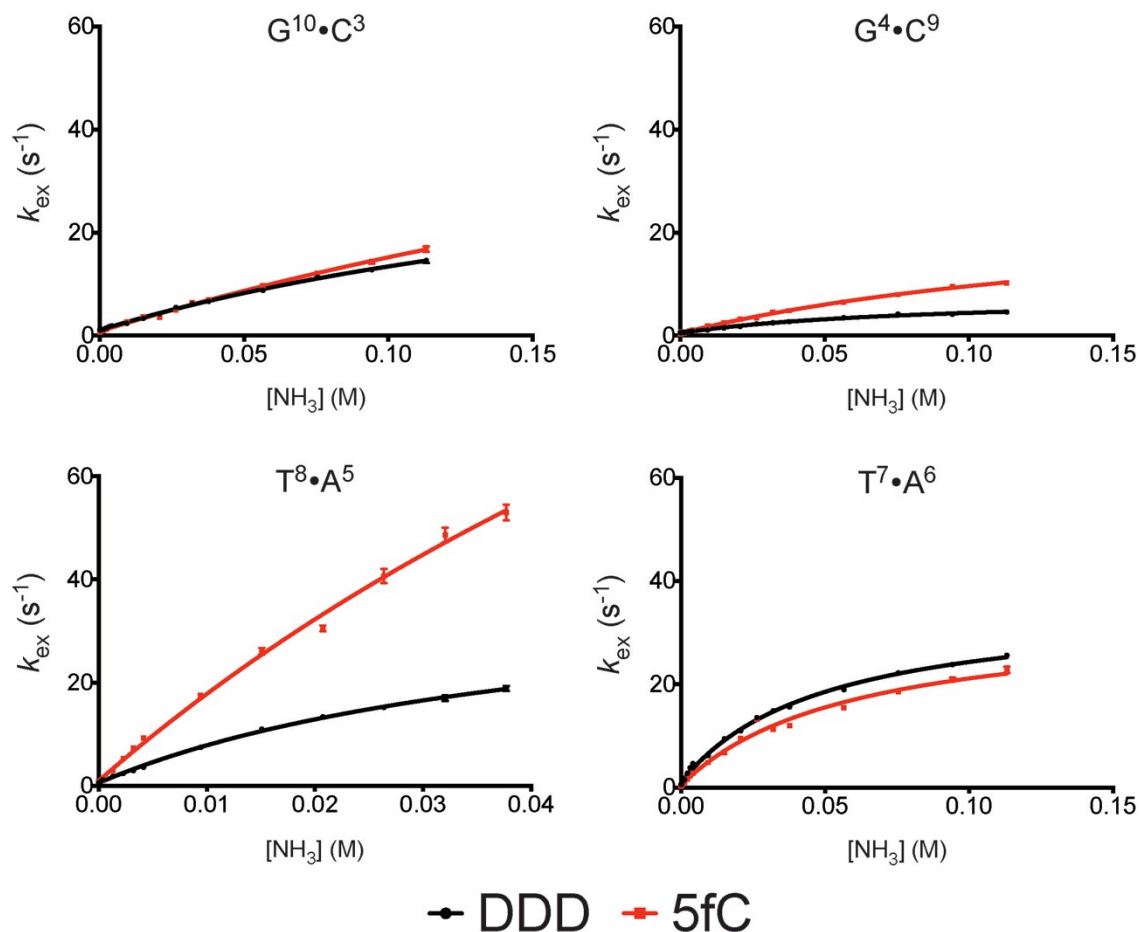


Figure 4.10. Plots showing imino proton exchange rates obtained by monitoring magnetization from water as a function of ammonia base catalyst. A. G¹⁰:C³. B. G⁴:C⁹. C. T⁸:A⁵. D. T⁷:A⁶ in DDD (black), and DDDf (red).

As is evident, for all of the studied imino protons in the three duplexes studied, the exchange rate reaches the EX1 regime as predicted by the eq. 9. As the concentration of ammonia increases, the exchange rates increase until a plateau is reached, which indicates a change in the rate limiting step from chemical exchange to base pair opening ($k_{\text{ex}} = k_{\text{op}}$ and exchange occurs in each opening event). For the imino proton of G⁴,

the exchange rate has a weak dependence on the concentration of ammonia base. This finding reflects the small equilibrium constant for opening of the G⁴·C⁹ base pair. However, comparing G⁴ from DDD^f with G⁴ from DDD, indicates that the opening rate is 3 times higher ($k_{\text{op}}=26$ vs. $k_{\text{op}}=8$, for G⁴ in DDD^f and DDD, respectively). The similar, low rate for base pair closing and base pair lifetime is consistent with the modified base pair spending more time exposed to solution than being embedded in a canonical base. This is also confirmed by the equilibrium constant for base pair opening, which is almost tripled for modified G⁴:X⁹ vs. unmodified G⁴:C⁹, ($\alpha K_{\text{op}} = 2.8$ and 1.2, respectively).

	k_0 (s ⁻¹) ^a		$K_{\text{op}} \times 10^7$	
	DDD	DDD ^f	DDD	DDD ^f
C ³ :G ¹⁰	1.1 ± 0.06	0.71 ± 0.03	2.9 ± 0.1	3.8 ± 0.1
G ⁴ :C ⁹	0.57 ± 0.03	0.39 ± 0.04	1.2 ± 0.04	2.8 ± 0.1
A ⁵ :T ⁸	0.59 ± 0.03	0.79 ± 0.04	41 ± 0.08	106 ± 4
A ⁶ :T ⁷	0.61 ± 0.03	0.27 ± 0.03	37 ± 5	29 ± 2

	k_{op} (s ⁻¹)		$k_{\text{cl}} (\times 10^{-7} \text{ s}^{-1})$	
	DDD	DDD ^f	DDD	DDD ^f
C ³ :G ¹⁰	45 ± 3	86 ± 18	15 ± 1	21 ± 2
G ⁴ :C ⁹	8 ± 0.5	26 ± 2	6.7 ± 0.2	9.3 ± 1
A ⁵ :T ⁸	40 ± 2	222 ± 53	0.97 ± 0.04	2.1 ± 0.06
A ⁶ :T ⁷	36 ± 1	33 ± 3	0.98 ± 0.09	1.1 ± 0.05

Table 4.1. Rate and Equilibrium Constants for DDD^f Base Pair Opening.
^aThe observed exchange rate without an ammonia catalyst.

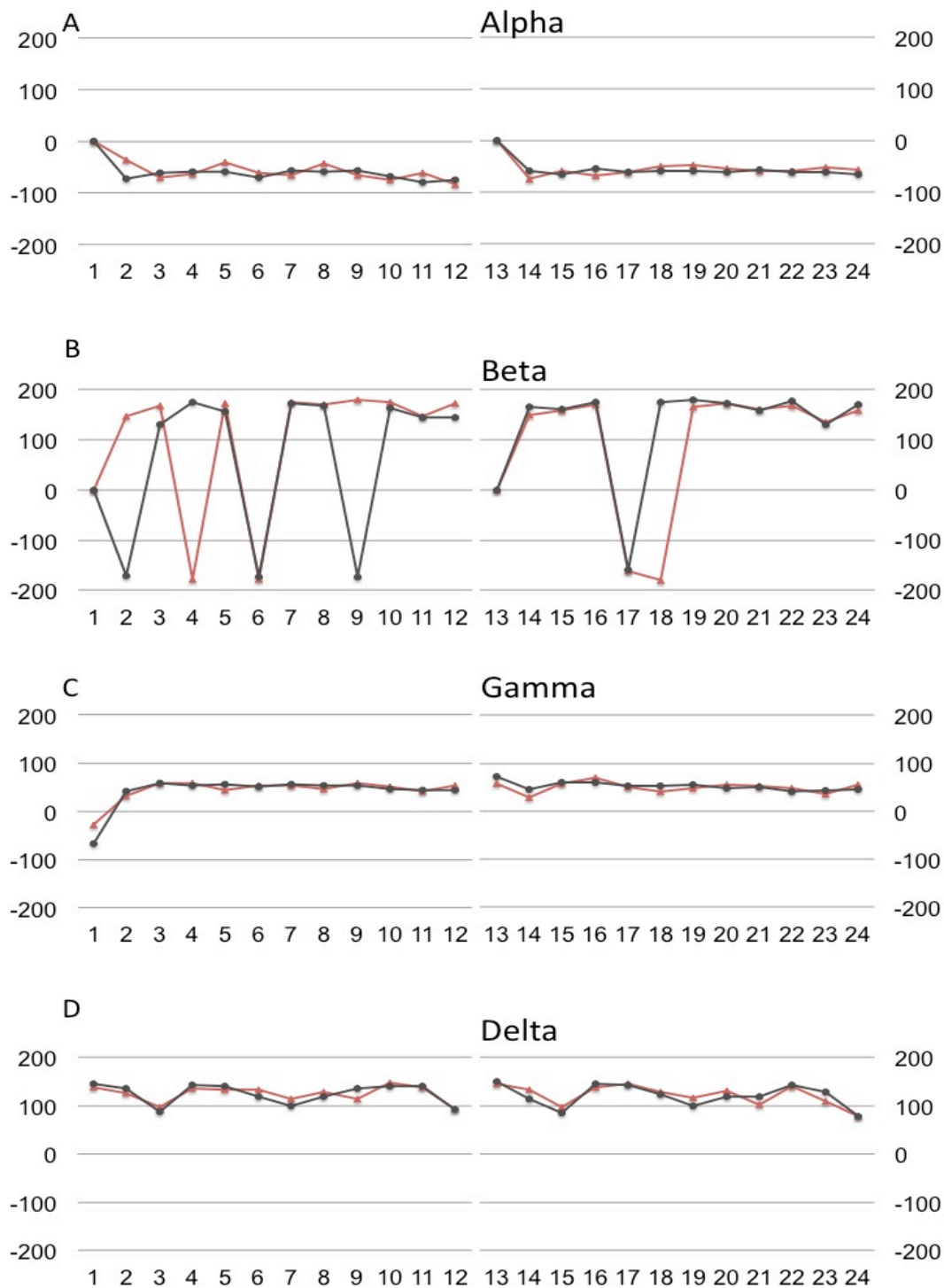
For G¹⁰·C³ imino protons, the exchange rates are faster even at low ammonia concentrations. With an increase in the catalyst concentration, the exchange rates further

increases until the EX1 regime is reached. The equilibrium constant for base pair opening is slightly higher for the modified base pair in the DDD^f relative to the native DDD ($\alpha K_{op} = 3.8 \times 10^7$ vs. 2.9×10^7 for $G^{10}:C^3$). However analysis of base pair opening shows twice the rate for $G^{10}:C^3$ in DDD^f , relative to the same base pair in the unmodified duplex (Table 4.1). The highest value for the equilibrium constant for base pair opening is for T^8 imino proton, which is the 5'- neighbor of the modification site. This applies to both duplexes. The dynamics of this base pair is remarkably different in the DDD^f duplex. As compared to $T^8:A^5$ in the DDD , this base pair in DDD^f opens ~ 6 -times faster, and has open lifetimes ($1/k_{cl}$) that are only 2-times shorter. A much higher rates for the equilibrium constant for base pair opening suggest that $T^8:A^5$ in DDD^f spends 2.5-fold more time in the open conformation than the corresponding base pair in the control duplex.

Helicoidal analysis

An analysis of the helical parameters of the crystal structure was performed with the program Curves+ (100). The analysis of the torsion angles (shown in Figure 4.11) demonstrates that there were no significant changes in the alpha, gamma, delta, epsilon and zeta angles. For the beta torsion angle, the DDD^f shows very similar conformation as the unmodified DDD (77). However, the peak at G^4 with the DDD^f reflects a change as a result of the presence of the modification opposite base X^9 . The analysis of interbase parameters shows no substantial changes in helical rise, roll and twist, as presented in Figure 4.12. Also, shear and stretch, which are intrabase

translation parameters, are similar and comparable between bases in all studied duplexes (Figure 4.13).



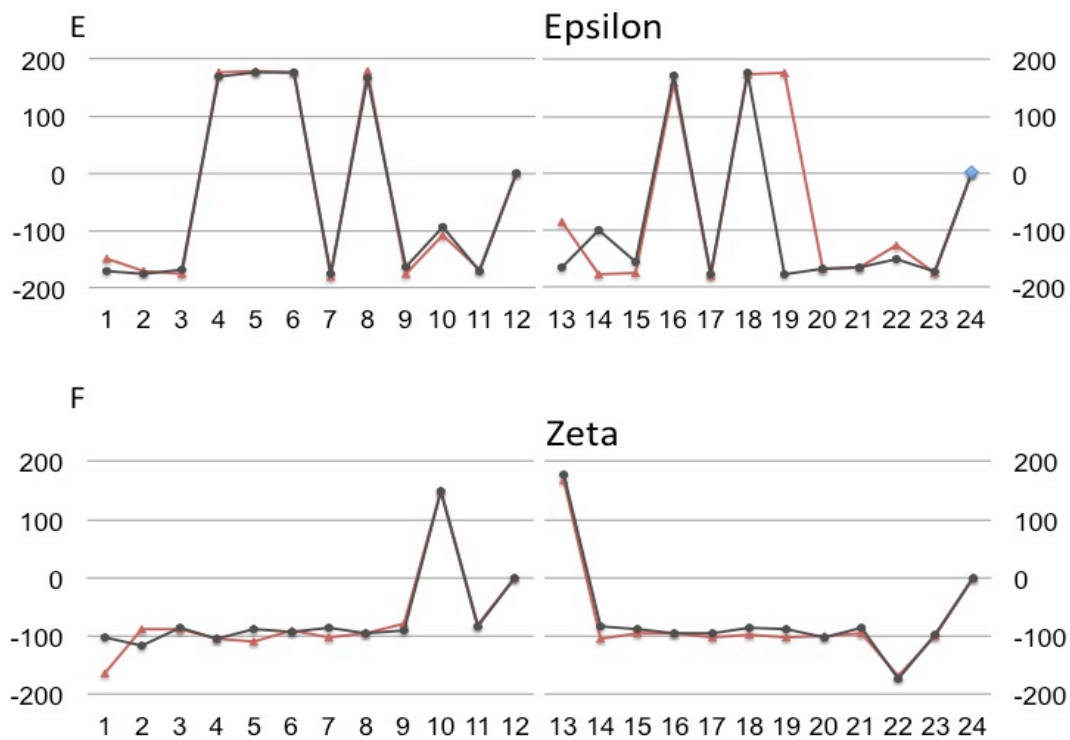


Figure 4.11. Comparison of backbone torsion angles (a) alpha, (b) beta, (c) gamma, (d) delta, (e) epsilon, and (f) zeta in the structure of DDD^f in red, and unmodified DDD (PDB code 436D (77)) in black.

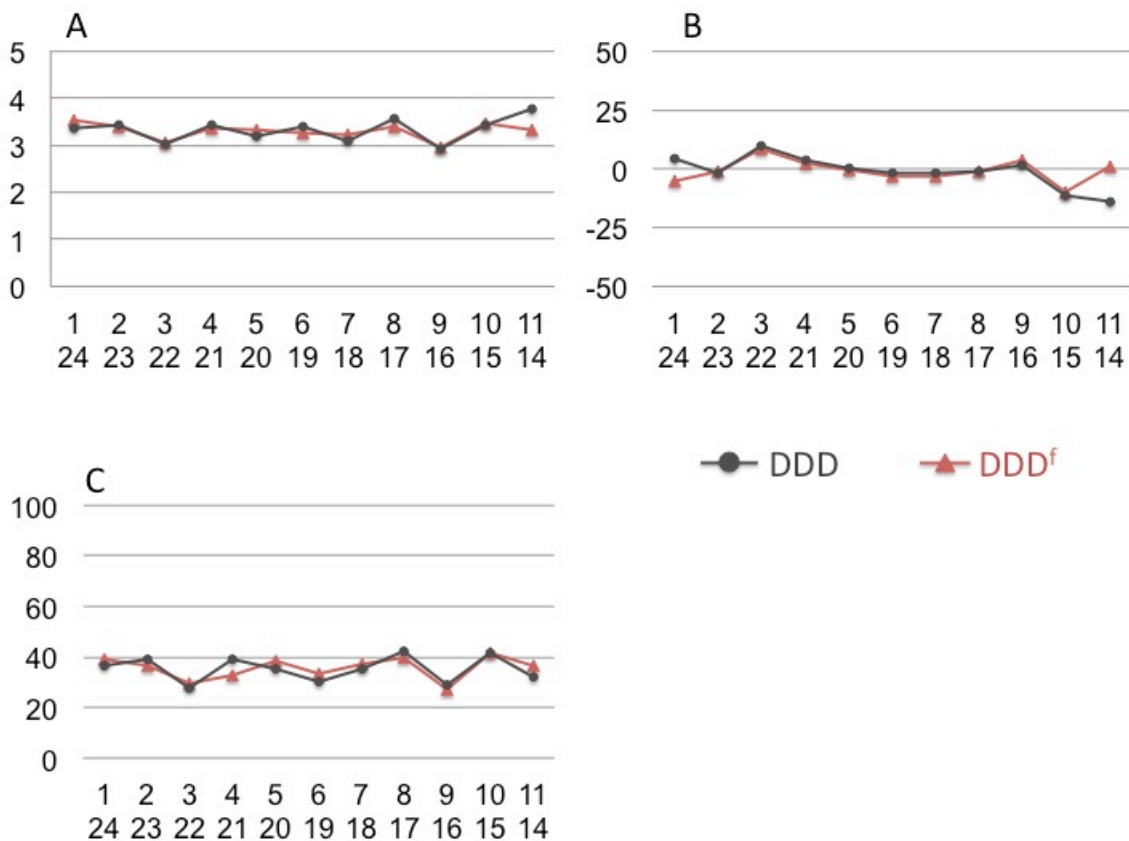


Figure 4.12. Interbase pair parameters: (a) helical rise, (b) roll, and (c) twist for the structure of DDD^f in red and unmodified DDD (PDB code 436D (77)) in black.

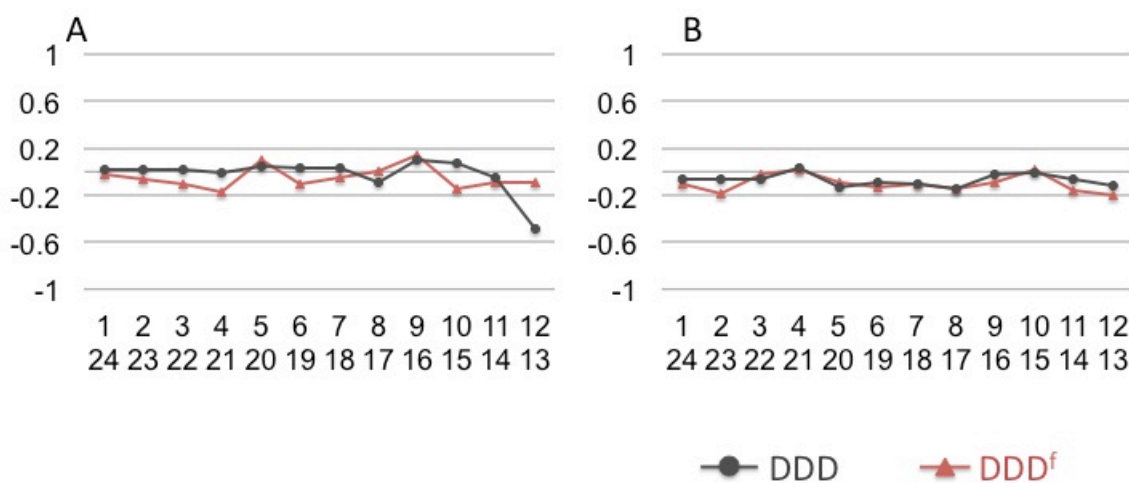


Figure 4.13. Intrabase pair parameters (a) shear, and (b) stretch for the structure of DDD^f in red, and unmodified DDD (PDB code 436D (77)) in black.

Discussion

The 5fC base is an active demethylation oxidation product derived from 5mC, formed post-replicatively by TET enzymes (39). The 5fC base has been detected in genomic DNA of mouse embryonic stem (ES) cells and organs (39, 133) at high levels in CpG islands of promoters and exons, which corresponded to transcriptionally active genes.

Effect of 5fC on DNA structure

TET deoxygenases act together with other DNA demethylation pathways, such as BER. In this process, 5fC is efficiently removed by thymine DNA glycosylase (TDG) resulting in the restoration of unmodified cytosine (78, 145, 146). Hashimoto *et al.* proposed that 5fC undergoes tautomerisation, which results in Wobble base pair formation. This unique base pair geometry was shown to be recognized by TDG in the BER pathway (57). In order to understand the basis by which 5fC pairs opposite guanine in DNA, the structure of 5fC:dG in DDD^f was determined. The structural data reveal that 5fC has a minimal effect on DNA conformation (Figure 4.8) when compared to the dC:dG base pair (77). The Watson-Crick base pairing geometry is conserved for the 5fC:dG base pair (Figure 4.9). There is no evidence in the electron density map for the imino tautomer of 5fC, which would result in protonation at the N3 position of the modified cytosine. This protonation would prevent the formation of a Watson-Crick base pair with the complementary guanine. In addition, the solution NMR data corroborate the crystallographic data, as can be seen from a comparison of NOESY spectra of the modified and unmodified duplexes. For example, the sequential pattern

of imino-imino and imino-amino NOEs is conserved in the presence of the 5fC:G base pair, confirming that base pairing and base stacking are maintained at the lesion sites (Figures 4.4 and 4.5). In addition, the pattern of sequential base aromatic→deoxyribose anomeric NOEs is also conserved (Figure 4.6), with no significant changes in chemical shifts, which indicates the presence of a normal undisrupted B-type helix.

Effect of 5fC on DNA stability

To investigate the thermal stability of 5fC in DNA we determined melting temperatures of the modified duplex by temperature-dependent UV spectroscopy. The T_M was calculated by the first derivative of the resulting UV melting curves. The resulting T_M of DDD^f is 2 °C lower than the T_M of the unmodified duplex, suggesting lower stability of formylated DNA. The same destabilizing effect is observed in temperature-dependent NMR spectroscopy (Figure 4.3) Overall, the NMR data show that all imino protons start to broaden at 45 °C in DDD^f, while the same resonances appear as sharp peaks in the control sequence at the same temperature. The G⁴ imino resonance, representing the modified base pair in DDD^f, broadened at 45 °C, but was still sharp and detectable at a temperature as high as 55 °C in the unmodified DDD.

Imino proton exchange and base pair opening kinetics

DNA glycosylase repair enzymes use an extrahelical recognition mechanism to bind and excise the damaged base from DNA (82, 147). In the series of studies done by Stivers *et al.*, it was shown that DNA stability and dynamics play a key role in the detection of the damaged base by the repair enzyme. The hypothesis is that the base pair with lower stability will have kinetically higher opening rates (54, 55, 148). In order to characterize the kinetics of base pair opening in nucleic acid duplexes,

we performed high-resolution proton exchange NMR spectroscopy, where by the rates of exchange of imino protons with solvent protons were measured by magnetization transfer from water for each DNA duplex as a function of the concentration of exchange catalyst. Comparing G^4 from the DDD^f with G^4 from DDD shows a 3 times higher opening rate ($k_{op}=26$ vs. $k_{op}=8$, for G^4 in DDD^f and DDD , respectively). Together with the similar, low rate for base pair closing and base pair lifetime, that means that in fact the modified base pair spends more time exposed to the solvent than the canonical base. This is also confirmed by the equilibrium constant for base pair opening, which is 2.3-fold higher for modified $G^4:X^9$ vs. unmodified $G^4:C^9$, ($\alpha K_{op} = 2.8 \times 10^7$ and 1.2×10^7 , respectively). This result implies that the possibility of 5fC adopting an extrahelical conformation for excision by thymine DNA glycosylase is high.

Summary

It was hypothesized that DNA glycosylases use an extrahelical mechanism to recognize damaged bases that may possess higher base pair opening rates and lower stability. In addition, structural characteristics recognized by TDG suggest that a Wobble base pairing is recognized geometry at the modification site. In our studies we showed that 5fC is stably stacked in DNA duplex, and that the melting temperature of the modified duplex is substantially lower than of the canonical duplex. Moreover, the dynamics at the modification site and the opening rates are higher, which suggests

that the probability of 5fC base flipped out the DNA duplex and adopting an extrahelical conformation to be recognized by repair glycosylase is high.

Acknowledgements

This work is based upon research conducted at the Advanced Photon Source on the Northeastern Collaborative Access Team beamlines, which are supported by a grant from the National Institute of General Medical Sciences (P41 GM103403) from the National Institutes of Health. Use of the Advanced Photon Source, an Office of Science User Facility operated for the U.S. Department of Energy (DOE) Office of Science by Argonne National Laboratory, was supported by the U.S. DOE under Contract No. DE-AC02-06CH11357.

This work was supported by NIH grant R01 CA-55678 (M.P.S.). Funding for the NMR instruments was provided by NIH grants S10 RR-05805, S10 RR-025677 and NSF Grant DBI 0922862, the latter funded by the American Recovery and Reinvestment Act of 2009 (Public Law 111-5). Vanderbilt University assisted with the purchase of NMR instrumentation.

CHAPTER 5

CHARACTERIZATION OF 5-CARBOXYL-CYTOSINE IN DICKERSON DREW DODECAMER

Introduction

5-carboxycytosine (5caC) is an oxidation derivative of 5mC, formed by the TET enzyme in the process of active demethylation (39, 133). The TET family of proteins has the capacity to convert 5mC to 5caC *in vitro*. The presence of 5caC in the genomic DNA was established in mouse embryonic stem (ES) cells (39). The functional effect on transcription of 5caC in the genome has been described, for mammalian and yeast RNA polymerase II. Pol II polymerization rates and specificity constants for GTP incorporation opposite 5caC were reduced significantly, although there were no changes in GTP incorporation opposite C, 5mC or 5hmC in the templates. Pol II can read and distinguish subtle changes at the 5 position of modified cytosines, and process them in different ways (135).

The TET-catalyzed oxidation is reminiscent of the thymine hydroxylase catalyzed conversion of thymine to iso-orotate, which was recently studied in the context of 5mC demethylation. It could be potentially achieved through a process similar to the conversion of thymine to uracil, which is achieved by conversion of thymine to iso-orotate decarboxylase (149, 150). However it is hypothetical pathway for DNA

demethylation, and the enzyme that is capable of decarboxylation of 5caC-containing DNA has not been identified yet. TET enzymes act together with other DNA demethylation pathways, such as BER. In this process, 5caC is removed by thymine DNA glycosylase (TDG) resulting in the formation of unmodified cytosine (78). It was shown, that TDG can excise 5caC from DNA in vitro, and this activity was subsequently found in mammalian cells (136-139). In order to understand the mechanism of active demethylation, that proceeds via 5caC formation chemical reactivity studies of the new nucleobases were reported (140). In this report, the sensitivity of these modifications toward the oxidation and deamination was measured, along with C-C bond cleaving reactivity, both in the absence and presence of thiols as catalysts. It turns out, that 5caC can undergo thiol-catalyzed and acid-catalyzed C-C bond cleavage reactions to form dC with the release of CO₂. This means that decarboxylation could take place as an alternative active demethylation pathway (151). Indeed, extensive studies by Schiesser *et al.* provided the first evidence that stem-cell nuclear extracts have the ability to decarboxylate 5caC (152).

However, recent biochemical and biological studies established that the pathway for active DNA demethylation involves BER and TDG enzyme. The latest studies of Maiti and Drohat report the investigation of the TDG mechanism excising 5caC from DNA, and the chemical properties indicating the catalytic requirements for the excision (78, 141). They showed that TDG can remove 5caC (78). TDG activity is greater for cytosine analogs with an electron-withdrawing substituent at the C5 position ($\sigma_m > 0$, σ_m - electronic substituent constant) that can stabilize negative charge developing on the excised base in the transition state. In the case of 5caC, σ_m poorly predicts

the electronic effect for the carboxyl group. At neutral or physiological pH, the carboxyl group exists in the deprotonated form with $\sigma_m = -0.10$ (indicating electron-donating effect). However, as reported by Maiti and Drohat, the anionic carboxyl substituent lowers the pK_a of cytosine by -0.3, suggesting a stabilization effect of COO^- by NH_2 group at position C4, which would indicate an electron withdrawing effect and implying that TDG could remove it (78, 141). The other aspect addressed by the authors was the tautomerism of 5caC under physiological conditions. It was proposed by Hashimoto *et al.* that 5caC favors an imino tautomeric conformation leading to a Wobble-like structure of the pair with G, similar to the structure of G/U or G/T mispairs (Figure 5.1) (142, 143).

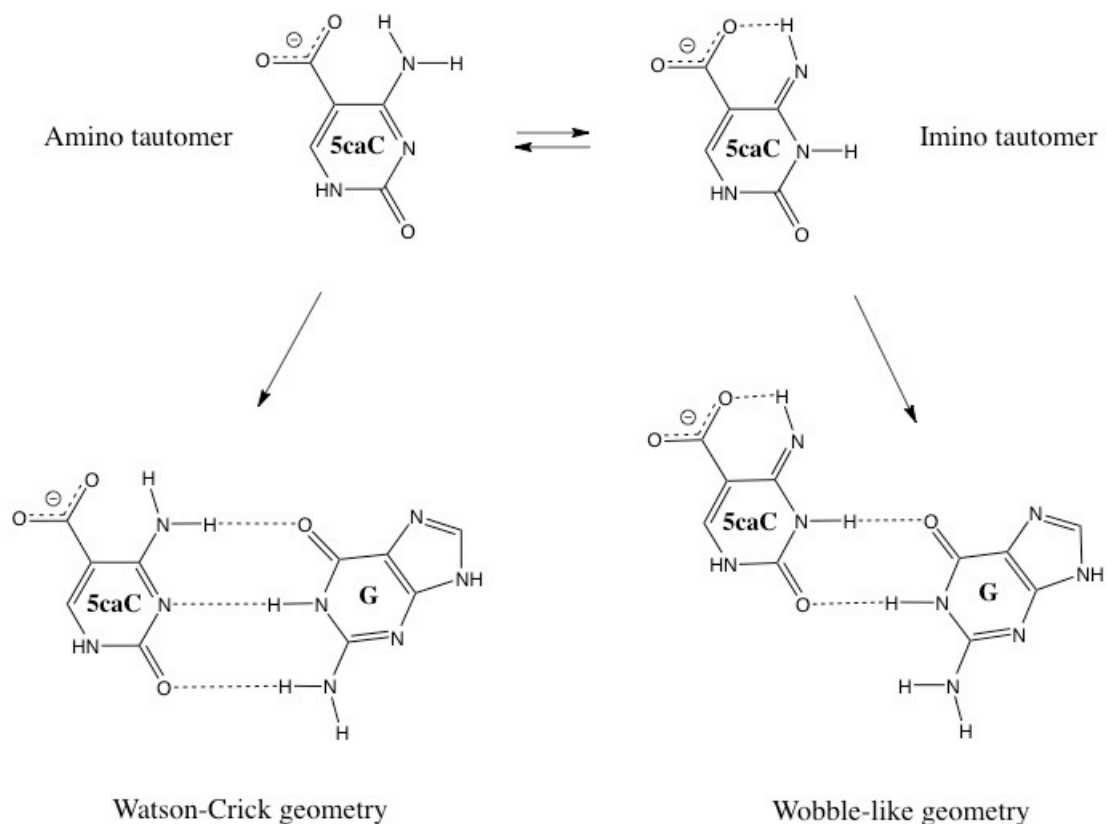


Figure 5.1. Tautomerism of 5caC modified base and base pairing schemes.

In the studies with TDG it was proposed that this unique structural geometry of the mismatch base pair is recognized by the enzyme, which results in excision of the mispaired base by TDG and repair. The hypothesis was tested by Maiti and Drohat recently, whereby they calculated the relative stability of the amino and imino tautomers for 5caC in a form of single base. Accordingly 5caC as an anionic tautomer is much more stable than its imino counterpart (Figure 5.1) in the gas phase and in the water phase (141), suggesting that 5caC when paired with G forms a Watson-Crick base pair with normal geometry. We addressed this hypothesis in the present work and investigated the stacking and stability of DNA containing a 5caC modified base paired with G.

DNA glycosylases use an extrahelical base recognition mechanism, which relies on kinetically enhanced base pair opening rates for destabilized base pairs (82). High-resolution proton exchange NMR spectroscopy was used to characterize the kinetics of base-pair opening in nucleic acid duplexes. The rates of exchange of imino protons with solvent protons were measured by magnetization transfer from water for each DNA duplex, as a function of the concentration of exchange catalyst. Here we discuss the potential of 5caC as a substrate for base excision repair.

Results

The sequence studied in this work is well-characterized Dickerson-Drew Dodecamer (76), which has C/G rich termini and A/T rich core. This sequence was selected because it is self-complementary and crystallized well. The 5caC modification was incorporated into the DDD oligodeoxynucleotide at the ninth position; consequently after annealing the modified duplex contains two 5caC residues. The DNA duplexes used in these studies are listed in Figure 5.2.

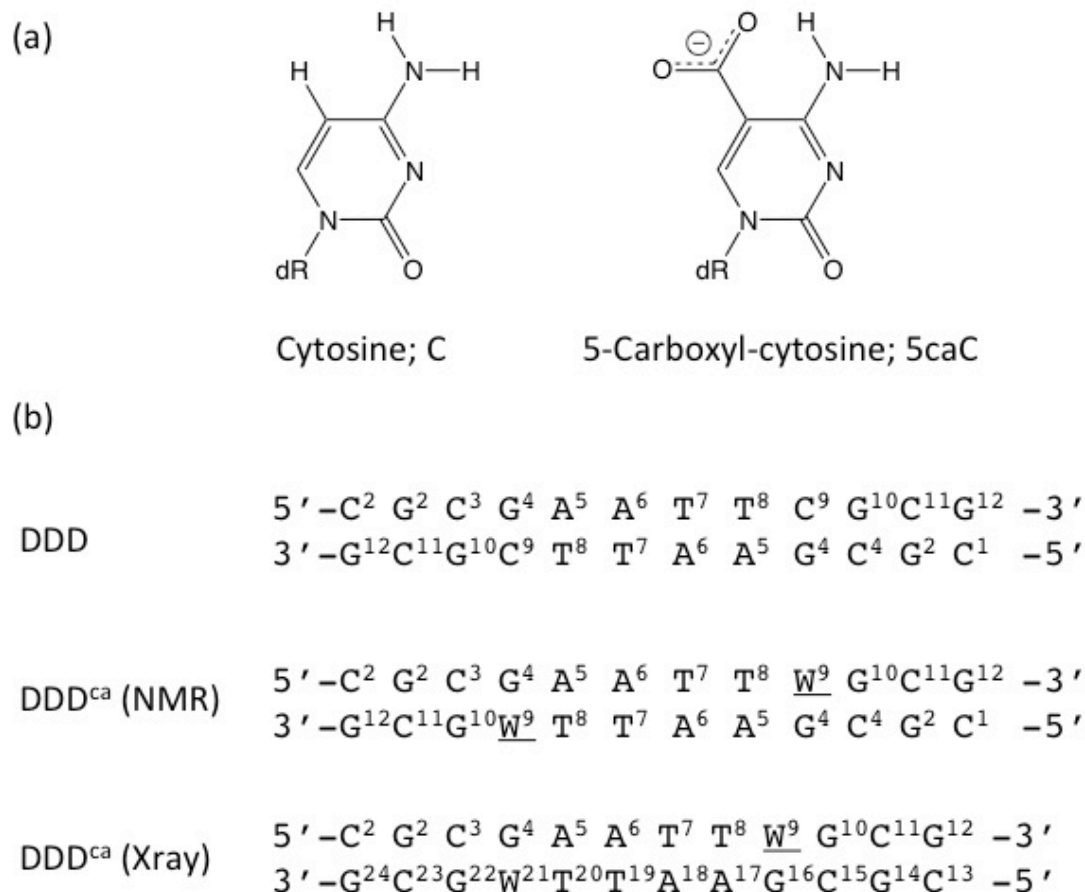


Figure 5.2. (a). Structure of dC, and 5caC. (b). Sequences and numbering of the nucleotides for unmodified DDD, and DDD^{ca} (NMR) and DDD^{ca} (X-ray) duplexes. In solution, the two strands of the DDD exhibit pseudo-dyad symmetry and the NMR resonances of symmetry-related nucleotides in the two strands are not individually observed. In the crystal, electron density for symmetry-related nucleotides in the two strands is observed and the nucleotides are numbered individually.

Thermal denaturation studies

At neutral pH, the melting temperature (T_M) of the modified duplex was 54 °C as measured by UV spectroscopy in 10mM phosphate buffer at pH 7.0, 100 mM NaCl,

50 μM Na_2EDTA . The corresponding T_M for the unmodified duplex under the same conditions was 48 $^\circ\text{C}$.

NMR melting studies

A series of 1D ^1H NMR spectra for the exchangeable protons were recorded in a range of 5 - 55 $^\circ\text{C}$ and are shown in Figure 5.3. Overall the data indicate that all imino protons are much stable in modified DDD^{ca} than in control sample. The N^l -imino proton of the $\text{G}^2\cdot\text{C}^{11}$ base pair in DDD was sharp and detectable up to 25 $^\circ\text{C}$, and at 35 $^\circ\text{C}$ it started to broaden. The same imino peak in the DDD^{ca} duplex started to broaden at much higher temperature (45 $^\circ\text{C}$). The next base pair, $\text{G}^{10}\cdot\text{C}^3$, which is the 5'- neighbor of the modification, was sharp and quite intense in DDD^{ca} , even at a temperature as high as 55 $^\circ\text{C}$, while already broadened at the same temperature in the control experiment. The N^l -imino proton of the $\text{W}^9\cdot\text{G}^4$ modified base pair in the DDD^{ca} is the most intense resonance among all, and remained sharp at 55 $^\circ\text{C}$. However the same resonance in DDD , even though it is the most stable, was much less intense and started to broaden. The N^3 -imino resonances of T^7 and T^8 are the sharpest in both duplexes; they start to broaden at 55 $^\circ\text{C}$ in the DDD and above that temperature in the DDD^{ca} , indicating that these thymine base pairs are much more stable than the same base pairs in the control sequence.

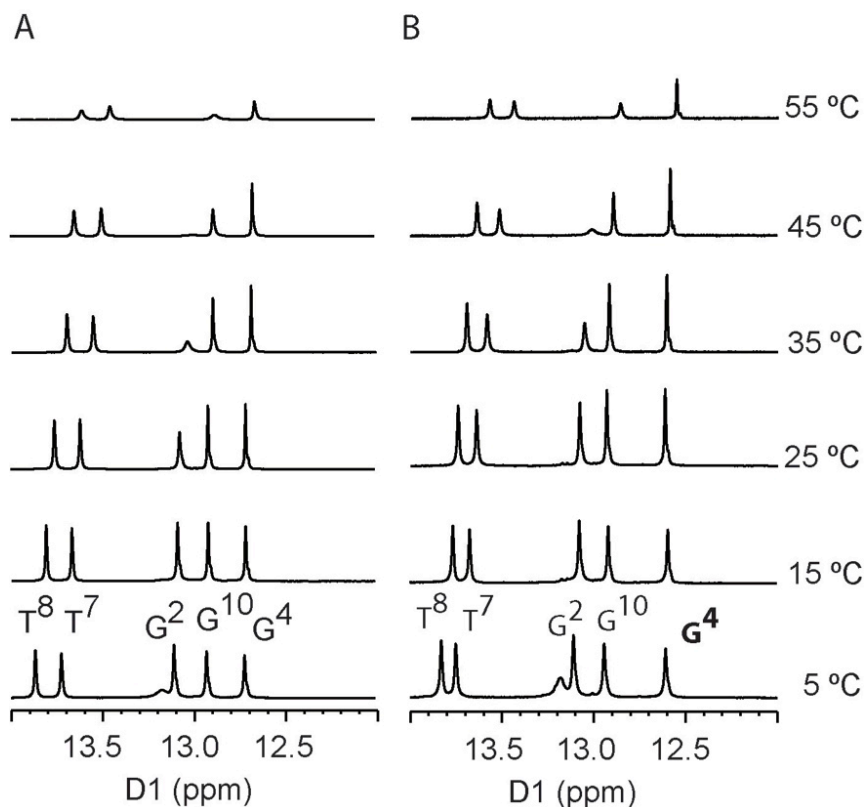


Figure 5.3. ¹H-NMR of imino proton resonances as a function of temperature for the unmodified DDD duplex (A), and the modified DDD^{ca} duplex (B).

Exchangeable protons.

Figure 5.4 shows the NOE connectivity of the purine N^1 and pyrimidine N^3 imino protons. The base imino protons were assigned based on their sequential connectivities in NOESY spectra, and these assignments were supported by their NOE cross-peaks to Watson-Crick base-paired amino protons. The sequential connectivities were obtained from base pairs $G^2:C^{11} \rightarrow G^{10}:C^3 \rightarrow G^4:C^9 \rightarrow T^8:A^5 \rightarrow T^7:A^6$. For the DDD and DDD^{ca} duplexes, the imino-proton resonances of the terminal base pairs $C^1:G^{12}$ are lost by fast exchange with water.

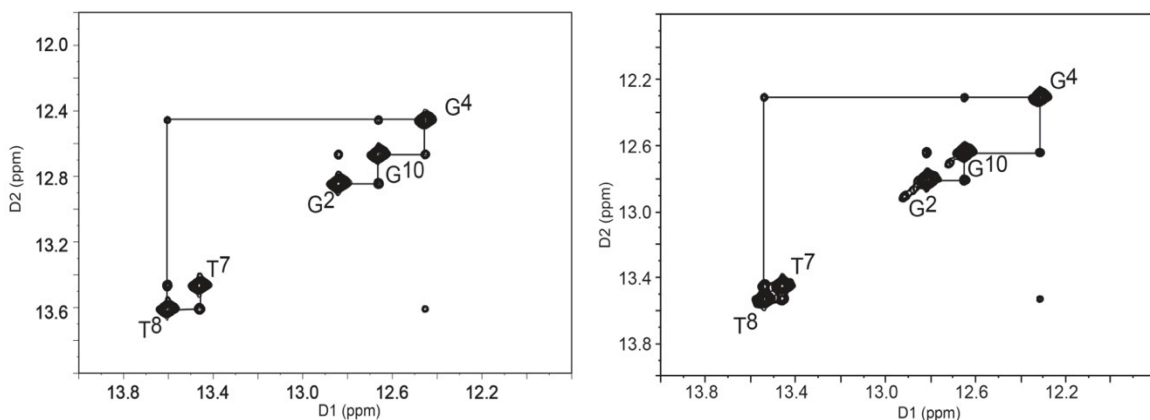


Figure 5.4. ^1H - ^1H NMR NOESY spectrum showing resonances for the thymine and guanine imino protons and sequential NOE connectivity for the imino protons of the base pairs $\text{G}^2:\text{C}^{11}$ to $\text{A}^6:\text{T}^7$ for unmodified DDD (left), and modified DDD^{ca} (right) duplexes.

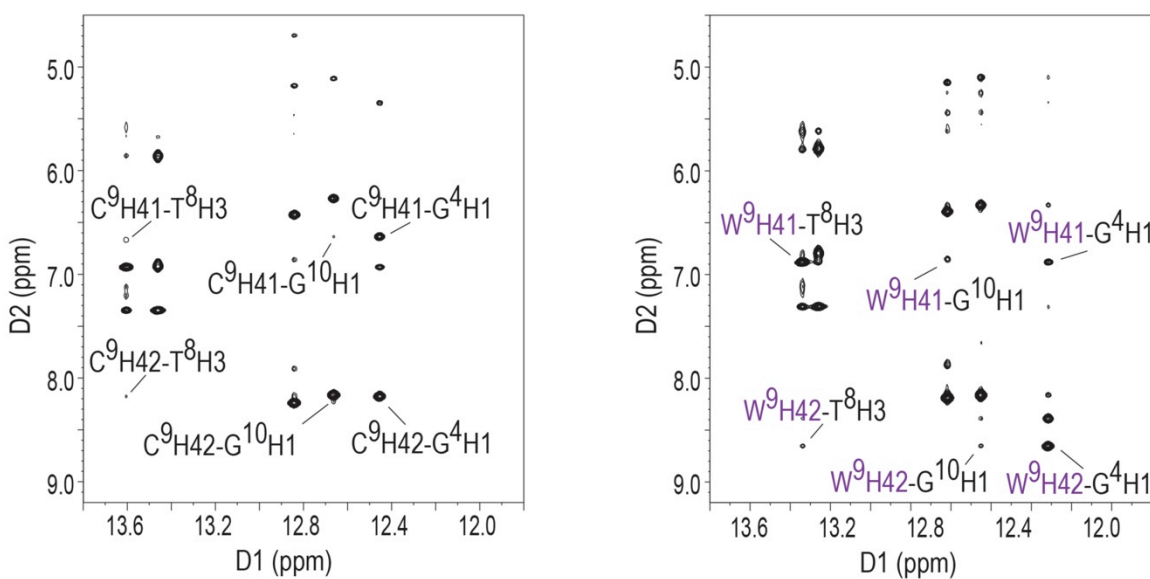


Figure 5.5. Expansion of the ^1H - ^1H NOESY spectra for DDD (left), and DDD^{ca} (right) showing the conservation of Watson-Crick base pairing and base stacking at the modification site.

The upfield region of the NOESY spectrum, shown in Figure 5.5, showed the NOEs between the imino and amino protons. Cross peaks from modified base C⁹H41, C⁹H42 to the opposite G⁴H1 base were observed, as well as interactions to neighbor T⁸H3 and G¹⁰H1 bases. No significant changes in intensities were observed in the DDD^{ca} duplex, compared with DDD.

Nonexchangeable protons

The sequential assignment of nonexchangeable protons was accomplished using standard protocols. The unmodified duplex was used as a control for NMR assignments for DDD^{ca}. For modified duplexes, the anticipated pattern of sequential base aromatic → deoxyribose anomeric nuclear Overhauser enhancement (NOE) was identified from C¹ → G¹² (Figure 5.6).

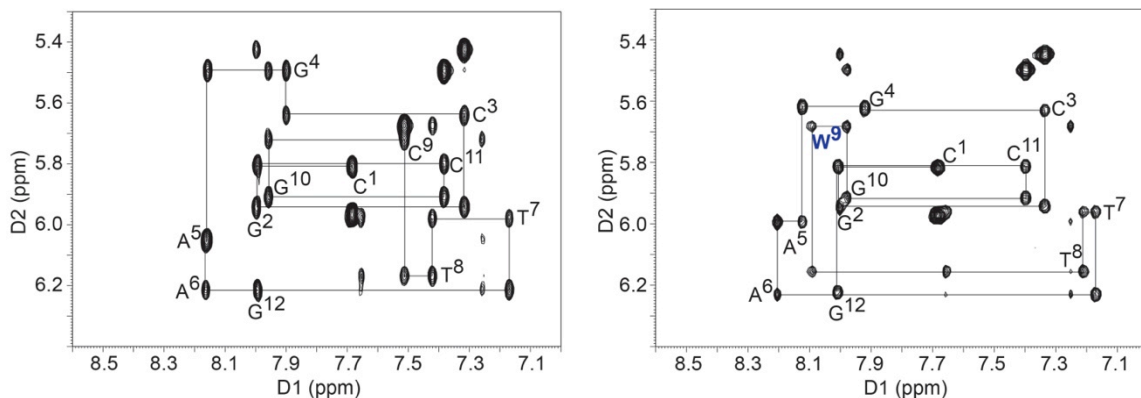


Figure 5.6. Expanded plot from the aromatic-anomeric region of the NOESY spectrum, showing sequential NOE connectivities in the unmodified DDD, and modified DDD^{ca} duplexes.

Only one set of resonances was observed because the sequences are self-complementary. The spectra exhibited well resolved cross-peaks. Three strong NOEs accounted for the H5-H6 cross-peaks of the cytosine residues (C¹, C³, and C¹¹). The H5-H6 resonance from W⁹ is missing due to carboxyl substitution at the 5-position in the DDD^{ca}. Cytosine assignments were confirmed by a DQF-COSY spectrum recorded under identical conditions (data not shown). Each base proton exhibited NOE peaks to its own and 5'-flanking H1' deoxyribose protons. For T⁸, W⁹ and G¹⁰ the NOE cross-peaks intensities between the base protons and the sugar H1' of the attached deoxyribose moieties were of the same relative magnitudes as those between other bases in the sequence. The 5caC H6 resonance was observed at 8.1 ppm, which is shifted downfield by approximately 0.6 ppm with respect to the unmodified oligodeoxynucleotide. This was attributed to the differential electronic density for 5caC as compared to G. Proton resonances from the opposite G⁴ and A⁵ bases exhibited chemical shift changes of <0.1 ppm compared with those bases in the unmodified DDD duplex.

Crystal structure of the DDD modified with 5caC

Crystals belonged to the orthorhombic $P2_12_12_1$ space group with unit cell parameters $a = 24.25 \text{ \AA}$, $b = 41.34 \text{ \AA}$, $c = 66.41 \text{ \AA}$. The crystal structure of DDD^{ca} was refined isotropically to a resolution of 1.95 \AA . The rmsd values for bond lengths were 0.009 \AA , and for angles they were 2.209 deg. The relatively high resolution achieved for the structure allowed for a detailed analysis of the geometric

and conformational changes in the duplex as a consequence of the introduced modification.

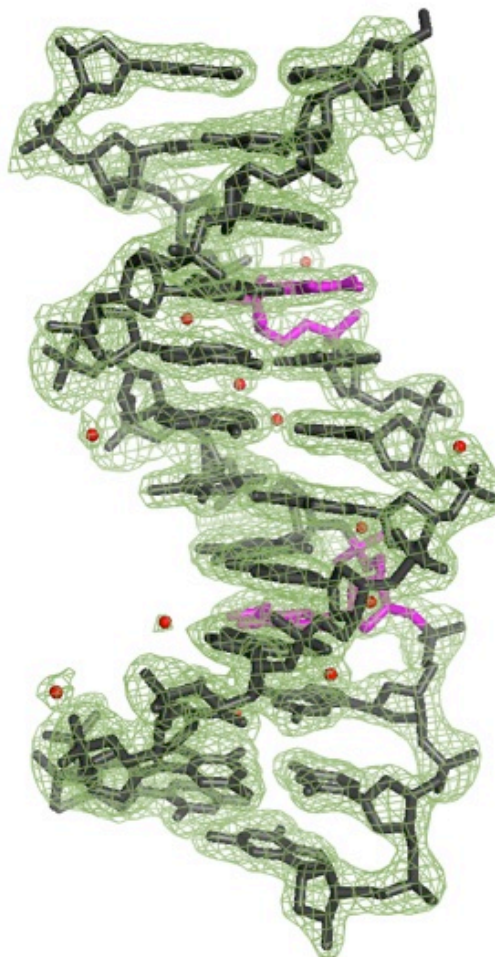


Figure 5.7. Fourier (2Fo-Fc) sum electron density contoured at the 1.0σ level (green meshwork) surrounding the DDD^{ca} duplex. The 5caC modified base is shown in magenta, and water molecules are shown in red.

Minimal perturbations of the DNA duplex were observed at the modification site (Figure 5.7). The carboxyl group was in the major groove (Figure 5.8), the oxygen from the COO^- group was within hydrogen bonding distance of the amino group at the C4 position of the modified cytosine.

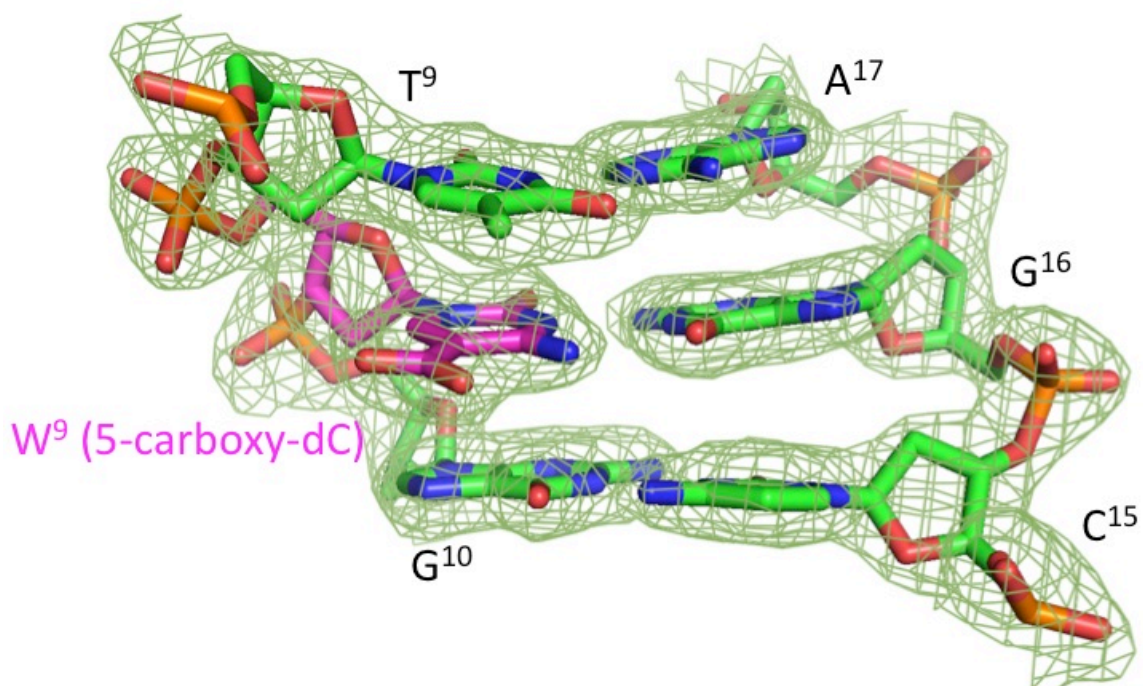


Figure 5.8. Fourier (2Fo-Fc) sum electron density contoured at the 1.0σ level (green meshwork) surrounding the DDD^{ca} duplex at one of the modification sites.

The DDD^{ca} structure is very similar to the other DDD structures. It overlays well with the canonical structure of DDD (PDB code: 436D) with an rmsd of 0.44 Å. The base stacking pattern in the modified DDD^{ca} is very similar to the one observed in unmodified B-type DNA (Figure 4.9), and helical parameters obtained with the program Curves+ (100) are comparable (Figures 4.11 – 4.13). The substitution of the cytidine nucleobase at the C5 position with the carboxyl group did not disrupt Watson-Crick base pairing or change the conformations of these residues (Figure 5.9). Analysis of the waters forming the “spine” of hydration, shows that the waters involved in the minor groove

hydration are conserved. Detailed information about data collection and refinement statistics are shown in Table 2.2.

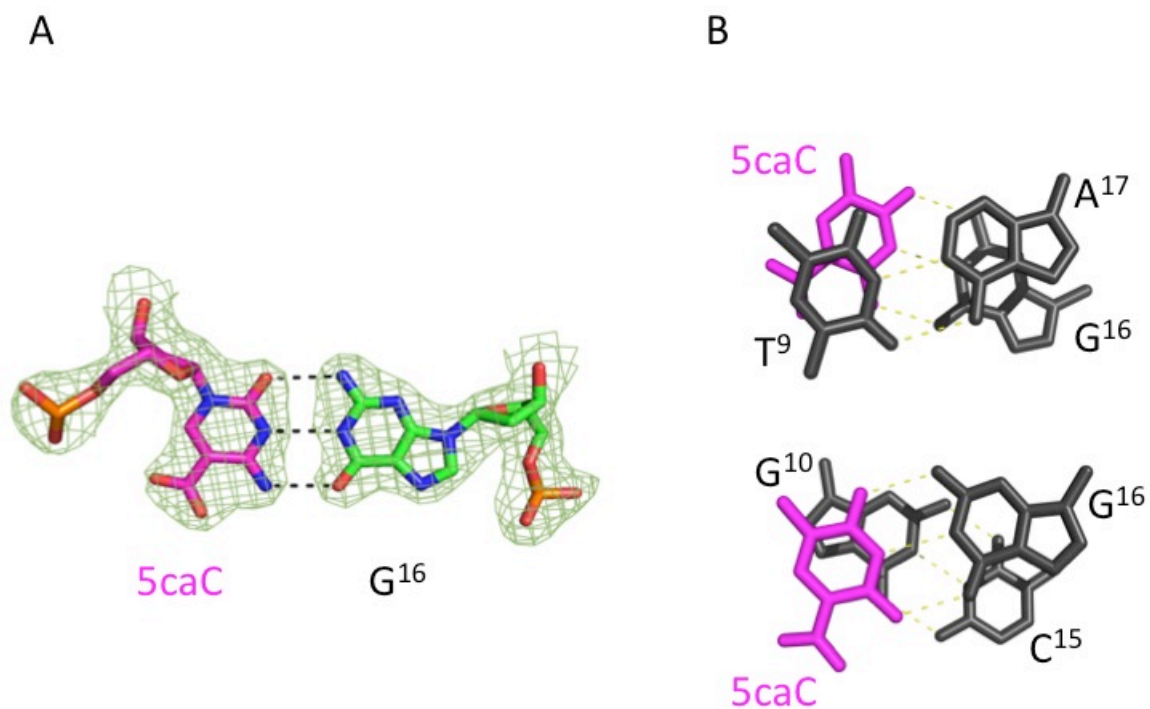


Figure 5.9. (A) Fourier (2Fo-Fc) sum electron density contoured at the 1.0σ level (green meshwork) around the 5caC:dG modified base pair showing conserved Watson-Crick base pairing geometry. (B) Stacking interactions in the DDD^{ca} duplex: (top) stacking of base pair T⁸:A¹⁷ above base pair W⁹:G¹⁶, and (bottom) stacking of base pair W⁹:G¹⁶ above base pair G¹⁰:C¹⁵.

Dynamics of base pair opening

To determine the dynamics of the base pairs, imino exchange rates were measured in the presence of base catalyst. In this work, ammonia base as the proton acceptor in imino proton exchange was used, due to its small size, lack of charge and high accessibility factor. Figure 5.10 shows imino proton exchange rates obtained by measuring magnetization transfer from water as a function of added ammonia base

catalyst. The results of the imino proton exchange analysis are summarized in Table 5.1. For the terminal (i.e. C¹·G¹²) and penultimate (G²·C¹¹) base pairs the exchange rates could not be measured because of the increased exchange with the solvent protons.

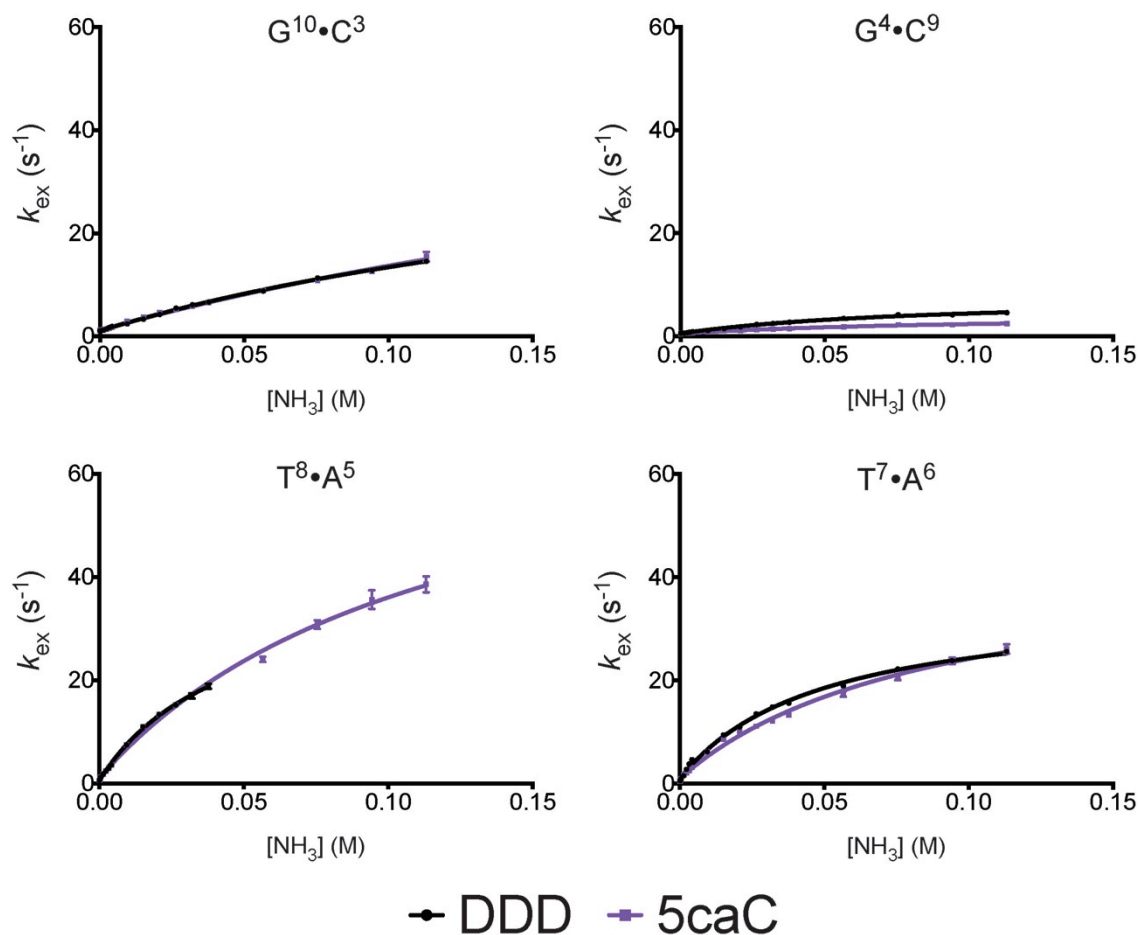


Figure 5.10. Plots showing imino proton exchange rates obtained by monitoring magnetization from water as a function of ammonia base catalyst. A. G¹⁰:C³. B. G⁴:C⁹. C. T⁸:A⁵. D. T⁷:A⁶ in DDD (black), and DDD^{ca} (purple).

For all studied imino protons, in both duplexes, the exchange rate reaches the EX1 regime, as predicted by eq. 8. As the concentration of base ammonia catalyst increases, the values of the exchange rate increase until a plateau is reached, indicating a change in the rate limiting step from chemical exchange to base pair opening ($k_{\text{ex}} = k_{\text{op}}$

and exchange occurring at each opening event). For the imino proton of G⁴, the exchange rate has a weak dependence on the concentration of ammonia base. This finding reflects the small equilibrium constant for opening of the G⁴·C⁹ base pair; in DDD^{ca} it is ~2 times smaller than in the DDD ($\alpha K_{op} = 0.6 \times 10^7$ vs. 1.2×10^7 for G¹⁰:C³ in DDD^{ca} and DDD, respectively). The same trend is observed with the opening rate, whereby G⁴·W⁹ base pair opens with a rate of 4 s⁻¹, while unmodified G⁴·C⁹ base pair has a rate of 8 s⁻¹. With almost the same an opening rate that is as the closing rate, the G⁴·W⁹ modified base pair is the least dynamic base pair in the analyzed duplexes.

	k_0 (s ⁻¹) ^a		$K_{op} \times 10^7$	
	DDD	DDD ^{ca}	DDD	DDD ^{ca}
C ³ :G ¹⁰	1.1 ± 0.06	1.1 ± 0.06	2.9 ± 0.1	3.2 ± 0.08
G ⁴ :C ⁹	0.57 ± 0.03	0.49 ± 0.04	1.2 ± 0.04	0.6 ± 0.04
A ⁵ :T ⁸	0.59 ± 0.03	0.59 ± 0.03	41 ± 0.08	34 ± 1
A ⁶ :T ⁷	0.61 ± 0.03	0.55 ± 0.03	37 ± 5	34 ± 0.9

	k_{op} (s ⁻¹)		k_{cl} (× 10 ⁻⁷ s ⁻¹)	
	DDD	DDD ^{ca}	DDD	DDD ^{ca}
C ³ :G ¹⁰	45 ± 3	64 ± 12	15 ± 1	20 ± 1
G ⁴ :C ⁹	8 ± 0.5	4 ± 0.3	6.7 ± 0.2	6.6 ± 0.1
A ⁵ :T ⁸	40 ± 2	78 ± 5	0.97 ± 0.04	2.3 ± 0.05
A ⁶ :T ⁷	36 ± 1	45 ± 3	0.98 ± 0.09	1.3 ± 0.01

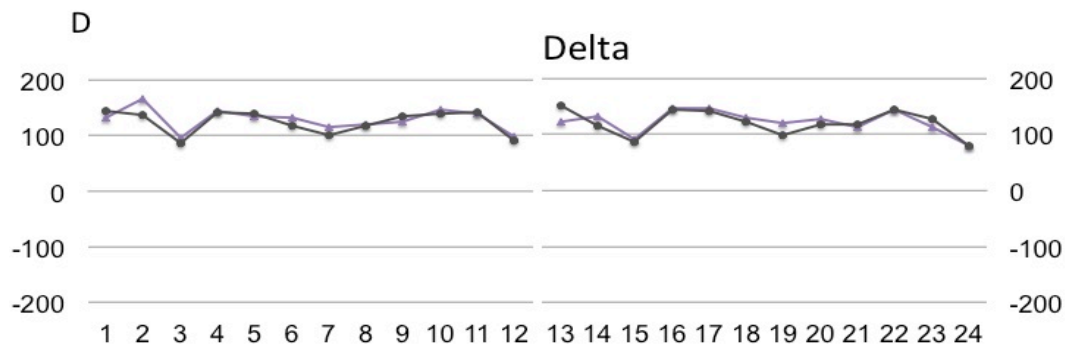
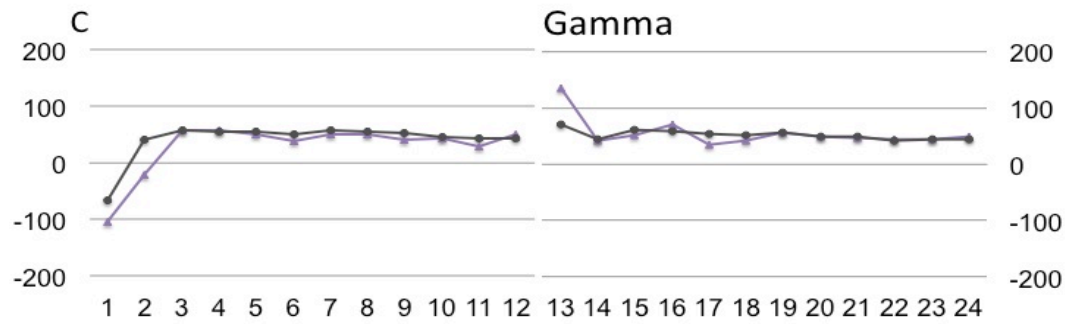
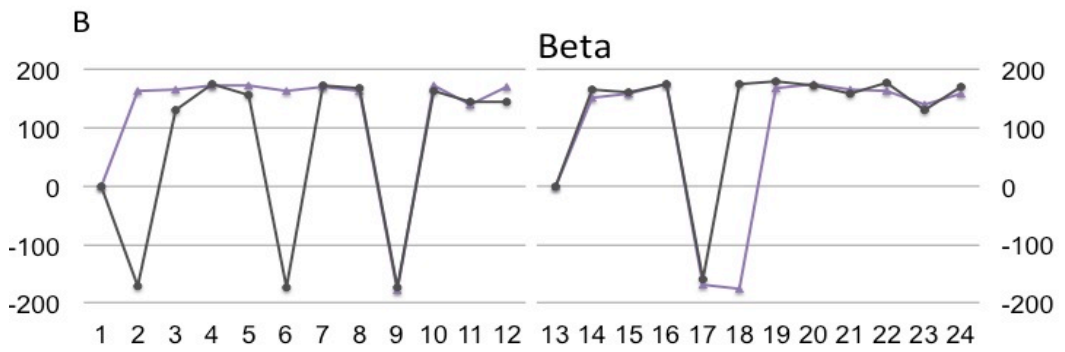
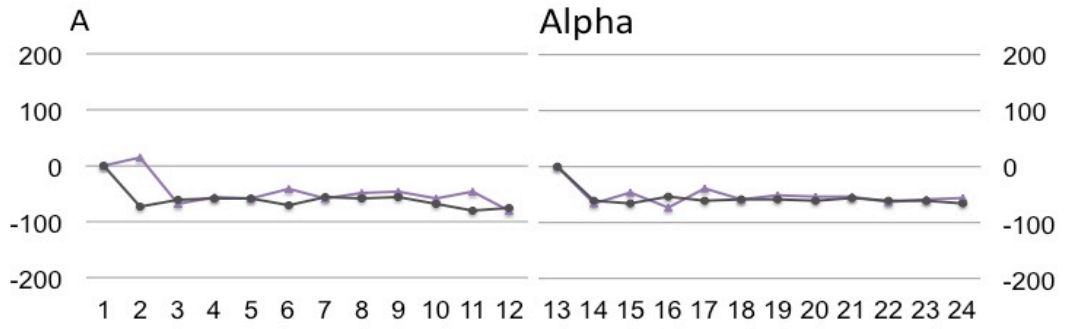
Table 5.1. Rate and Equilibrium Constants for DNA Base Pair Opening. ^aThe observed exchange rate without an ammonia catalyst.

For G¹⁰·C³ imino protons, the exchange rates and all kinetic parameters are at the same or similarly low level for both the DDD^{ca} and the DDD duplex.

The highest value for the equilibrium constant of base pair opening is for the T⁸ imino proton, which is the 5' neighbor of the modification site. This applies to all three duplexes. The dynamics behavior of this base pair is remarkably different in the studied duplexes. The dynamics behavior of this base pair is remarkably different in the studied duplexes. With an opening rates that are almost doubled in DDD^{ca}, and doubled closing rate (Table 5.1), it appears that the frequency for base pair opening and closing are the same.

Helical analysis

An analysis of the helical parameters of the DDD^{ca} was performed with the program Curves+ (100). The analysis of the backbone torsion angles (Figure 5.11) showed no significant changes in alpha, gamma, delta, and zeta angles. For the beta torsion angle, the DDD^{ca} shows similar conformations as the unmodified DDD (77). However, peaks at the A⁶ and A¹⁷ reflect changes induced by the interacting Mg²⁺ ion observed in the unmodified DDD structure. Similarly the peak at T²⁰ residue for DDD^f relative to DDD for torsion angle epsilon reflects the change as the result of the presence of the modification at the flanking X²¹ residue. The analysis of interbase parameters shows no substantial changes in helical rise, roll and twist, as shown in Figure 5.12. Moreover shear and stretch, that characterize intrabase translations are comparable between bases in all studied duplexes (Figure 5.13).



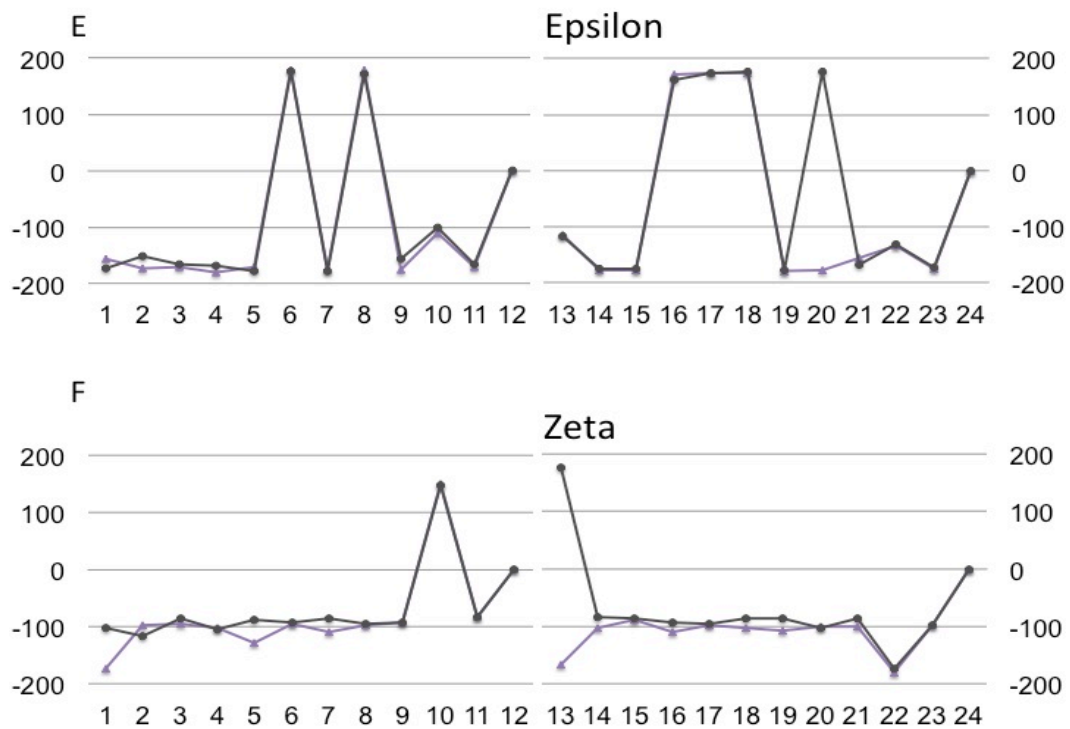


Figure 5.11. Comparison of backbone torsion angles (a) alpha, (b) beta, (c) gamma, (d) delta, (e) epsilon, and (f) zeta in the structure of the DDD^{ca} (in purple), and in the unmodified DDD (PDB entry 436D (124)) (in black).

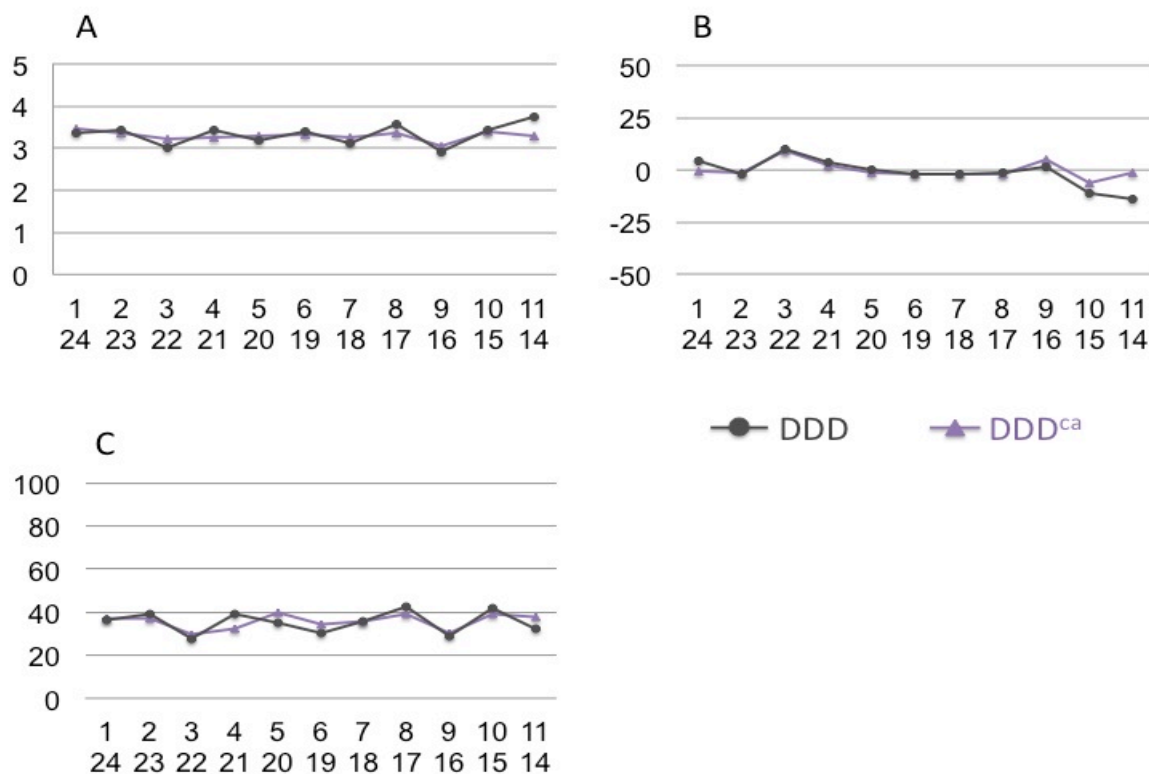


Figure 5.12. Interbase pair parameters: (a) helical rise, (b) roll, and (c) twist for the structure of DDD^{ca} in purple, and unmodified DDD (PDB entry 436D (124)) in black.

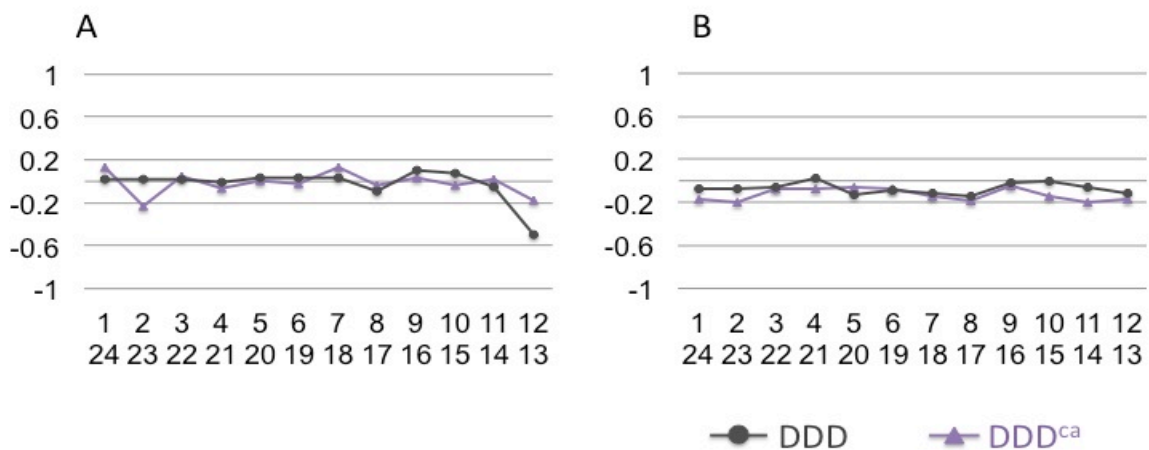


Figure 5.13. Intrabase pair parameters: (a) shear, (b) stretch in the structure of the DDD^{ca} (in purple), and the unmodified DDD (PDB entry 436D (124)) (in black).

Discussion

The 5caC base is an active demethylation oxidation product derived from 5mC, formed post-replicatively by TET enzymes (39). The 5caC base has been detected in genomic DNA of mouse embryonic stem (ES) cells and was found to be at level of 3 5caC in every 10^6 C (39).

Effect of 5caC on DNA stability.

The T_m of the DDD^{ca} duplex is 6 °C higher than the T_m of the unmodified duplex, demonstrating a stabilizing effect of carboxylated DNA. The same effect is observed in temperature-dependent NMR spectroscopy (Figure 5.3). Overall, the NMR data show that all imino protons appear as sharp peaks at a temperature as high as 55 °C. Only the penultimate base pair in DDD^{ca} is broadened at 55 °C, but resonances coming from other base pairs are still observed. The G⁴ imino resonance, representing the modified base pair G⁴:W⁹ appears as the sharpest and most intense peak in the spectrum. The imino protons of the unmodified sequence are broadening at lower temperature. Previous studies on DNA electrostatics and stability showed that the presence of positive charge on the DNA stabilizes the duplex (153). It would appear by analogy that additional negative charge in DNA would result in destabilization. In fact, Maiti and Drohat (141) showed that 5caC ionizes with pK_a values of 4.28 at the N3 position, and 2.45 at the carboxyl, which suggests that 5caC exists as a monoanion at physiological pH. Substitution with 5caC introduces a negatively charged carboxyl group, which affects the electrostatics of the nucleobase. Double bond character on the oxygen from the carboxyl group serves as a proton acceptor and is capable to form a hydrogen bond. The amino group at C4 is in close proximity and donates a hydrogen. This additional hydrogen bond is formed within the 5caC:G base pair, which enlarges the modified

cytosine by an additional ring. In this fashion it mimics the two-ring purine nucleic base. This results in increased stacking and increased DNA stability (Figure 5.9). Base stacking interactions are of importance in stabilizing nucleic acid duplex.

Imino Proton Exchange and Base Pair Opening Kinetics.

DNA glycosylase repair enzymes are believed to employ an extrahelical recognition mechanism to bind and excise damaged bases from DNA (147, 154-158). Stivers and co-workers (54, 148, 159) showed that DNA stability and dynamics play a crucial role in detection of damaged base by repair enzyme. The assumption is that base pairs with lower stabilities will have greater opening rates. For the G⁴ imino proton, which represents the modified base pair, our data show that the exchange rate has a weak dependence on the concentration of the ammonia base, resulting in a small equilibrium constant for opening the G⁴:W⁹ base pair. In the DDD^{ca}, αK_{op} is small and $\alpha K_{op} = 0.6 \times 10^7$ for the modified base pair, while in control DDD $\alpha K_{op} = 1.2 \times 10^7$. The same trend is observed for the opening rate, whereby G⁴:W⁹ base pair opens 4 times per second, while the unmodified G⁴:C⁹ base pair opens with a rate of 8 s⁻¹. With similar values for opening and closing rates, the G⁴:W⁹ modified base pair represents the least dynamic base pair among all pairs in the studied duplexes. This result implies that the possibility of 5caC adopting an extrahelical conformation for excision by thymine DNA glycosylase is unlikely.

Effect of 5caC on DNA Structure and Structure-Activity Relationships.

In order to understand the mechanism of active demethylation through 5caC formation, chemical reactivity studies on the modified nucleobases were reported (140). The sensitivity of these modifications toward oxidation and deamination was measured,

along with C-C bond cleaving reactivity, in the presence and absence of thiols as catalysts. 5caC can undergo thiol-catalyzed and acid-catalyzed C-C bond cleavage reactions to form dC with the release of CO₂. Thus, decarboxylation could occur as an alternative active demethylation pathway (151). It was proposed that in the process of active DNA demethylation, in the last step, 5caC could decarboxylate to dC (160). In that case it is believed that DNA decarboxylase might share some structural, sequential and mechanistic similarities with isoorotate decarboxylase (IDCase), which catalyzes decarboxylation of 5caU to U in fungi (160). Indeed, Xu *et al.* (160) reported biochemical studies with fungi IDCase catalyzing decarboxylation of 5caC to dC, albeit with weak activity. Nonetheless this was the first *in vitro* evidence for direct decarboxylation of 5caC to dC by the enzyme. However, with respect to DNA demethylation in mammals, no enzyme that is capable of decarboxylation of 5caC-containing DNA has been identified. TET enzymes act together with other DNA demethylation pathways such as BER. In this process 5caC is removed by thymine DNA glycosylase (TDG) resulting in the restoration of unmodified cytosine (43, 78, 161, 162). TDG can excise 5caC from DNA *in vitro*, and this activity was confirmed in mammalian cells (136-139).

It was proposed by Hashimoto *et al.* (142, 143) that 5caC undergoes tautomerization caused by the presence of the negatively charged carboxyl group at C5 on modified cytosine, and forms a Wobble base pair with the opposite guanine, similar to the structure of G:U or G:T mismatches (Figure 5.1) (142, 143). This mismatch base pair geometry is recognized by TDG, and results in the excision of the mispaired base and repair. In order to determine the basis by which 5caC pairs with the opposite guanine in DNA, the structure of 5caC placed opposite dG (DDD^{ca}) was determined. The crystals of DDD^{ca} diffracted at a resolution of 1.95 Å. The structure revealed that 5caC has a minimal effect on duplex conformation (Figures 5.7 and 5.8), and base pair geometry (Figure 5.9), as compared to canonical dC:dG base pair. The Watson-Crick

geometry was conserved for the 5caC:G base pair, indicating the amino tautomer for the modified base (Figure 5.9). In addition, the solution NMR data corroborate the crystallographic data as can be seen from a comparison of NOESY spectra of the modified and unmodified duplexes. The sequential pattern of imino-imino and imino-amino NOEs is conserved in the presence of the modified base pair, confirming that the base pairing and base stacking is maintained at the lesion sites (Figures 5.4 and 5.5). In addition, the pattern of sequential base aromatic→deoxyribose anomeric NOEs is also conserved (Figure 5.6), with no significant changes in chemical shifts, which indicates the presence of a normal undisturbed B-type helix. It was confirmed at the nucleobase level as well, by Maiti and Drohat (78, 141), who showed higher relative stabilities of the amino tautomers for 5caC. These authors also reported an investigation of the TDG mechanism excising 5caC from DNA, and the chemical properties indicating the catalytic requirements for the excision. They showed that TDG activity is greater for cytosine analogs with an electron-withdrawing substituent at the C5 position, which can stabilize negative charge developing on the excised base in the transition state. In the case of 5caC, σ_m does not predict the electronic effect for the carboxyl group well. At physiological pH, the carboxyl group is in the anionic form, and $\sigma_m = -0.10$, which indicated an electron-donating effect. However, Drohat *et al.* (78, 141) calculated that the anionic carboxyl substituent lowers the pK_a of cytosine by -0.3. This suggests a stabilizing effect of COO^- by NH_2 group at position C4, which would indicate an electron withdrawing effect. It implies that TDG could remove 5caC from DNA (78, 141).

Summary

The 5caC base is stably stacked in the DNA duplex, and the melting temperature of the DDD^{ca} is higher than that of the canonical duplex. The dynamics at the modification site and the opening rates are low, which suggests a low probability of 5caC being flipped out of the duplex and adopting an extrahelical conformation to be recognized by thymine DNA glycosylase is low.

Acknowledgements

This work is based upon research conducted at the Advanced Photon Source on the Northeastern Collaborative Access Team beamlines, which are supported by a grant from the National Institute of General Medical Sciences (P41 GM103403) from the National Institutes of Health. Use of the Advanced Photon Source, an Office of Science User Facility operated for the U.S. Department of Energy (DOE) Office of Science by Argonne National Laboratory, was supported by the U.S. DOE under Contract No. DE-AC02-06CH11357.

This work was supported by NIH grant R01 CA-55678 (M.P.S.). Funding for NMR was supplied by NIH grants S10 RR-05805, S10 RR-025677 and NSF Grant DBI 0922862, the latter funded by the American Recovery and Reinvestment Act of 2009 (Public Law 111-5). Vanderbilt University assisted with the purchase of NMR instrumentation.

CHAPTER 6

SUMMARY, CONCLUSION AND FUTURE DIRECTIONS

This dissertation describes studies toward the characterization of oxidation derivatives of 5-methylcytosine (5mC): 5-hydroxymethylcytosine (5hmC), 5-formylcytosine (5fC), and 5-carboxylcytosine (5caC), which were site-specifically incorporated into the Dickerson-Drew Dodecamer (DDD) DNA sequence. The structures of modified DNA in the crystalline state were determined by X-ray crystallography, and the characterization of modified DNA in solution was performed using high-resolution NMR spectroscopy. In order to characterize the kinetics of base-pair opening in the native and modified nucleic acid duplexes, high-resolution proton exchange NMR spectroscopy was used. The rates of exchange of imino protons with solvent protons were measured by magnetization transfer from water for each DNA duplex as a function of the concentration of exchange base catalyst.

The methylation of cytosine in DNA is very important in epigenetics, as cells use methylation cells to switch genes on and off on their way to specialization (23-25, 163). While it is clear how the methylation occurs, the question of demethylation pathways is of high interest and many different studies have addressed this issue recently. The search for active demethylation pathways led to the discovery of 5hmC in the mammalian genome. As it turns out 5hmC is oxidized by TET enzymes from 5mC (33, 34, 163). Since 5hmC was found to be tissue specific and high levels of 5hmC were found in the central nervous system, it was proposed that 5hmC is an epigenetic marker

similar to 5mC (33, 34, 163). However, at approximately at the same time it was found that TET enzymes can further oxidize 5hmC to 5fC and 5caC, and the latter could be decarboxylated to unmodified cytosine. Therefore it was proposed that active demethylation of 5mC proceeds through oxidation to 5hmC, 5fC, and 5caC by TET enzymes (39). However, TET enzymes act together with other DNA demethylation pathways such as base excision repair (BER). Maiti and Drohat showed that TDG might be involved in demethylation when 5fC and 5caC, but not 5hmC, are removed via BER (115). While thymine DNA glycosylase (TDG) removes 5fC and 5caC, it is unable to remove 5hmC (115).

The initial and most important step in BER involves finding the damaged base by the repair enzyme. There are many different hypotheses about the recognition of the lesion by the enzyme, and some of them were addressed in this dissertation. In one of the hypothesis, the structural characteristics recognized by TDG involve Wobble base pairing geometry at the modification site (57, 141-143, 146, 164). 5hmC was found not to be a substrate for TDG and BER, but rather an epigenetic marker (78, 165). High resolution NMR spectroscopy and the X-ray crystal structure of the DDD substituted with 5hmC at 1 Å resolution showed no perturbations in the DNA duplex conformation as the structure was of normal B-type geometry. At the modification site the 5hmC base was stably stacked and paired with opposite G according to Watson-Crick geometry. The hydroxymethyl group of the modified base was oriented toward the 3'- end of the duplex and interacted via two structural water molecules with O6 and N7 on flanking 3'- G¹⁰ base. That suggested a stabilizing effect as a result of the oxidized modified base. The further oxidation products 5fC and 5caC were proposed to undergo tautomerization,

which would lead to protonation at N3 of the modified cytosine and affect pairing with guanine (142, 143). Thus, in order to avoid two potentially repulsive hydrogens in the base pair, both nucleobases would have to relocate into a different conformation to form, for instance, Wobble-like base pair geometry, as it was observed in case of G/T or G/U mismatches in DNA, which were shown to be recognized and excised by TDG (57). However there is no evidence for dual or any different conformations for modified 5fC:G and 5caC:G base pair in the studied DDD^f and DDD^{ca} structures. Both modifications form normal Watson-Crick geometry in the crystalline state, and there was no evidence for imino tautomers present in solution as shown by NMR high-resolution spectroscopy.

It was hypothesized that DNA glycosylases use an extrahelical mechanism to recognize the modified base possessing higher opening rates and lower stability. When 5hmC analog was introduced to DDD sequence, no change in the thermal stability was observed relative to unmodified DDD. The dynamics at the modification site and the opening rates are low, which suggests that the probability of 5hmC to flip out of the DNA duplex and adopting an extrahelical conformation to be recognized by repair glycosylase is low. These findings are consistent with current studies done by other labs in that 5hmC is not a substrate for TDG glycosylase (78). The presence of the 5fC in a DNA duplex decreases its stability and the kinetics are remarkably different from the control sequence. The opening rate for the modified base pair is 3-times higher than that for the canonical sample. In addition, the dynamics for base pair opening for the flanking 5'-neighboring base pair is also five times higher. The high frequency for opening of bases at the modification site indicates a highly dynamic “behavior” that could be captured by the repair DNA glycosylase and attract the enzyme

to the damaged base. In the case of 5caC, the melting temperature is higher relative to the canonical duplex. The dynamics at the modification site and the opening rates are low, suggesting a low probability of 5caC to flip out the duplex and adopt an extrahelical conformation that can be recognized by thymine DNA glycosylase. Since the 5caC modification was found to be excised from DNA by TDG it raises the question about the recognition of the lesion by the enzyme. There was no structural indication of unusual base pairing geometry in the crystal structure. Moreover, the low dynamic properties and high stability of the base pair does not support this hypothesis. That means that TDG may use a different recognition mechanism to process the damaged base.

The work on many different glycosylases demonstrated that there is not one universal mechanism used by all the enzymes to locate the lesion (*147, 166-169*). In the studies done by Plum and Breslauer the relationship between DNA lesions and DNA repair involving a “thermodynamic signature” was proposed (*170*). This led to the hypothesis that repair enzymes can sense the damaged nucleotide based on its local thermodynamic effect, even when the structures of the DNA with and without the lesion are indistinguishable by structural analysis using either crystallography or NMR. Indeed, this scenario was tested in studies with 5-hydroxy-2'-deoxycytidine (5ohC) embedded in the DDD sequence (*171*). This oxidative lesion is capable of Watson-Crick pairing with guanine, although it is highly destabilizing when compared to unmodified DDD, due to the reduced enthalpic term as a result of disrupted base stacking. Temperature-dependent NMR studies revealed local destabilization of the modified base pair, whereas it is the most stable base pair in native sequence. The crystal structure with the 5ohC

modification is indistinguishable from the canonical sequence (172). Thus it was proposed, that DNA glycosylase uses the thermodynamic signature mechanism to find the lesion based on local thermodynamic destabilization (171). The detailed analysis of the 5hmC modification in the DDD duplex revealed no thermodynamic changes compared to the native DDD. 5hmC is a stable base in DNA and with no enzyme discovered yet that can excise 5hmC from the genome, it is possible that it serves as an epigenetic marker. For the other oxidation products such as 5fC and 5caC, the complete thermodynamic analysis is still to be performed. In particular, as far as 5caC is concerned, additional information about the effects of this modification on the DNA stability could provide the specifics, which could be sensed by the repair enzyme.

APPENDIX
PDB COORDINATE FILES

File A-1: Crystal structure 5-hydroxymethyl-2'-deoxycytidine in B-type DNA, DDD^{hm}
(PDB code 4I9V).

```
HEADER      DNA                               05-DEC-12  4I9V
TITLE       THE ATOMIC STRUCTURE OF 5-HYDROXYMETHYL 2'-DEOXYCITIDINE BASE PAIRED
TITLE       2 WITH 2'-DEOXYGUANOSINE IN DICKERSON DREW DODECAMER
COMPND      MOL_ID: 1;
COMPND      2 MOLECULE: DNA (5'-D(*CP*GP*CP*GP*AP*AP*TP*TP*(5HC)P*GP*CP*G)-3');
COMPND      3 CHAIN: A, B;
COMPND      4 ENGINEERED: YES
SOURCE      MOL_ID: 1;
SOURCE      2 SYNTHETIC: YES;
SOURCE      3 ORGANISM_SCIENTIFIC: SYNTHETIC DNA;
SOURCE      4 ORGANISM_TAXID: 32630
KEYWDS      5-HYDROXYMETHYL 2' DEOXYCITIDINE, 5-HYDROXYMETHYL-DC ADDUCT, MODIFIED
KEYWDS      2 DDD, DODECAMER OF B-DNA, DNA
EXPDTA      X-RAY DIFFRACTION
AUTHOR      M.W.SZULIK, B.NOCEK, A.JOACHIMIAK, M.P.STONE
REVDAT      1 20-NOV-13 4I9V 0
JRNL        AUTH  M.W.SZULIK, B.NOCEK, A.JOACHIMIAK, M.P.STONE
JRNL        TITL  THE ATOMIC STRUCTURE OF 5-HYDROXYMETHYL 2'-DEOXYCITIDINE
JRNL        TITL  2 BASE PAIRED WITH 2'-DEOXYGUANOSINE IN DICKERSON DREW
JRNL        TITL  3 DODECAMER
JRNL        REF   TO BE PUBLISHED
JRNL        REFN
REMARK      2
REMARK      2 RESOLUTION.      1.02 ANGSTROMS.
REMARK      3
REMARK      3 REFINEMENT.
REMARK      3 PROGRAM          : REFMAC 5.7.0029
REMARK      3 AUTHORS           : MURSHUDOV, VAGIN, DODSON
REMARK      3
REMARK      3 REFINEMENT TARGET : MAXIMUM LIKELIHOOD
REMARK      3
REMARK      3 DATA USED IN REFINEMENT.
REMARK      3 RESOLUTION RANGE HIGH (ANGSTROMS) : 1.02
REMARK      3 RESOLUTION RANGE LOW  (ANGSTROMS) : 14.88
REMARK      3 DATA CUTOFF              (SIGMA(F)) : NULL
REMARK      3 COMPLETENESS FOR RANGE      (%)       : 95.0
REMARK      3 NUMBER OF REFLECTIONS          : 32113
REMARK      3
REMARK      3 FIT TO DATA USED IN REFINEMENT.
REMARK      3 CROSS-VALIDATION METHOD          : THROUGHOUT
REMARK      3 FREE R VALUE TEST SET SELECTION : RANDOM
REMARK      3 R VALUE          (WORKING + TEST SET) : 0.161
REMARK      3 R VALUE          (WORKING SET)       : 0.159
REMARK      3 FREE R VALUE                               : 0.182
```

REMARK 3 FREE R VALUE TEST SET SIZE (%) : 5.000
REMARK 3 FREE R VALUE TEST SET COUNT : 1700
REMARK 3
REMARK 3 FIT IN THE HIGHEST RESOLUTION BIN.
REMARK 3 TOTAL NUMBER OF BINS USED : 20
REMARK 3 BIN RESOLUTION RANGE HIGH (A) : 1.02
REMARK 3 BIN RESOLUTION RANGE LOW (A) : 1.05
REMARK 3 REFLECTION IN BIN (WORKING SET) : 1374
REMARK 3 BIN COMPLETENESS (WORKING+TEST) (%) : 56.96
REMARK 3 BIN R VALUE (WORKING SET) : 0.3350
REMARK 3 BIN FREE R VALUE SET COUNT : 82
REMARK 3 BIN FREE R VALUE : 0.3020
REMARK 3
REMARK 3 NUMBER OF NON-HYDROGEN ATOMS USED IN REFINEMENT.
REMARK 3 PROTEIN ATOMS : 0
REMARK 3 NUCLEIC ACID ATOMS : 490
REMARK 3 HETEROGEN ATOMS : 26
REMARK 3 SOLVENT ATOMS : 187
REMARK 3
REMARK 3 B VALUES.
REMARK 3 FROM WILSON PLOT (A**2) : NULL
REMARK 3 MEAN B VALUE (OVERALL, A**2) : 22.68
REMARK 3 OVERALL ANISOTROPIC B VALUE.
REMARK 3 B11 (A**2) : 0.24000
REMARK 3 B22 (A**2) : -1.49000
REMARK 3 B33 (A**2) : 1.26000
REMARK 3 B12 (A**2) : 0.00000
REMARK 3 B13 (A**2) : 0.00000
REMARK 3 B23 (A**2) : 0.00000
REMARK 3
REMARK 3 ESTIMATED OVERALL COORDINATE ERROR.
REMARK 3 ESU BASED ON R VALUE (A): 0.029
REMARK 3 ESU BASED ON FREE R VALUE (A): 0.030
REMARK 3 ESU BASED ON MAXIMUM LIKELIHOOD (A): 0.023
REMARK 3 ESU FOR B VALUES BASED ON MAXIMUM LIKELIHOOD (A**2): 0.990
REMARK 3
REMARK 3 CORRELATION COEFFICIENTS.
REMARK 3 CORRELATION COEFFICIENT FO-FC : 0.979
REMARK 3 CORRELATION COEFFICIENT FO-FC FREE : 0.974
REMARK 3
REMARK 3 RMS DEVIATIONS FROM IDEAL VALUES COUNT RMS WEIGHT
REMARK 3 BOND LENGTHS REFINED ATOMS (A): 629 ; 0.020 ; 0.012
REMARK 3 BOND LENGTHS OTHERS (A): 375 ; 0.006 ; 0.020
REMARK 3 BOND ANGLES REFINED ATOMS (DEGREES): 937 ; 2.488 ; 1.354
REMARK 3 BOND ANGLES OTHERS (DEGREES): 888 ; 2.562 ; 3.000
REMARK 3 TORSION ANGLES, PERIOD 1 (DEGREES): NULL ; NULL ; NULL
REMARK 3 TORSION ANGLES, PERIOD 2 (DEGREES): NULL ; NULL ; NULL
REMARK 3 TORSION ANGLES, PERIOD 3 (DEGREES): NULL ; NULL ; NULL
REMARK 3 TORSION ANGLES, PERIOD 4 (DEGREES): NULL ; NULL ; NULL
REMARK 3 CHIRAL-CENTER RESTRAINTS (A**3): 78 ; 0.406 ; 0.200
REMARK 3 GENERAL PLANES REFINED ATOMS (A): 337 ; 0.041 ; 0.020
REMARK 3 GENERAL PLANES OTHERS (A): 137 ; 0.008 ; 0.020
REMARK 3 NON-BONDED CONTACTS REFINED ATOMS (A): NULL ; NULL ; NULL
REMARK 3 NON-BONDED CONTACTS OTHERS (A): NULL ; NULL ; NULL
REMARK 3 NON-BONDED TORSION REFINED ATOMS (A): NULL ; NULL ; NULL
REMARK 3 NON-BONDED TORSION OTHERS (A): NULL ; NULL ; NULL

REMARK 3 H-BOND (X...Y) REFINED ATOMS (A): NULL ; NULL ; NULL
REMARK 3 H-BOND (X...Y) OTHERS (A): NULL ; NULL ; NULL
REMARK 3 POTENTIAL METAL-ION REFINED ATOMS (A): NULL ; NULL ; NULL
REMARK 3 POTENTIAL METAL-ION OTHERS (A): NULL ; NULL ; NULL
REMARK 3 SYMMETRY VDW REFINED ATOMS (A): NULL ; NULL ; NULL
REMARK 3 SYMMETRY VDW OTHERS (A): NULL ; NULL ; NULL
REMARK 3 SYMMETRY H-BOND REFINED ATOMS (A): NULL ; NULL ; NULL
REMARK 3 SYMMETRY H-BOND OTHERS (A): NULL ; NULL ; NULL
REMARK 3 SYMMETRY METAL-ION REFINED ATOMS (A): NULL ; NULL ; NULL
REMARK 3 SYMMETRY METAL-ION OTHERS (A): NULL ; NULL ; NULL
REMARK 3
REMARK 3 ISOTROPIC THERMAL FACTOR RESTRAINTS. COUNT RMS WEIGHT
REMARK 3 MAIN-CHAIN BOND REFINED ATOMS (A**2): NULL ; NULL ; NULL
REMARK 3 MAIN-CHAIN BOND OTHER ATOMS (A**2): NULL ; NULL ; NULL
REMARK 3 MAIN-CHAIN ANGLE REFINED ATOMS (A**2): NULL ; NULL ; NULL
REMARK 3 MAIN-CHAIN ANGLE OTHER ATOMS (A**2): NULL ; NULL ; NULL
REMARK 3 SIDE-CHAIN BOND REFINED ATOMS (A**2): NULL ; NULL ; NULL
REMARK 3 SIDE-CHAIN BOND OTHER ATOMS (A**2): NULL ; NULL ; NULL
REMARK 3 SIDE-CHAIN ANGLE REFINED ATOMS (A**2): NULL ; NULL ; NULL
REMARK 3 SIDE-CHAIN ANGLE OTHER ATOMS (A**2): NULL ; NULL ; NULL
REMARK 3 LONG RANGE B REFINED ATOMS (A**2): NULL ; NULL ; NULL
REMARK 3 LONG RANGE B OTHER ATOMS (A**2): NULL ; NULL ; NULL
REMARK 3
REMARK 3 ANISOTROPIC THERMAL FACTOR RESTRAINTS. COUNT RMS WEIGHT
REMARK 3 RIGID-BOND RESTRAINTS (A**2): 1002 ; 4.594 ; 3.000
REMARK 3 SPHERICITY; FREE ATOMS (A**2): 2 ; 8.840 ; 5.000
REMARK 3 SPHERICITY; BONDED ATOMS (A**2): 1134 ; 10.669 ; 5.000
REMARK 3
REMARK 3 NCS RESTRAINTS STATISTICS
REMARK 3 NUMBER OF DIFFERENT NCS GROUPS : NULL
REMARK 3
REMARK 3 TLS DETAILS
REMARK 3 NUMBER OF TLS GROUPS : NULL
REMARK 3
REMARK 3 BULK SOLVENT MODELLING.
REMARK 3 METHOD USED : MASK
REMARK 3 PARAMETERS FOR MASK CALCULATION
REMARK 3 VDW PROBE RADIUS : 1.20
REMARK 3 ION PROBE RADIUS : 0.80
REMARK 3 SHRINKAGE RADIUS : 0.80
REMARK 3
REMARK 3 OTHER REFINEMENT REMARKS: HYDROGENS HAVE BEEN ADDED IN THE RIDING
REMARK 3 POSITIONS
REMARK 4
REMARK 4 4I9V COMPLIES WITH FORMAT V. 3.30, 13-JUL-11
REMARK 100
REMARK 100 THIS ENTRY HAS BEEN PROCESSED BY RCSB ON 18-JAN-13.
REMARK 100 THE RCSB ID CODE IS RCSB076486.
REMARK 200
REMARK 200 EXPERIMENTAL DETAILS
REMARK 200 EXPERIMENT TYPE : X-RAY DIFFRACTION
REMARK 200 DATE OF DATA COLLECTION : 14-JUN-12
REMARK 200 TEMPERATURE (KELVIN) : 100
REMARK 200 PH : 7.0
REMARK 200 NUMBER OF CRYSTALS USED : 1
REMARK 200

REMARK 200 SYNCHROTRON (Y/N) : Y
 REMARK 200 RADIATION SOURCE : APS
 REMARK 200 BEAMLINE : 19-ID
 REMARK 200 X-RAY GENERATOR MODEL : NULL
 REMARK 200 MONOCHROMATIC OR LAUE (M/L) : M
 REMARK 200 WAVELENGTH OR RANGE (A) : 0.979
 REMARK 200 MONOCHROMATOR : DOUBLE CRYSTAL
 REMARK 200 OPTICS : MIRRORS
 REMARK 200
 REMARK 200 DETECTOR TYPE : CCD
 REMARK 200 DETECTOR MANUFACTURER : ADSC QUANTUM 315R
 REMARK 200 INTENSITY-INTEGRATION SOFTWARE : HKL3000
 REMARK 200 DATA SCALING SOFTWARE : HKL3000
 REMARK 200
 REMARK 200 NUMBER OF UNIQUE REFLECTIONS : 35974
 REMARK 200 RESOLUTION RANGE HIGH (A) : 1.020
 REMARK 200 RESOLUTION RANGE LOW (A) : 14.900
 REMARK 200 REJECTION CRITERIA (SIGMA(I)) : 2.000
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 REMARK 200 OVERALL.
 REMARK 200 COMPLETENESS FOR RANGE (%) : 98.0
 REMARK 200 DATA REDUNDANCY : 8.800
 REMARK 200 R MERGE (I) : 0.04300
 REMARK 200 R SYM (I) : NULL
 REMARK 200 <I/SIGMA(I)> FOR THE DATA SET : 44.6700
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 REMARK 200 IN THE HIGHEST RESOLUTION SHELL.
 REMARK 200 HIGHEST RESOLUTION SHELL, RANGE HIGH (A) : 1.02
 REMARK 200 HIGHEST RESOLUTION SHELL, RANGE LOW (A) : 1.03
 REMARK 200 COMPLETENESS FOR SHELL (%) : 99.9
 REMARK 200 DATA REDUNDANCY IN SHELL : 6.50
 REMARK 200 R MERGE FOR SHELL (I) : 0.78000
 REMARK 200 R SYM FOR SHELL (I) : NULL
 REMARK 200 <I/SIGMA(I)> FOR SHELL : NULL
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 REMARK 200 DIFFRACTION PROTOCOL: SINGLE WAVELENGTH
 REMARK 200 METHOD USED TO DETERMINE THE STRUCTURE: MOLECULAR REPLACEMENT
 REMARK 200 SOFTWARE USED: MOLREP
 REMARK 200 STARTING MODEL: PDB ENTRY 436D
 REMARK 200
 REMARK 200 REMARK: NULL
 REMARK 280
 REMARK 280 CRYSTAL
 REMARK 280 SOLVENT CONTENT, VS (%): 46.63
 REMARK 280 MATTHEWS COEFFICIENT, VM (ANGSTROMS**3/DA): 2.30
 REMARK 280
 REMARK 280 CRYSTALLIZATION CONDITIONS: 10% MPD,40MM CACODYLATE, 80MM NAACL,12
 REMARK 280 MM SPERMINE TETRACHLORIDE, 20MM MGCL2, PH 7.0, VAPOR DIFFUSION,
 REMARK 280 HANGING DROP, TEMPERATURE 291K
 REMARK 290
 REMARK 290 CRYSTALLOGRAPHIC SYMMETRY
 REMARK 290 SYMMETRY OPERATORS FOR SPACE GROUP: P 21 21 21
 REMARK 290
 REMARK 290 SYMOP SYMMETRY
 REMARK 290 NNNMMM OPERATOR
 REMARK 290 1555 X,Y,Z

REMARK 290 2555 -X+1/2,-Y,Z+1/2
REMARK 290 3555 -X,Y+1/2,-Z+1/2
REMARK 290 4555 X+1/2,-Y+1/2,-Z
REMARK 290
REMARK 290 WHERE NNN -> OPERATOR NUMBER
REMARK 290 MMM -> TRANSLATION VECTOR
REMARK 290
REMARK 290 CRYSTALLOGRAPHIC SYMMETRY TRANSFORMATIONS
REMARK 290 THE FOLLOWING TRANSFORMATIONS OPERATE ON THE ATOM/HETATM
REMARK 290 RECORDS IN THIS ENTRY TO PRODUCE CRYSTALLOGRAPHICALLY
REMARK 290 RELATED MOLECULES.
REMARK 290 SMTRY1 1 1.000000 0.000000 0.000000 0.000000
REMARK 290 SMTRY2 1 0.000000 1.000000 0.000000 0.000000
REMARK 290 SMTRY3 1 0.000000 0.000000 1.000000 0.000000
REMARK 290 SMTRY1 2 -1.000000 0.000000 0.000000 12.80300
REMARK 290 SMTRY2 2 0.000000 -1.000000 0.000000 0.000000
REMARK 290 SMTRY3 2 0.000000 0.000000 1.000000 32.15950
REMARK 290 SMTRY1 3 -1.000000 0.000000 0.000000 0.000000
REMARK 290 SMTRY2 3 0.000000 1.000000 0.000000 20.67200
REMARK 290 SMTRY3 3 0.000000 0.000000 -1.000000 32.15950
REMARK 290 SMTRY1 4 1.000000 0.000000 0.000000 12.80300
REMARK 290 SMTRY2 4 0.000000 -1.000000 0.000000 20.67200
REMARK 290 SMTRY3 4 0.000000 0.000000 -1.000000 0.000000
REMARK 290
REMARK 290 REMARK: NULL
REMARK 300
REMARK 300 BIOMOLECULE: 1
REMARK 300 SEE REMARK 350 FOR THE AUTHOR PROVIDED AND/OR PROGRAM
REMARK 300 GENERATED ASSEMBLY INFORMATION FOR THE STRUCTURE IN
REMARK 300 THIS ENTRY. THE REMARK MAY ALSO PROVIDE INFORMATION ON
REMARK 300 BURIED SURFACE AREA.
REMARK 350
REMARK 350 COORDINATES FOR A COMPLETE MULTIMER REPRESENTING THE KNOWN
REMARK 350 BIOLOGICALLY SIGNIFICANT OLIGOMERIZATION STATE OF THE
REMARK 350 MOLECULE CAN BE GENERATED BY APPLYING BIOMT TRANSFORMATIONS
REMARK 350 GIVEN BELOW. BOTH NON-CRYSTALLOGRAPHIC AND
REMARK 350 CRYSTALLOGRAPHIC OPERATIONS ARE GIVEN.
REMARK 350
REMARK 350 BIOMOLECULE: 1
REMARK 350 AUTHOR DETERMINED BIOLOGICAL UNIT: DIMERIC
REMARK 350 SOFTWARE DETERMINED QUATERNARY STRUCTURE: DIMERIC
REMARK 350 SOFTWARE USED: PISA
REMARK 350 TOTAL BURIED SURFACE AREA: 1470 ANGSTROM**2
REMARK 350 SURFACE AREA OF THE COMPLEX: 5150 ANGSTROM**2
REMARK 350 CHANGE IN SOLVENT FREE ENERGY: -11.0 KCAL/MOL
REMARK 350 APPLY THE FOLLOWING TO CHAINS: A, B
REMARK 350 BIOMT1 1 1.000000 0.000000 0.000000 0.000000
REMARK 350 BIOMT2 1 0.000000 1.000000 0.000000 0.000000
REMARK 350 BIOMT3 1 0.000000 0.000000 1.000000 0.000000
REMARK 500
REMARK 500 GEOMETRY AND STEREOCHEMISTRY
REMARK 500 SUBTOPIC: COVALENT BOND ANGLES
REMARK 500
REMARK 500 THE STEREOCHEMICAL PARAMETERS OF THE FOLLOWING RESIDUES
REMARK 500 HAVE VALUES WHICH DEVIATE FROM EXPECTED VALUES BY MORE
REMARK 500 THAN 6*RMSD (M=MODEL NUMBER; RES=RESIDUE NAME; C=CHAIN

REMARK 500 IDENTIFIER; SSEQ=SEQUENCE NUMBER; I=INSERTION CODE).

REMARK 500

REMARK 500 STANDARD TABLE:

REMARK 500 FORMAT: (10X,I3,1X,A3,1X,A1,I4,A1,3(1X,A4,2X),12X,F5.1)

REMARK 500

REMARK 500 EXPECTED VALUES PROTEIN: ENGH AND HUBER, 1999

REMARK 500 EXPECTED VALUES NUCLEIC ACID: CLOWNEY ET AL 1996

REMARK 500

REMARK 500 M RES CSSEQI ATM1 ATM2 ATM3

REMARK 500 DG A 12 O5' - P - OP2 ANGL. DEV. = -16.2 DEGREES

REMARK 500

REMARK 500 REMARK: NULL

REMARK 525

REMARK 525 SOLVENT

REMARK 525

REMARK 525 THE SOLVENT MOLECULES HAVE CHAIN IDENTIFIERS THAT

REMARK 525 INDICATE THE POLYMER CHAIN WITH WHICH THEY ARE MOST

REMARK 525 CLOSELY ASSOCIATED. THE REMARK LISTS ALL THE SOLVENT

REMARK 525 MOLECULES WHICH ARE MORE THAN 5A AWAY FROM THE

REMARK 525 NEAREST POLYMER CHAIN (M = MODEL NUMBER;

REMARK 525 RES=RESIDUE NAME; C=CHAIN IDENTIFIER; SSEQ=SEQUENCE

REMARK 525 NUMBER; I=INSERTION CODE):

REMARK 525

REMARK 525 M RES CSSEQI

REMARK 525 HOH B 283 DISTANCE = 6.25 ANGSTROMS

REMARK 525 HOH B 292 DISTANCE = 5.42 ANGSTROMS

REMARK 525 HOH B 293 DISTANCE = 6.88 ANGSTROMS

REMARK 610

REMARK 610 MISSING HETEROATOM

REMARK 610 THE FOLLOWING RESIDUES HAVE MISSING ATOMS (M=MODEL NUMBER;

REMARK 610 RES=RESIDUE NAME; C=CHAIN IDENTIFIER; SSEQ=SEQUENCE NUMBER;

REMARK 610 I=INSERTION CODE):

REMARK 610 M RES C SSEQI

REMARK 610 SPK B 101

REMARK 615

REMARK 615 ZERO OCCUPANCY ATOM

REMARK 615 THE FOLLOWING RESIDUES HAVE ATOMS MODELED WITH ZERO

REMARK 615 OCCUPANCY. THE LOCATION AND PROPERTIES OF THESE ATOMS

REMARK 615 MAY NOT BE RELIABLE. (M=MODEL NUMBER; RES=RESIDUE NAME;

REMARK 615 C=CHAIN IDENTIFIER; SSEQ=SEQUENCE NUMBER; I=INSERTION CODE):

REMARK 615 M RES C SSEQI

REMARK 615 HOH B 285

REMARK 620

REMARK 620 METAL COORDINATION

REMARK 620 (M=MODEL NUMBER; RES=RESIDUE NAME; C=CHAIN IDENTIFIER;

REMARK 620 SSEQ=SEQUENCE NUMBER; I=INSERTION CODE):

REMARK 620

REMARK 620 COORDINATION ANGLES FOR: M RES CSSEQI METAL

REMARK 620 MG A 101 MG

REMARK 620 N RES CSSEQI ATOM

REMARK 620 1 HOH B 270 O

REMARK 620 2 HOH A 208 O 93.2

REMARK 620 3 HOH A 249 O 89.9 93.8

REMARK 620 4 HOH B 206 O 179.8 86.7 90.0

REMARK 620 5 HOH B 261 O 88.8 86.2 178.7 91.4

REMARK 620 6 HOH B 262 O 92.3 173.7 89.2 87.9 90.9

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REMARK 620 N          1      2      3      4      5
REMARK 800
REMARK 800 SITE
REMARK 800 SITE_IDENTIFIER: AC1
REMARK 800 EVIDENCE_CODE: SOFTWARE
REMARK 800 SITE_DESCRIPTION: BINDING SITE FOR RESIDUE MG A 101
REMARK 800
REMARK 800 SITE_IDENTIFIER: AC2
REMARK 800 EVIDENCE_CODE: SOFTWARE
REMARK 800 SITE_DESCRIPTION: BINDING SITE FOR RESIDUE SPK B 101
REMARK 800
REMARK 800 SITE_IDENTIFIER: AC3
REMARK 800 EVIDENCE_CODE: SOFTWARE
REMARK 800 SITE_DESCRIPTION: BINDING SITE FOR RESIDUE SPK B 102
REMARK 900
REMARK 900 RELATED ENTRIES
REMARK 900 RELATED ID: 355D   RELATED DB: PDB
DBREF  4I9V A   1   12 PDB   4I9V   4I9V           1   12
DBREF  4I9V B  13   24 PDB   4I9V   4I9V           13  24
SEQRES  1 A  12  DC  DG  DC  DG  DA  DA  DT  DT 5HC  DG  DC  DG
SEQRES  1 B  12  DC  DG  DC  DG  DA  DA  DT  DT 5HC  DG  DC  DG
MODRES  4I9V 5HC A   9  DC
MODRES  4I9V 5HC B  21  DC
HET     5HC  A   9      23
HET     5HC  B  21      21
HET     MG   A 101      1
HET     SPK  B 101      11
HET     SPK  B 102      28
HETNAM   5HC 2'-DEOXY-5-(HYDROXYMETHYL)CYTIDINE 5'-(DIHYDROGEN
HETNAM   2 5HC PHOSPHATE)
HETNAM   MG MAGNESIUM ION
HETNAM   SPK SPERMINE (FULLY PROTONATED FORM)
FORMUL   1 5HC 2(C10 H16 N3 O8 P)
FORMUL   3 MG MG 2+
FORMUL   4 SPK 2(C10 H30 N4 4+)
FORMUL   6 HOH *187(H2 O)
LINK     O3' DT A 8          P 5HC A 9      1555 1555 1.59
LINK     O3' 5HC A 9          P  DG A 10      1555 1555 1.53
LINK     O3' DT B 20          P 5HC B 21      1555 1555 1.57
LINK     O3' 5HC B 21          P  DG B 22      1555 1555 1.54
LINK     MG  MG A 101          O  HOH B 270    1555 1555 2.04
LINK     MG  MG A 101          O  HOH A 208    1555 1555 2.04
LINK     MG  MG A 101          O  HOH A 249    1555 1555 2.08
LINK     MG  MG A 101          O  HOH B 206    1555 1555 2.08
LINK     MG  MG A 101          O  HOH B 261    1555 1555 2.09
LINK     MG  MG A 101          O  HOH B 262    1555 1555 2.09
SITE     1 AC1 6 HOH A 208 HOH A 249 HOH B 206 HOH B 261
SITE     2 AC1 6 HOH B 262 HOH B 270
SITE     1 AC2 7 DG A 12 HOH A 264 HOH A 271 DA B 18
SITE     2 AC2 7 DT B 19 DT B 20 HOH B 250
SITE     1 AC3 8 5HC A 9 DG A 10 HOH A 231 DG B 14
SITE     2 AC3 8 DC B 15 DC B 23 HOH B 219 HOH B 257
CRYST1  25.606 41.344 64.319 90.00 90.00 90.00 P 21 21 21 8
ORIGX1  1.000000 0.000000 0.000000 0.000000
ORIGX2  0.000000 1.000000 0.000000 0.000000
ORIGX3  0.000000 0.000000 1.000000 0.000000

```

SCALE1	0.039053	0.000000	0.000000	0.000000								
SCALE2	0.000000	0.024187	0.000000	0.000000								
SCALE3	0.000000	0.000000	0.015548	0.000000								
ATOM	1	O5'	DC	A	1	-8.013	15.825	-22.967	1.00	23.56	O	
ANISOU	1	O5'	DC	A	1	2844	1616	4488	-240	-258	343	O
ATOM	2	C5'	DC	A	1	-7.285	14.638	-23.310	1.00	20.62	C	
ANISOU	2	C5'	DC	A	1	2631	1722	3481	-370	-244	141	C
ATOM	3	C4'	DC	A	1	-8.198	13.451	-23.135	1.00	20.18	C	
ANISOU	3	C4'	DC	A	1	2716	1507	3442	-207	-150	-8	C
ATOM	4	O4'	DC	A	1	-7.529	12.266	-23.624	1.00	20.51	O	
ANISOU	4	O4'	DC	A	1	3049	1590	3154	-131	-204	173	O
ATOM	5	C3'	DC	A	1	-8.576	13.131	-21.695	1.00	21.54	C	
ANISOU	5	C3'	DC	A	1	2907	1690	3586	-95	10	73	C
ATOM	6	O3'	DC	A	1	-9.964	12.764	-21.710	1.00	23.86	O	
ANISOU	6	O3'	DC	A	1	2883	2083	4096	-18	88	201	O
ATOM	7	C2'	DC	A	1	-7.599	12.032	-21.314	1.00	20.54	C	
ANISOU	7	C2'	DC	A	1	3130	1586	3086	-78	-26	-42	C
ATOM	8	C1'	DC	A	1	-7.444	11.277	-22.613	1.00	19.55	C	
ANISOU	8	C1'	DC	A	1	2929	1498	3000	-76	-4	48	C
ATOM	9	N1	DC	A	1	-6.181	10.566	-22.816	1.00	18.68	N	
ANISOU	9	N1	DC	A	1	2789	1439	2867	-198	-219	14	N
ATOM	10	C2	DC	A	1	-6.211	9.314	-23.439	1.00	18.66	C	
ANISOU	10	C2	DC	A	1	2736	1438	2916	-397	-105	7	C
ATOM	11	O2	DC	A	1	-7.306	8.809	-23.716	1.00	20.32	O	
ANISOU	11	O2	DC	A	1	2650	1535	3535	-261	29	-236	O
ATOM	12	N3	DC	A	1	-5.046	8.678	-23.701	1.00	18.09	N	
ANISOU	12	N3	DC	A	1	2592	1496	2782	-369	-83	4	N
ATOM	13	C4	DC	A	1	-3.887	9.264	-23.398	1.00	18.74	C	
ANISOU	13	C4	DC	A	1	2478	1618	3024	-353	-272	166	C
ATOM	14	N4	DC	A	1	-2.762	8.606	-23.681	1.00	19.80	N	
ANISOU	14	N4	DC	A	1	2437	1794	3291	-381	-281	67	N
ATOM	15	C5	DC	A	1	-3.828	10.553	-22.795	1.00	19.77	C	
ANISOU	15	C5	DC	A	1	2549	1772	3188	-365	-303	-11	C
ATOM	16	C6	DC	A	1	-4.987	11.167	-22.537	1.00	19.55	C	
ANISOU	16	C6	DC	A	1	2729	1588	3109	-203	-303	35	C
ATOM	17	P	DG	A	2	-10.765	12.548	-20.383	1.00	26.04	P	
ANISOU	17	P	DG	A	2	3323	2376	4194	58	480	291	P
ATOM	18	OP1	DG	A	2	-12.130	12.923	-20.616	1.00	29.73	O	
ANISOU	18	OP1	DG	A	2	3423	2872	5002	582	819	579	O
ATOM	19	OP2	DG	A	2	-10.071	13.067	-19.272	1.00	29.00	O	
ANISOU	19	OP2	DG	A	2	3850	3431	3737	-619	532	18	O
ATOM	20	O5'	DG	A	2	-10.673	10.977	-20.163	1.00	23.68	O	
ANISOU	20	O5'	DG	A	2	2834	2190	3972	-249	301	204	O
ATOM	21	C5'	DG	A	2	-11.428	10.127	-21.004	1.00	22.65	C	
ANISOU	21	C5'	DG	A	2	2579	2588	3437	95	246	80	C
ATOM	22	C4'	DG	A	2	-11.239	8.706	-20.550	1.00	21.99	C	
ANISOU	22	C4'	DG	A	2	2253	2490	3611	-64	224	175	C
ATOM	23	O4'	DG	A	2	-9.897	8.270	-20.844	1.00	24.02	O	
ANISOU	23	O4'	DG	A	2	2351	3330	3445	116	365	-66	O
ATOM	24	C3'	DG	A	2	-11.452	8.467	-19.060	1.00	21.91	C	
ANISOU	24	C3'	DG	A	2	2809	1787	3725	124	597	386	C
ATOM	25	O3'	DG	A	2	-12.228	7.270	-19.048	1.00	29.79	O	
ANISOU	25	O3'	DG	A	2	4090	2395	4831	-635	1827	-166	O
ATOM	26	C2'	DG	A	2	-10.039	8.294	-18.532	1.00	23.04	C	
ANISOU	26	C2'	DG	A	2	2865	2403	3484	717	674	412	C
ATOM	27	C1'	DG	A	2	-9.333	7.669	-19.716	1.00	21.83	C	

ANISOU	27	C1'	DG	A	2	2651	2196	3444	64	670	29	C
ATOM	28	N9	DG	A	2	-7.886	7.824	-19.805	1.00	19.47		N
ANISOU	28	N9	DG	A	2	2494	1812	3088	481	299	129	N
ATOM	29	C8	DG	A	2	-7.132	8.909	-19.442	1.00	19.61		C
ANISOU	29	C8	DG	A	2	2708	1725	3017	309	262	187	C
ATOM	30	N7	DG	A	2	-5.865	8.760	-19.703	1.00	18.90		N
ANISOU	30	N7	DG	A	2	2713	1537	2929	313	193	178	N
ATOM	31	C5	DG	A	2	-5.778	7.502	-20.278	1.00	17.60		C
ANISOU	31	C5	DG	A	2	2289	1572	2825	209	-6	156	C
ATOM	32	C6	DG	A	2	-4.651	6.782	-20.753	1.00	16.44		C
ANISOU	32	C6	DG	A	2	2246	1491	2507	216	-71	89	C
ATOM	33	O6	DG	A	2	-3.481	7.125	-20.773	1.00	17.75		O
ANISOU	33	O6	DG	A	2	2223	1701	2818	63	-49	1	O
ATOM	34	N1	DG	A	2	-5.016	5.544	-21.263	1.00	16.93		N
ANISOU	34	N1	DG	A	2	2072	1788	2571	233	-82	76	N
ATOM	35	C2	DG	A	2	-6.291	5.053	-21.295	1.00	16.68		C
ANISOU	35	C2	DG	A	2	2139	1642	2554	257	-73	100	C
ATOM	36	N2	DG	A	2	-6.438	3.840	-21.820	1.00	17.23		N
ANISOU	36	N2	DG	A	2	1966	1849	2730	55	-53	-1	N
ATOM	37	N3	DG	A	2	-7.348	5.714	-20.863	1.00	17.69		N
ANISOU	37	N3	DG	A	2	2120	1851	2748	288	57	226	N
ATOM	38	C4	DG	A	2	-7.020	6.923	-20.371	1.00	17.73		C
ANISOU	38	C4	DG	A	2	2405	1558	2772	248	179	140	C
ATOM	39	P	DC	A	3	-13.104	6.799	-17.821	1.00	25.26		P
ANISOU	39	P	DC	A	3	3187	2125	4283	299	1239	301	P
ATOM	40	OP1	DC	A	3	-14.465	6.749	-18.257	1.00	35.26		O
ANISOU	40	OP1	DC	A	3	3301	4648	5446	1453	1135	1252	O
ATOM	41	OP2	DC	A	3	-12.786	7.522	-16.667	1.00	32.75		O
ANISOU	41	OP2	DC	A	3	5338	2468	4638	760	1702	-146	O
ATOM	42	O5'	DC	A	3	-12.717	5.272	-17.713	1.00	22.98		O
ANISOU	42	O5'	DC	A	3	2926	2212	3594	546	276	59	O
ATOM	43	C5'	DC	A	3	-12.462	4.488	-18.858	1.00	19.40		C
ANISOU	43	C5'	DC	A	3	2142	1922	3306	128	-157	139	C
ATOM	44	C4'	DC	A	3	-11.304	3.565	-18.568	1.00	18.42		C
ANISOU	44	C4'	DC	A	3	2240	1849	2907	129	-95	32	C
ATOM	45	O4'	DC	A	3	-10.083	4.304	-18.569	1.00	18.58		O
ANISOU	45	O4'	DC	A	3	1974	2115	2967	138	-104	163	O
ATOM	46	C3'	DC	A	3	-11.339	2.846	-17.219	1.00	19.10		C
ANISOU	46	C3'	DC	A	3	2263	2096	2897	76	14	26	C
ATOM	47	O3'	DC	A	3	-11.831	1.540	-17.514	1.00	20.43		O
ANISOU	47	O3'	DC	A	3	2189	2118	3455	-43	-164	266	O
ATOM	48	C2'	DC	A	3	-9.894	2.888	-16.733	1.00	19.14		C
ANISOU	48	C2'	DC	A	3	2240	2094	2937	-36	-5	240	C
ATOM	49	C1'	DC	A	3	-9.117	3.539	-17.867	1.00	18.51		C
ANISOU	49	C1'	DC	A	3	2096	2023	2913	104	-74	90	C
ATOM	50	N1	DC	A	3	-8.047	4.456	-17.482	1.00	17.85		N
ANISOU	50	N1	DC	A	3	2124	1906	2752	129	-129	61	N
ATOM	51	C2	DC	A	3	-6.730	4.200	-17.880	1.00	17.27		C
ANISOU	51	C2	DC	A	3	2255	1674	2630	257	-26	152	C
ATOM	52	O2	DC	A	3	-6.472	3.143	-18.456	1.00	17.85		O
ANISOU	52	O2	DC	A	3	2160	1840	2781	228	-168	0	O
ATOM	53	N3	DC	A	3	-5.771	5.111	-17.611	1.00	17.19		N
ANISOU	53	N3	DC	A	3	2113	1740	2679	197	-35	155	N
ATOM	54	C4	DC	A	3	-6.087	6.243	-16.986	1.00	17.32		C
ANISOU	54	C4	DC	A	3	2242	1700	2637	227	-43	161	C
ATOM	55	N4	DC	A	3	-5.113	7.114	-16.743	1.00	17.71		N

ANISOU	55	N4	DC	A	3	2282	1741	2703	151	-64	178	N
ATOM	56	C5	DC	A	3	-7.419	6.533	-16.581	1.00	18.69		C
ANISOU	56	C5	DC	A	3	2217	2088	2795	249	-27	-70	C
ATOM	57	C6	DC	A	3	-8.360	5.628	-16.858	1.00	18.34		C
ANISOU	57	C6	DC	A	3	2227	1977	2763	334	-122	14	C
ATOM	58	P	DG	A	4	-12.189	0.490	-16.381	1.00	21.70		P
ANISOU	58	P	DG	A	4	2141	2288	3814	62	78	389	P
ATOM	59	OP1	DG	A	4	-13.100	-0.475	-16.951	1.00	25.45		O
ANISOU	59	OP1	DG	A	4	2312	2407	4948	-268	-163	466	O
ATOM	60	OP2	DG	A	4	-12.537	1.163	-15.199	1.00	23.67		O
ANISOU	60	OP2	DG	A	4	2544	2838	3612	325	472	565	O
ATOM	61	O5'	DG	A	4	-10.814	-0.234	-16.073	1.00	19.80		O
ANISOU	61	O5'	DG	A	4	2078	2312	3132	26	0	292	O
ATOM	62	C5'	DG	A	4	-10.138	-0.970	-17.073	1.00	19.65		C
ANISOU	62	C5'	DG	A	4	2355	2139	2972	-104	-51	176	C
ATOM	63	C4'	DG	A	4	-8.743	-1.279	-16.592	1.00	19.72		C
ANISOU	63	C4'	DG	A	4	2463	1904	3125	9	-163	58	C
ATOM	64	O4'	DG	A	4	-7.975	-0.060	-16.507	1.00	18.58		O
ANISOU	64	O4'	DG	A	4	2216	1911	2930	16	-128	77	O
ATOM	65	C3'	DG	A	4	-8.683	-1.903	-15.197	1.00	19.70		C
ANISOU	65	C3'	DG	A	4	2537	1747	3202	46	-21	177	C
ATOM	66	O3'	DG	A	4	-7.591	-2.822	-15.160	1.00	21.01		O
ANISOU	66	O3'	DG	A	4	2506	1870	3606	86	-189	230	O
ATOM	67	C2'	DG	A	4	-8.301	-0.724	-14.327	1.00	18.78		C
ANISOU	67	C2'	DG	A	4	2183	1875	3075	-43	-138	205	C
ATOM	68	C1'	DG	A	4	-7.337	-0.055	-15.264	1.00	18.13		C
ANISOU	68	C1'	DG	A	4	2208	1817	2863	18	-13	59	C
ATOM	69	N9	DG	A	4	-6.944	1.307	-14.942	1.00	17.98		N
ANISOU	69	N9	DG	A	4	2249	1803	2780	117	-130	127	N
ATOM	70	C8	DG	A	4	-7.671	2.279	-14.307	1.00	18.66		C
ANISOU	70	C8	DG	A	4	2408	1868	2814	259	-120	109	C
ATOM	71	N7	DG	A	4	-6.999	3.380	-14.123	1.00	18.42		N
ANISOU	71	N7	DG	A	4	2272	1880	2843	269	-35	154	N
ATOM	72	C5	DG	A	4	-5.763	3.128	-14.699	1.00	17.78		C
ANISOU	72	C5	DG	A	4	2275	1757	2723	205	-135	90	C
ATOM	73	C6	DG	A	4	-4.601	3.937	-14.783	1.00	17.31		C
ANISOU	73	C6	DG	A	4	2226	1716	2633	176	-164	139	C
ATOM	74	O6	DG	A	4	-4.447	5.099	-14.406	1.00	18.39		O
ANISOU	74	O6	DG	A	4	2392	1575	3020	102	-159	111	O
ATOM	75	N1	DG	A	4	-3.554	3.265	-15.404	1.00	17.25		N
ANISOU	75	N1	DG	A	4	2251	1687	2617	101	-88	233	N
ATOM	76	C2	DG	A	4	-3.613	1.979	-15.874	1.00	17.22		C
ANISOU	76	C2	DG	A	4	2072	1835	2636	109	-150	156	C
ATOM	77	N2	DG	A	4	-2.506	1.506	-16.443	1.00	17.77		N
ANISOU	77	N2	DG	A	4	2308	1842	2600	260	-115	73	N
ATOM	78	N3	DG	A	4	-4.684	1.216	-15.793	1.00	17.40		N
ANISOU	78	N3	DG	A	4	2238	1683	2688	180	-108	117	N
ATOM	79	C4	DG	A	4	-5.708	1.843	-15.185	1.00	17.27		C
ANISOU	79	C4	DG	A	4	2265	1828	2467	82	-86	123	C
ATOM	80	P	DA	A	5	-7.823	-4.354	-15.073	1.00	26.07		P
ANISOU	80	P	DA	A	5	2700	1805	5400	-215	-281	202	P
ATOM	81	OP1	DA	A	5	-8.585	-4.719	-16.242	1.00	34.24		O
ANISOU	81	OP1	DA	A	5	3082	2918	7009	-320	-912	-1342	O
ATOM	82	OP2	DA	A	5	-8.290	-4.567	-13.720	1.00	31.70		O
ANISOU	82	OP2	DA	A	5	3493	3237	5314	714	88	1091	O
ATOM	83	O5'	DA	A	5	-6.355	-4.926	-15.183	1.00	25.02		O

ANISOU	83	O5'	DA	A	5	2898	1758	4850	161	-844	76	O
ATOM	84	C5'	DA	A	5	-5.666	-4.872	-16.400	1.00	25.58		C
ANISOU	84	C5'	DA	A	5	2828	2432	4458	326	-868	-541	C
ATOM	85	C4'	DA	A	5	-4.189	-4.656	-16.171	1.00	21.18		C
ANISOU	85	C4'	DA	A	5	2686	1952	3407	429	-622	-666	C
ATOM	86	O4'	DA	A	5	-3.928	-3.295	-15.744	1.00	21.12		O
ANISOU	86	O4'	DA	A	5	2938	1786	3298	383	-703	-353	O
ATOM	87	C3'	DA	A	5	-3.541	-5.550	-15.118	1.00	22.28		C
ANISOU	87	C3'	DA	A	5	2746	1758	3960	554	-845	-378	C
ATOM	88	O3'	DA	A	5	-2.273	-5.885	-15.698	1.00	22.49		O
ANISOU	88	O3'	DA	A	5	2905	1798	3842	404	-716	-432	O
ATOM	89	C2'	DA	A	5	-3.487	-4.650	-13.895	1.00	19.79		C
ANISOU	89	C2'	DA	A	5	2573	1456	3490	45	-505	-107	C
ATOM	90	C1'	DA	A	5	-3.224	-3.293	-14.513	1.00	18.90		C
ANISOU	90	C1'	DA	A	5	2494	1453	3232	214	-533	-253	C
ATOM	91	N9	DA	A	5	-3.680	-2.136	-13.746	1.00	18.25		N
ANISOU	91	N9	DA	A	5	2540	1495	2899	49	-403	-76	N
ATOM	92	C8	DA	A	5	-4.920	-1.933	-13.205	1.00	19.12		C
ANISOU	92	C8	DA	A	5	2614	1682	2967	-35	-321	-63	C
ATOM	93	N7	DA	A	5	-5.068	-0.760	-12.643	1.00	18.43		N
ANISOU	93	N7	DA	A	5	2467	1622	2911	17	-253	76	N
ATOM	94	C5	DA	A	5	-3.838	-0.149	-12.821	1.00	17.03		C
ANISOU	94	C5	DA	A	5	2363	1552	2554	82	-225	99	C
ATOM	95	C6	DA	A	5	-3.355	1.123	-12.469	1.00	16.52		C
ANISOU	95	C6	DA	A	5	2301	1472	2502	192	-241	59	C
ATOM	96	N6	DA	A	5	-4.076	2.035	-11.812	1.00	16.72		N
ANISOU	96	N6	DA	A	5	2257	1521	2574	104	-84	103	N
ATOM	97	N1	DA	A	5	-2.078	1.413	-12.785	1.00	16.93		N
ANISOU	97	N1	DA	A	5	2375	1477	2578	85	-115	112	N
ATOM	98	C2	DA	A	5	-1.362	0.510	-13.458	1.00	17.09		C
ANISOU	98	C2	DA	A	5	2346	1448	2698	161	-215	42	C
ATOM	99	N3	DA	A	5	-1.709	-0.709	-13.859	1.00	17.55		N
ANISOU	99	N3	DA	A	5	2484	1467	2715	217	-246	19	N
ATOM	100	C4	DA	A	5	-2.975	-0.981	-13.508	1.00	17.26		C
ANISOU	100	C4	DA	A	5	2446	1501	2611	121	-322	68	C
ATOM	101	P	DA	A	6	-1.195	-6.801	-14.946	1.00	21.29		P
ANISOU	101	P	DA	A	6	2864	1740	3485	289	-432	-76	P
ATOM	102	OP1	DA	A	6	-0.352	-7.448	-15.980	1.00	22.29		O
ANISOU	102	OP1	DA	A	6	2900	1976	3590	386	-379	-195	O
ATOM	103	OP2	DA	A	6	-1.891	-7.542	-13.910	1.00	19.81		O
ANISOU	103	OP2	DA	A	6	2433	1867	3227	-54	-414	44	O
ATOM	104	O5'	DA	A	6	-0.314	-5.761	-14.155	1.00	20.69		O
ANISOU	104	O5'	DA	A	6	2622	1862	3375	307	-645	-251	O
ATOM	105	C5'	DA	A	6	0.604	-4.951	-14.874	1.00	21.58		C
ANISOU	105	C5'	DA	A	6	2857	1900	3440	213	-357	-276	C
ATOM	106	C4'	DA	A	6	1.423	-4.227	-13.844	1.00	19.89		C
ANISOU	106	C4'	DA	A	6	2763	1941	2853	603	-317	-260	C
ATOM	107	O4'	DA	A	6	0.570	-3.264	-13.199	1.00	20.52		O
ANISOU	107	O4'	DA	A	6	2989	1949	2859	851	-198	-259	O
ATOM	108	C3'	DA	A	6	1.994	-5.115	-12.732	1.00	19.27		C
ANISOU	108	C3'	DA	A	6	2601	1821	2897	389	-337	-250	C
ATOM	109	O3'	DA	A	6	3.395	-4.886	-12.801	1.00	19.32		O
ANISOU	109	O3'	DA	A	6	2550	1974	2815	427	-148	-182	O
ATOM	110	C2'	DA	A	6	1.321	-4.617	-11.461	1.00	17.99		C
ANISOU	110	C2'	DA	A	6	2221	1647	2967	284	-245	-143	C
ATOM	111	C1'	DA	A	6	0.888	-3.212	-11.827	1.00	17.77		C

ANISOU	111	C1'	DA	A	6	2361	1681	2710	358	-182	-160	C
ATOM	112	N9	DA	A	6	-0.299	-2.724	-11.152	1.00	16.84		N
ANISOU	112	N9	DA	A	6	2298	1458	2640	197	-101	-30	N
ATOM	113	C8	DA	A	6	-1.503	-3.367	-11.041	1.00	17.41		C
ANISOU	113	C8	DA	A	6	2179	1689	2746	148	-267	-97	C
ATOM	114	N7	DA	A	6	-2.446	-2.635	-10.504	1.00	16.93		N
ANISOU	114	N7	DA	A	6	2321	1397	2713	131	-201	-48	N
ATOM	115	C5	DA	A	6	-1.816	-1.428	-10.230	1.00	16.24		C
ANISOU	115	C5	DA	A	6	2328	1373	2469	65	-90	63	C
ATOM	116	C6	DA	A	6	-2.277	-0.221	-9.670	1.00	15.50		C
ANISOU	116	C6	DA	A	6	2180	1324	2381	107	-181	125	C
ATOM	117	N6	DA	A	6	-3.524	-0.031	-9.236	1.00	16.31		N
ANISOU	117	N6	DA	A	6	2151	1475	2569	123	-91	6	N
ATOM	118	N1	DA	A	6	-1.388	0.784	-9.536	1.00	16.10		N
ANISOU	118	N1	DA	A	6	2207	1375	2535	106	-89	108	N
ATOM	119	C2	DA	A	6	-0.138	0.603	-9.981	1.00	16.39		C
ANISOU	119	C2	DA	A	6	2171	1451	2606	173	-144	98	C
ATOM	120	N3	DA	A	6	0.407	-0.473	-10.542	1.00	16.61		N
ANISOU	120	N3	DA	A	6	2295	1376	2638	182	-35	86	N
ATOM	121	C4	DA	A	6	-0.498	-1.462	-10.647	1.00	16.35		C
ANISOU	121	C4	DA	A	6	2337	1277	2595	142	-161	63	C
ATOM	122	P	DT	A	7	4.430	-5.497	-11.750	1.00	18.20		P
ANISOU	122	P	DT	A	7	2379	1638	2895	363	-152	-214	P
ATOM	123	OP1	DT	A	7	5.718	-5.600	-12.454	1.00	20.55		O
ANISOU	123	OP1	DT	A	7	2628	1954	3224	279	-51	-356	O
ATOM	124	OP2	DT	A	7	3.886	-6.716	-11.089	1.00	18.55		O
ANISOU	124	OP2	DT	A	7	2459	1509	3081	190	-349	-31	O
ATOM	125	O5'	DT	A	7	4.517	-4.365	-10.642	1.00	17.72		O
ANISOU	125	O5'	DT	A	7	2370	1501	2861	222	-90	-204	O
ATOM	126	C5'	DT	A	7	4.940	-3.056	-11.019	1.00	17.94		C
ANISOU	126	C5'	DT	A	7	2331	1632	2852	146	-12	-143	C
ATOM	127	C4'	DT	A	7	4.733	-2.109	-9.863	1.00	18.23		C
ANISOU	127	C4'	DT	A	7	2338	1624	2962	218	34	-278	C
ATOM	128	O4'	DT	A	7	3.329	-1.995	-9.571	1.00	17.48		O
ANISOU	128	O4'	DT	A	7	2323	1632	2685	250	13	-20	O
ATOM	129	C3'	DT	A	7	5.410	-2.537	-8.557	1.00	18.60		C
ANISOU	129	C3'	DT	A	7	2443	1673	2948	156	32	-264	C
ATOM	130	O3'	DT	A	7	6.509	-1.634	-8.378	1.00	20.96		O
ANISOU	130	O3'	DT	A	7	2371	2129	3463	185	17	-560	O
ATOM	131	C2'	DT	A	7	4.302	-2.460	-7.519	1.00	18.04		C
ANISOU	131	C2'	DT	A	7	2277	1750	2825	92	-144	-127	C
ATOM	132	C1'	DT	A	7	3.236	-1.629	-8.208	1.00	17.24		C
ANISOU	132	C1'	DT	A	7	2385	1470	2693	231	5	-9	C
ATOM	133	N1	DT	A	7	1.850	-1.858	-7.782	1.00	16.44		N
ANISOU	133	N1	DT	A	7	2416	1232	2596	137	-25	-24	N
ATOM	134	C2	DT	A	7	1.145	-0.804	-7.243	1.00	16.52		C
ANISOU	134	C2	DT	A	7	2376	1324	2576	257	-38	103	C
ATOM	135	O2	DT	A	7	1.637	0.291	-7.027	1.00	17.09		O
ANISOU	135	O2	DT	A	7	2339	1356	2798	195	31	-9	O
ATOM	136	N3	DT	A	7	-0.160	-1.084	-6.938	1.00	16.15		N
ANISOU	136	N3	DT	A	7	2280	1299	2556	246	-52	61	N
ATOM	137	C4	DT	A	7	-0.814	-2.289	-7.100	1.00	16.39		C
ANISOU	137	C4	DT	A	7	2333	1345	2549	103	-54	202	C
ATOM	138	O4	DT	A	7	-2.004	-2.377	-6.811	1.00	17.18		O
ANISOU	138	O4	DT	A	7	2312	1492	2722	126	-20	127	O
ATOM	139	C5	DT	A	7	-0.008	-3.366	-7.641	1.00	16.66		C

ANISOU	139	C5	DT	A	7	2439	1295	2593	83	-151	145	C
ATOM	140	C7	DT	A	7	-0.626	-4.717	-7.832	1.00	17.97		C
ANISOU	140	C7	DT	A	7	2539	1425	2864	12	-33	-64	C
ATOM	141	C6	DT	A	7	1.263	-3.096	-7.963	1.00	16.58		C
ANISOU	141	C6	DT	A	7	2381	1343	2573	144	-230	23	C
ATOM	142	P	DT	A	8	7.404	-1.655	-7.058	1.00	23.59		P
ANISOU	142	P	DT	A	8	2520	2276	4165	600	-618	-704	P
ATOM	143	OP1	DT	A	8	8.716	-1.057	-7.396	1.00	28.71		O
ANISOU	143	OP1	DT	A	8	2039	3643	5225	714	-261	-1370	O
ATOM	144	OP2	DT	A	8	7.350	-2.994	-6.446	1.00	27.86		O
ANISOU	144	OP2	DT	A	8	3104	2502	4980	856	-1331	-168	O
ATOM	145	O5'	DT	A	8	6.646	-0.681	-6.072	1.00	20.37		O
ANISOU	145	O5'	DT	A	8	2633	1887	3216	-80	-275	-232	O
ATOM	146	C5'	DT	A	8	6.439	0.676	-6.459	1.00	19.53		C
ANISOU	146	C5'	DT	A	8	2383	1973	3063	6	152	-204	C
ATOM	147	C4'	DT	A	8	5.680	1.380	-5.362	1.00	20.08		C
ANISOU	147	C4'	DT	A	8	2200	2310	3117	-54	180	-294	C
ATOM	148	O4'	DT	A	8	4.354	0.825	-5.306	1.00	20.46		O
ANISOU	148	O4'	DT	A	8	2191	2642	2941	-131	-43	-96	O
ATOM	149	C3'	DT	A	8	6.267	1.220	-3.954	1.00	22.98		C
ANISOU	149	C3'	DT	A	8	2364	3222	3145	50	-43	-773	C
ATOM	150	O3'	DT	A	8	6.556	2.525	-3.484	1.00	26.53		O
ANISOU	150	O3'	DT	A	8	2665	3750	3666	-1024	321	-1190	O
ATOM	151	C2'	DT	A	8	5.164	0.558	-3.145	1.00	20.72		C
ANISOU	151	C2'	DT	A	8	2509	2420	2944	469	-192	-281	C
ATOM	152	C1'	DT	A	8	3.924	0.828	-3.960	1.00	19.02		C
ANISOU	152	C1'	DT	A	8	2187	2171	2868	131	-5	-23	C
ATOM	153	N1	DT	A	8	2.862	-0.168	-3.853	1.00	17.78		N
ANISOU	153	N1	DT	A	8	2332	1678	2743	206	70	88	N
ATOM	154	C2	DT	A	8	1.612	0.244	-3.454	1.00	17.15		C
ANISOU	154	C2	DT	A	8	2260	1589	2665	154	57	45	C
ATOM	155	O2	DT	A	8	1.362	1.387	-3.105	1.00	17.74		O
ANISOU	155	O2	DT	A	8	2250	1559	2930	123	58	21	O
ATOM	156	N3	DT	A	8	0.659	-0.738	-3.465	1.00	17.07		N
ANISOU	156	N3	DT	A	8	2376	1457	2652	135	66	1	N
ATOM	157	C4	DT	A	8	0.827	-2.062	-3.822	1.00	17.23		C
ANISOU	157	C4	DT	A	8	2525	1496	2523	244	-42	104	C
ATOM	158	O4	DT	A	8	-0.132	-2.822	-3.797	1.00	18.34		O
ANISOU	158	O4	DT	A	8	2732	1476	2758	119	90	3	O
ATOM	159	C5	DT	A	8	2.169	-2.433	-4.217	1.00	17.88		C
ANISOU	159	C5	DT	A	8	2594	1615	2582	406	-27	187	C
ATOM	160	C7	DT	A	8	2.452	-3.851	-4.602	1.00	19.45		C
ANISOU	160	C7	DT	A	8	2818	1653	2917	457	127	104	C
ATOM	161	C6	DT	A	8	3.106	-1.476	-4.220	1.00	18.15		C
ANISOU	161	C6	DT	A	8	2474	1809	2611	411	-39	-16	C
HETATM	162	P	5HC	A	9	7.082	2.882	-2.025	1.00	26.54		P
ANISOU	162	P	5HC	A	9	2766	3801	3515	-219	-15	-949	P
HETATM	163	OP1	5HC	A	9	7.981	4.304	-2.363	1.00	32.38		O
ANISOU	163	OP1	5HC	A	9	2654	5035	4612	-898	-413	-1226	O
HETATM	164	OP2	5HC	A	9	7.671	1.403	-1.322	1.00	29.03		O
ANISOU	164	OP2	5HC	A	9	3330	4006	3691	-151	-424	-596	O
HETATM	165	O5'	5HC	A	9	5.662	3.284	-1.320	1.00	24.30		O
ANISOU	165	O5'	5HC	A	9	2482	3409	3340	-32	-331	-740	O
HETATM	166	C5'	5HC	A	9	4.856	4.263	-1.813	1.00	21.88		C
ANISOU	166	C5'	5HC	A	9	2601	2590	3121	-394	-8	-336	C
HETATM	167	C4'	5HC	A	9	3.652	4.368	-0.897	1.00	19.77		C

ANISOU	167	C4'	5HC	A	9	2579	1836	3097	-465	152	63	C
HETATM	168	O4'	5HC	A	9	2.865	3.190	-0.969	1.00	19.06		O
ANISOU	168	O4'	5HC	A	9	2488	1801	2953	-409	59	-1	O
HETATM	169	C3'	5HC	A	9	4.027	4.561	0.573	1.00	23.06		C
ANISOU	169	C3'	5HC	A	9	3570	2079	3112	-273	253	-347	C
HETATM	170	O3'	5HC	A	9	3.638	5.840	1.005	1.00	26.29		O
ANISOU	170	O3'	5HC	A	9	4596	1791	3601	126	1136	-150	O
HETATM	171	C2'	5HC	A	9	3.441	3.440	1.258	1.00	20.08		C
ANISOU	171	C2'	5HC	A	9	2447	2275	2905	-301	149	-72	C
HETATM	172	C1'	5HC	A	9	2.396	2.869	0.356	1.00	19.02		C
ANISOU	172	C1'	5HC	A	9	2534	1745	2949	-209	198	-8	C
HETATM	173	N1	5HC	A	9	2.070	1.425	0.298	1.00	17.21		N
ANISOU	173	N1	5HC	A	9	2206	1705	2626	-75	-87	94	N
HETATM	174	C2	5HC	A	9	0.816	0.996	0.402	1.00	16.83		C
ANISOU	174	C2	5HC	A	9	2188	1612	2593	-24	57	2	C
HETATM	175	O2	5HC	A	9	-0.097	1.753	0.704	1.00	18.21		O
ANISOU	175	O2	5HC	A	9	2278	1768	2872	-82	120	-19	O
HETATM	176	N3	5HC	A	9	0.543	-0.304	0.249	1.00	17.07		N
ANISOU	176	N3	5HC	A	9	2332	1658	2495	-49	85	82	N
HETATM	177	C4	5HC	A	9	1.508	-1.157	-0.060	1.00	16.89		C
ANISOU	177	C4	5HC	A	9	2355	1619	2443	80	-49	-41	C
HETATM	178	N4	5HC	A	9	1.139	-2.459	-0.257	1.00	18.16		N
ANISOU	178	N4	5HC	A	9	2606	1448	2845	201	96	41	N
HETATM	179	C5	5HC	A	9	2.826	-0.737	-0.188	1.00	18.02		C
ANISOU	179	C5	5HC	A	9	2302	1939	2604	176	-291	-16	C
HETATM	180	C5MA5HC	A	9	3.976	-1.675	-0.518	0.80	19.74			C
ANISOU	180	C5MA5HC	A	9	2375	2030	3096	171	-346	-119		C
HETATM	181	C5MB5HC	A	9	3.840	-1.760	-0.548	0.20	19.99			C
ANISOU	181	C5MB5HC	A	9	2473	2323	2798	343	-17	111		C
HETATM	182	O5	A5HC	A	9	4.206	-2.565	0.569	0.80	23.25		O
ANISOU	182	O5	A5HC	A	9	3083	2171	3576	415	-645	-123	O
HETATM	183	O5	B5HC	A	9	5.041	-1.307	-0.089	0.20	19.97		O
ANISOU	183	O5	B5HC	A	9	2202	2585	2798	567	40	208	O
HETATM	184	C6	5HC	A	9	3.048	0.595	-0.017	0.80	18.28		C
ANISOU	184	C6	5HC	A	9	2446	1898	2601	114	-193	56	C
ATOM	185	P	DG	A	10	3.993	6.501	2.333	0.80	31.69		P
ANISOU	185	P	DG	A	10	6116	1836	4088	-1307	1879	-671	P
ATOM	186	OP1	DG	A	10	3.727	7.932	2.185	1.00	41.30		O
ANISOU	186	OP1	DG	A	10	7630	3204	4857	436	2837	155	O
ATOM	187	OP2	DG	A	10	5.266	6.076	2.754	1.00	42.79		O
ANISOU	187	OP2	DG	A	10	6028	4987	5240	-432	1473	242	O
ATOM	188	O5'	DG	A	10	2.943	5.870	3.345	1.00	24.57		O
ANISOU	188	O5'	DG	A	10	3823	1829	3682	-814	939	-367	O
ATOM	189	C5'	DG	A	10	1.546	6.114	3.298	1.00	23.54		C
ANISOU	189	C5'	DG	A	10	4063	1602	3280	-248	393	-30	C
ATOM	190	C4'	DG	A	10	0.867	5.179	4.273	1.00	20.58		C
ANISOU	190	C4'	DG	A	10	2825	1678	3314	-63	377	-174	C
ATOM	191	O4'	DG	A	10	0.960	3.829	3.765	1.00	19.55		O
ANISOU	191	O4'	DG	A	10	2631	1635	3158	-41	116	-220	O
ATOM	192	C3'	DG	A	10	1.499	5.138	5.669	1.00	21.57		C
ANISOU	192	C3'	DG	A	10	3088	1887	3219	-463	429	-151	C
ATOM	193	O3'	DG	A	10	0.495	5.018	6.673	1.00	21.79		O
ANISOU	193	O3'	DG	A	10	3075	1857	3345	-387	447	-322	O
ATOM	194	C2'	DG	A	10	2.279	3.840	5.658	1.00	21.13		C
ANISOU	194	C2'	DG	A	10	2768	2093	3164	-405	15	-263	C
ATOM	195	C1'	DG	A	10	1.322	3.003	4.850	1.00	19.40		C

ANISOU	195	C1'	DG	A	10	2396	1821	3153	-116	45	-215	C
ATOM	196	N9	DG	A	10	1.779	1.714	4.331	1.00	18.52		N
ANISOU	196	N9	DG	A	10	2351	1743	2940	-69	-65	-24	N
ATOM	197	C8	DG	A	10	3.046	1.308	3.996	1.00	18.90		C
ANISOU	197	C8	DG	A	10	2247	1971	2963	-163	-68	-83	C
ATOM	198	N7	DG	A	10	3.090	0.083	3.552	1.00	18.74		N
ANISOU	198	N7	DG	A	10	2197	1965	2957	-51	-68	-46	N
ATOM	199	C5	DG	A	10	1.767	-0.339	3.588	1.00	17.85		C
ANISOU	199	C5	DG	A	10	2187	1851	2741	-33	-74	172	C
ATOM	200	C6	DG	A	10	1.183	-1.591	3.244	1.00	17.40		C
ANISOU	200	C6	DG	A	10	2394	1588	2627	111	-46	55	C
ATOM	201	O6	DG	A	10	1.728	-2.595	2.789	1.00	18.91		O
ANISOU	201	O6	DG	A	10	2504	1719	2959	176	62	36	O
ATOM	202	N1	DG	A	10	-0.189	-1.589	3.463	1.00	17.26		N
ANISOU	202	N1	DG	A	10	2312	1545	2701	49	-48	33	N
ATOM	203	C2	DG	A	10	-0.906	-0.536	3.965	1.00	17.06		C
ANISOU	203	C2	DG	A	10	2255	1514	2712	-10	105	67	C
ATOM	204	N2	DG	A	10	-2.214	-0.720	4.110	1.00	17.62		N
ANISOU	204	N2	DG	A	10	2307	1538	2850	-86	71	17	N
ATOM	205	N3	DG	A	10	-0.377	0.634	4.268	1.00	17.60		N
ANISOU	205	N3	DG	A	10	2219	1625	2843	31	-14	31	N
ATOM	206	C4	DG	A	10	0.950	0.660	4.060	1.00	18.03		C
ANISOU	206	C4	DG	A	10	2280	1761	2808	-85	10	186	C
ATOM	207	P	DC	A	11	-0.029	6.285	7.476	1.00	22.76		P
ANISOU	207	P	DC	A	11	3301	1860	3487	-318	438	-338	P
ATOM	208	OP1	DC	A	11	0.033	7.472	6.653	1.00	25.52		O
ANISOU	208	OP1	DC	A	11	3766	1740	4188	-541	454	-97	O
ATOM	209	OP2	DC	A	11	0.628	6.239	8.711	1.00	26.17		O
ANISOU	209	OP2	DC	A	11	3624	2717	3601	-445	471	-400	O
ATOM	210	O5'	DC	A	11	-1.524	5.867	7.785	1.00	22.04		O
ANISOU	210	O5'	DC	A	11	3192	1774	3407	24	231	-324	O
ATOM	211	C5'	DC	A	11	-2.505	5.873	6.774	1.00	22.16		C
ANISOU	211	C5'	DC	A	11	3134	1800	3485	12	398	9	C
ATOM	212	C4'	DC	A	11	-3.490	4.750	6.985	1.00	20.24		C
ANISOU	212	C4'	DC	A	11	2880	1639	3171	210	297	72	C
ATOM	213	O4'	DC	A	11	-2.857	3.481	6.709	1.00	20.19		O
ANISOU	213	O4'	DC	A	11	3123	1546	3000	172	262	99	O
ATOM	214	C3'	DC	A	11	-4.075	4.612	8.381	1.00	20.69		C
ANISOU	214	C3'	DC	A	11	3085	1601	3174	120	279	61	C
ATOM	215	O3'	DC	A	11	-5.423	4.210	8.168	1.00	23.34		O
ANISOU	215	O3'	DC	A	11	3019	2382	3465	156	306	306	O
ATOM	216	C2'	DC	A	11	-3.234	3.505	8.997	1.00	20.20		C
ANISOU	216	C2'	DC	A	11	2991	1629	3054	94	257	61	C
ATOM	217	C1'	DC	A	11	-3.018	2.594	7.809	1.00	19.06		C
ANISOU	217	C1'	DC	A	11	2567	1737	2936	70	47	126	C
ATOM	218	N1	DC	A	11	-1.842	1.729	7.823	1.00	17.91		N
ANISOU	218	N1	DC	A	11	2391	1590	2823	2	-2	62	N
ATOM	219	C2	DC	A	11	-1.981	0.394	7.430	1.00	17.91		C
ANISOU	219	C2	DC	A	11	2404	1585	2816	-105	35	62	C
ATOM	220	O2	DC	A	11	-3.108	-0.047	7.223	1.00	18.39		O
ANISOU	220	O2	DC	A	11	2409	1571	3005	55	-58	33	O
ATOM	221	N3	DC	A	11	-0.883	-0.375	7.317	1.00	17.84		N
ANISOU	221	N3	DC	A	11	2386	1632	2758	-136	-137	37	N
ATOM	222	C4	DC	A	11	0.322	0.145	7.544	1.00	18.55		C
ANISOU	222	C4	DC	A	11	2443	1730	2874	-127	-164	-45	C
ATOM	223	N4	DC	A	11	1.380	-0.634	7.367	1.00	18.95		N

ANISOU	223	N4	DC	A	11	2374	1795	3030	-121	-173	86	N
ATOM	224	C5	DC	A	11	0.495	1.510	7.915	1.00	19.97		C
ANISOU	224	C5	DC	A	11	2655	1736	3195	-333	-45	-121	C
ATOM	225	C6	DC	A	11	-0.603	2.259	8.035	1.00	19.96		C
ANISOU	225	C6	DC	A	11	2550	1925	3108	-252	1	-54	C
ATOM	226	P	DG	A	12	-6.471	4.222	9.346	1.00	27.72		P
ANISOU	226	P	DG	A	12	3890	2384	4257	449	1216	243	P
ATOM	227	OP1	DG	A	12	-7.712	4.652	8.853	1.00	27.56		O
ANISOU	227	OP1	DG	A	12	4070	1611	4787	430	1126	204	O
ATOM	228	OP2	DG	A	12	-5.880	4.726	10.519	1.00	32.88		O
ANISOU	228	OP2	DG	A	12	3826	4655	4011	873	933	779	O
ATOM	229	O5'	DG	A	12	-6.416	2.793	9.948	1.00	33.13		O
ANISOU	229	O5'	DG	A	12	4367	3386	4833	413	740	1459	O
ATOM	230	C5'	DG	A	12	-6.983	1.731	9.279	1.00	26.12		C
ANISOU	230	C5'	DG	A	12	4018	2511	3394	-1214	-8	-405	C
ATOM	231	C4'	DG	A	12	-7.091	0.575	10.253	1.00	19.74		C
ANISOU	231	C4'	DG	A	12	2340	2197	2962	2	-51	154	C
ATOM	232	O4'	DG	A	12	-5.967	-0.320	10.200	1.00	26.91		O
ANISOU	232	O4'	DG	A	12	2590	4225	3408	1099	-236	-570	O
ATOM	233	C3'	DG	A	12	-7.276	0.962	11.716	1.00	19.40		C
ANISOU	233	C3'	DG	A	12	2370	1821	3180	-52	-15	-17	C
ATOM	234	O3'	DG	A	12	-8.437	0.297	12.187	1.00	19.83		O
ANISOU	234	O3'	DG	A	12	2304	1934	3295	151	109	151	O
ATOM	235	C2'	DG	A	12	-6.016	0.476	12.412	1.00	20.33		C
ANISOU	235	C2'	DG	A	12	2408	2328	2987	-67	-80	-149	C
ATOM	236	C1'	DG	A	12	-5.400	-0.543	11.466	1.00	21.19		C
ANISOU	236	C1'	DG	A	12	2293	2660	3098	303	-126	-67	C
ATOM	237	N9	DG	A	12	-3.955	-0.487	11.294	1.00	19.72		N
ANISOU	237	N9	DG	A	12	2266	2133	3091	-6	-95	82	N
ATOM	238	C8	DG	A	12	-3.141	0.613	11.373	1.00	19.31		C
ANISOU	238	C8	DG	A	12	2328	2080	2928	17	-97	287	C
ATOM	239	N7	DG	A	12	-1.898	0.350	11.081	1.00	19.76		N
ANISOU	239	N7	DG	A	12	2268	2160	3079	-209	-74	138	N
ATOM	240	C5	DG	A	12	-1.897	-1.000	10.763	1.00	18.91		C
ANISOU	240	C5	DG	A	12	2280	1971	2934	-80	-157	284	C
ATOM	241	C6	DG	A	12	-0.832	-1.858	10.375	1.00	18.84		C
ANISOU	241	C6	DG	A	12	2165	2137	2855	-25	66	194	C
ATOM	242	O6	DG	A	12	0.355	-1.585	10.213	1.00	20.63		O
ANISOU	242	O6	DG	A	12	2267	2267	3303	-172	-45	277	O
ATOM	243	N1	DG	A	12	-1.277	-3.156	10.156	1.00	19.50		N
ANISOU	243	N1	DG	A	12	2201	2162	3043	-22	-75	190	N
ATOM	244	C2	DG	A	12	-2.566	-3.583	10.311	1.00	18.89		C
ANISOU	244	C2	DG	A	12	2027	2360	2788	-74	57	-101	C
ATOM	245	N2	DG	A	12	-2.790	-4.876	10.081	1.00	19.80		N
ANISOU	245	N2	DG	A	12	2201	2206	3113	204	41	-134	N
ATOM	246	N3	DG	A	12	-3.562	-2.797	10.675	1.00	18.80		N
ANISOU	246	N3	DG	A	12	2023	2068	3052	-52	-19	-13	N
ATOM	247	C4	DG	A	12	-3.160	-1.528	10.878	1.00	18.99		C
ANISOU	247	C4	DG	A	12	2088	2251	2875	30	-17	175	C
TER	248		DG	A	12							
ATOM	249	O5'	DC	B	13	4.517	-9.654	9.217	1.00	35.24		O
ANISOU	249	O5'	DC	B	13	3239	4591	5560	696	60	-31	O
ATOM	250	C5'	DC	B	13	3.797	-10.590	10.029	1.00	29.09		C
ANISOU	250	C5'	DC	B	13	2877	3536	4637	630	-305	-548	C
ATOM	251	C4'	DC	B	13	2.328	-10.515	9.691	1.00	25.51		C
ANISOU	251	C4'	DC	B	13	2689	2485	4518	346	-199	-305	C

ATOM	252	O4'	DC	B	13	1.738	-9.234	9.994	1.00	23.51		O
ANISOU	252	O4'	DC	B	13	2730	2316	3886	324	-133	-160	O
ATOM	253	C3'	DC	B	13	1.960	-10.856	8.245	1.00	25.96		C
ANISOU	253	C3'	DC	B	13	3069	2481	4314	175	70	-67	C
ATOM	254	O3'	DC	B	13	0.875	-11.787	8.202	1.00	26.95		O
ANISOU	254	O3'	DC	B	13	3325	2033	4881	190	-436	-427	O
ATOM	255	C2'	DC	B	13	1.552	-9.526	7.641	1.00	25.51		C
ANISOU	255	C2'	DC	B	13	2903	2837	3952	399	152	115	C
ATOM	256	C1'	DC	B	13	1.112	-8.680	8.842	1.00	24.13		C
ANISOU	256	C1'	DC	B	13	2889	2273	4003	665	250	230	C
ATOM	257	N1	DC	B	13	1.468	-7.247	8.815	1.00	22.20		N
ANISOU	257	N1	DC	B	13	2316	2634	3481	76	76	-41	N
ATOM	258	C2	DC	B	13	0.501	-6.289	9.162	1.00	21.44		C
ANISOU	258	C2	DC	B	13	2363	2222	3561	96	-246	151	C
ATOM	259	O2	DC	B	13	-0.664	-6.654	9.353	1.00	21.81		O
ANISOU	259	O2	DC	B	13	2494	2316	3475	307	43	109	O
ATOM	260	N3	DC	B	13	0.855	-4.985	9.223	1.00	20.55		N
ANISOU	260	N3	DC	B	13	2163	2487	3155	24	-98	6	N
ATOM	261	C4	DC	B	13	2.116	-4.626	8.981	1.00	20.39		C
ANISOU	261	C4	DC	B	13	2172	2394	3179	-127	-35	-28	C
ATOM	262	N4	DC	B	13	2.423	-3.331	9.071	1.00	21.23		N
ANISOU	262	N4	DC	B	13	2287	2420	3359	-139	-191	-34	N
ATOM	263	C5	DC	B	13	3.130	-5.583	8.679	1.00	22.11		C
ANISOU	263	C5	DC	B	13	2214	2522	3664	-65	211	-37	C
ATOM	264	C6	DC	B	13	2.769	-6.871	8.623	1.00	23.23		C
ANISOU	264	C6	DC	B	13	2309	2561	3957	-85	-98	-33	C
ATOM	265	P	DG	B	14	0.431	-12.476	6.825	1.00	29.20		P
ANISOU	265	P	DG	B	14	3962	2238	4896	349	-420	-442	P
ATOM	266	OP1	DG	B	14	-0.166	-13.796	7.166	1.00	34.49		O
ANISOU	266	OP1	DG	B	14	4505	1899	6698	514	-722	-331	O
ATOM	267	OP2	DG	B	14	1.568	-12.436	5.862	1.00	36.05		O
ANISOU	267	OP2	DG	B	14	3738	3314	6642	912	399	-1545	O
ATOM	268	O5'	DG	B	14	-0.698	-11.466	6.343	1.00	24.63		O
ANISOU	268	O5'	DG	B	14	2953	2188	4215	394	-145	-598	O
ATOM	269	C5'	DG	B	14	-1.863	-11.411	7.143	1.00	22.11		C
ANISOU	269	C5'	DG	B	14	3129	1915	3354	112	-296	-36	C
ATOM	270	C4'	DG	B	14	-2.697	-10.233	6.720	1.00	19.91		C
ANISOU	270	C4'	DG	B	14	2493	1700	3370	-177	128	-262	C
ATOM	271	O4'	DG	B	14	-1.969	-9.030	6.977	1.00	21.17		O
ANISOU	271	O4'	DG	B	14	2653	1536	3855	-37	80	-252	O
ATOM	272	C3'	DG	B	14	-2.997	-10.225	5.224	1.00	21.45		C
ANISOU	272	C3'	DG	B	14	2602	1974	3571	17	-115	107	C
ATOM	273	O3'	DG	B	14	-4.371	-10.525	5.087	1.00	20.62		O
ANISOU	273	O3'	DG	B	14	2691	1520	3622	-268	19	-14	O
ATOM	274	C2'	DG	B	14	-2.672	-8.817	4.768	1.00	21.93		C
ANISOU	274	C2'	DG	B	14	2879	2072	3381	-240	-33	2	C
ATOM	275	C1'	DG	B	14	-2.439	-8.063	6.061	1.00	21.04		C
ANISOU	275	C1'	DG	B	14	2781	1410	3802	-16	-110	-200	C
ATOM	276	N9	DG	B	14	-1.411	-7.044	5.948	1.00	20.97		N
ANISOU	276	N9	DG	B	14	2391	1492	4082	48	-77	-164	N
ATOM	277	C8	DG	B	14	-0.116	-7.261	5.545	1.00	21.40		C
ANISOU	277	C8	DG	B	14	2366	1770	3993	-6	-51	1	C
ATOM	278	N7	DG	B	14	0.610	-6.179	5.555	1.00	20.65		N
ANISOU	278	N7	DG	B	14	2433	1745	3665	-49	-12	-171	N
ATOM	279	C5	DG	B	14	-0.254	-5.189	6.000	1.00	18.90		C
ANISOU	279	C5	DG	B	14	2265	1654	3258	18	-146	31	C

ATOM	280	C6	DG	B	14	-0.032	-3.809	6.197	1.00	17.21		C
ANISOU	280	C6	DG	B	14	2102	1591	2844	-86	-125	130	C
ATOM	281	O6	DG	B	14	1.018	-3.177	6.059	1.00	18.11		O
ANISOU	281	O6	DG	B	14	2246	1643	2988	-120	-102	115	O
ATOM	282	N1	DG	B	14	-1.176	-3.164	6.656	1.00	17.18		N
ANISOU	282	N1	DG	B	14	2187	1596	2743	-130	-103	48	N
ATOM	283	C2	DG	B	14	-2.390	-3.768	6.874	1.00	17.31		C
ANISOU	283	C2	DG	B	14	2208	1500	2868	-86	-104	65	C
ATOM	284	N2	DG	B	14	-3.381	-2.964	7.296	1.00	17.84		N
ANISOU	284	N2	DG	B	14	2293	1547	2937	-129	-21	151	N
ATOM	285	N3	DG	B	14	-2.611	-5.064	6.695	1.00	18.31		N
ANISOU	285	N3	DG	B	14	2186	1454	3316	-84	-61	39	N
ATOM	286	C4	DG	B	14	-1.504	-5.708	6.260	1.00	18.67		C
ANISOU	286	C4	DG	B	14	2256	1414	3420	15	-82	84	C
ATOM	287	P	DC	B	15	-5.063	-10.543	3.652	1.00	21.03		P
ANISOU	287	P	DC	B	15	3190	1621	3179	-138	135	-121	P
ATOM	288	OP1	DC	B	15	-6.189	-11.484	3.750	1.00	22.84		O
ANISOU	288	OP1	DC	B	15	3755	1536	3385	-565	-10	-97	O
ATOM	289	OP2	DC	B	15	-4.025	-10.756	2.613	1.00	24.83		O
ANISOU	289	OP2	DC	B	15	3865	2288	3281	412	393	-217	O
ATOM	290	O5'	DC	B	15	-5.581	-9.055	3.465	1.00	20.14		O
ANISOU	290	O5'	DC	B	15	2887	1744	3021	-155	160	4	O
ATOM	291	C5'	DC	B	15	-6.578	-8.504	4.328	1.00	18.89		C
ANISOU	291	C5'	DC	B	15	2582	1743	2850	-313	119	45	C
ATOM	292	C4'	DC	B	15	-6.552	-6.999	4.236	1.00	19.01		C
ANISOU	292	C4'	DC	B	15	2726	1712	2784	-285	10	-81	C
ATOM	293	O4'	DC	B	15	-5.232	-6.528	4.539	1.00	20.19		O
ANISOU	293	O4'	DC	B	15	2992	1572	3108	-371	-280	114	O
ATOM	294	C3'	DC	B	15	-6.829	-6.413	2.861	1.00	18.81		C
ANISOU	294	C3'	DC	B	15	2590	1767	2788	-312	163	-65	C
ATOM	295	O3'	DC	B	15	-8.231	-6.410	2.647	1.00	19.90		O
ANISOU	295	O3'	DC	B	15	2690	1898	2971	-407	70	66	O
ATOM	296	C2'	DC	B	15	-6.216	-5.031	2.975	1.00	20.48		C
ANISOU	296	C2'	DC	B	15	2816	1765	3197	-333	-105	162	C
ATOM	297	C1'	DC	B	15	-5.060	-5.248	3.921	1.00	19.84		C
ANISOU	297	C1'	DC	B	15	2742	1594	3203	-157	-39	96	C
ATOM	298	N1	DC	B	15	-3.727	-5.223	3.329	1.00	19.11		N
ANISOU	298	N1	DC	B	15	2686	1538	3033	-62	-94	9	N
ATOM	299	C2	DC	B	15	-2.994	-4.043	3.428	1.00	17.83		C
ANISOU	299	C2	DC	B	15	2512	1420	2842	-48	94	56	C
ATOM	300	O2	DC	B	15	-3.542	-3.037	3.890	1.00	18.22		O
ANISOU	300	O2	DC	B	15	2483	1386	3052	33	31	1	O
ATOM	301	N3	DC	B	15	-1.714	-4.022	3.005	1.00	18.25		N
ANISOU	301	N3	DC	B	15	2587	1499	2845	150	93	-20	N
ATOM	302	C4	DC	B	15	-1.186	-5.105	2.435	1.00	20.21		C
ANISOU	302	C4	DC	B	15	2825	1732	3120	178	40	-246	C
ATOM	303	N4	DC	B	15	0.077	-5.036	2.018	1.00	21.23		N
ANISOU	303	N4	DC	B	15	2944	1752	3368	267	128	-226	N
ATOM	304	C5	DC	B	15	-1.932	-6.305	2.260	1.00	22.52		C
ANISOU	304	C5	DC	B	15	3269	1589	3695	113	110	-150	C
ATOM	305	C6	DC	B	15	-3.189	-6.321	2.719	1.00	21.63		C
ANISOU	305	C6	DC	B	15	3028	1720	3470	54	-94	-319	C
ATOM	306	P	DG	B	16	-8.845	-6.049	1.228	1.00	21.07		P
ANISOU	306	P	DG	B	16	2850	1989	3167	-448	-202	5	P
ATOM	307	OP1	DG	B	16	-10.248	-6.523	1.248	1.00	24.23		O
ANISOU	307	OP1	DG	B	16	3275	2365	3566	-930	-400	84	O

ATOM	308	OP2	DG	B	16	-7.938	-6.522	0.147	1.00	22.93		O
ANISOU	308	OP2	DG	B	16	3515	2169	3028	-241	-230	-197	O
ATOM	309	O5'	DG	B	16	-8.836	-4.461	1.193	1.00	19.96		O
ANISOU	309	O5'	DG	B	16	2604	2012	2965	-340	47	67	O
ATOM	310	C5'	DG	B	16	-9.564	-3.684	2.157	1.00	19.81		C
ANISOU	310	C5'	DG	B	16	2461	2100	2964	-53	146	226	C
ATOM	311	C4'	DG	B	16	-9.280	-2.221	1.926	1.00	20.01		C
ANISOU	311	C4'	DG	B	16	2526	2200	2874	-418	-75	198	C
ATOM	312	O4'	DG	B	16	-7.856	-1.984	2.051	1.00	18.88		O
ANISOU	312	O4'	DG	B	16	2411	1926	2834	-218	-1	135	O
ATOM	313	C3'	DG	B	16	-9.675	-1.681	0.548	1.00	20.05		C
ANISOU	313	C3'	DG	B	16	2263	2214	3139	5	-121	302	C
ATOM	314	O3'	DG	B	16	-10.152	-0.343	0.773	1.00	20.65		O
ANISOU	314	O3'	DG	B	16	2440	2246	3159	9	40	392	O
ATOM	315	C2'	DG	B	16	-8.390	-1.789	-0.242	1.00	20.01		C
ANISOU	315	C2'	DG	B	16	2515	2128	2961	-309	-23	99	C
ATOM	316	C1'	DG	B	16	-7.330	-1.541	0.813	1.00	18.79		C
ANISOU	316	C1'	DG	B	16	2347	1878	2913	-138	-45	201	C
ATOM	317	N9	DG	B	16	-6.079	-2.241	0.534	1.00	17.72		N
ANISOU	317	N9	DG	B	16	2405	1644	2683	-173	71	52	N
ATOM	318	C8	DG	B	16	-5.915	-3.477	-0.043	1.00	18.68		C
ANISOU	318	C8	DG	B	16	2563	1660	2871	-166	61	-2	C
ATOM	319	N7	DG	B	16	-4.664	-3.778	-0.263	1.00	19.08		N
ANISOU	319	N7	DG	B	16	2610	1786	2851	-191	3	-132	N
ATOM	320	C5	DG	B	16	-3.963	-2.665	0.182	1.00	17.51		C
ANISOU	320	C5	DG	B	16	2480	1544	2627	-65	16	85	C
ATOM	321	C6	DG	B	16	-2.573	-2.375	0.145	1.00	17.43		C
ANISOU	321	C6	DG	B	16	2487	1461	2674	-46	60	23	C
ATOM	322	O6	DG	B	16	-1.648	-3.088	-0.261	1.00	18.30		O
ANISOU	322	O6	DG	B	16	2679	1565	2710	27	216	14	O
ATOM	323	N1	DG	B	16	-2.301	-1.099	0.634	1.00	17.05		N
ANISOU	323	N1	DG	B	16	2271	1546	2661	-57	66	7	N
ATOM	324	C2	DG	B	16	-3.240	-0.227	1.122	1.00	16.74		C
ANISOU	324	C2	DG	B	16	2259	1502	2598	-36	103	159	C
ATOM	325	N2	DG	B	16	-2.782	0.963	1.547	1.00	17.18		N
ANISOU	325	N2	DG	B	16	2339	1447	2740	-20	59	66	N
ATOM	326	N3	DG	B	16	-4.533	-0.492	1.175	1.00	16.99		N
ANISOU	326	N3	DG	B	16	2311	1440	2705	-52	6	116	N
ATOM	327	C4	DG	B	16	-4.824	-1.707	0.669	1.00	16.89		C
ANISOU	327	C4	DG	B	16	2318	1517	2580	-104	78	139	C
ATOM	328	P	DA	B	17	-10.173	0.833	-0.331	1.00	21.05		P
ANISOU	328	P	DA	B	17	2404	2346	3245	68	6	434	P
ATOM	329	OP1	DA	B	17	-11.279	1.754	0.053	1.00	22.46		O
ANISOU	329	OP1	DA	B	17	2387	2667	3480	198	90	506	O
ATOM	330	OP2	DA	B	17	-10.137	0.270	-1.695	1.00	23.21		O
ANISOU	330	OP2	DA	B	17	2612	2778	3429	56	-122	473	O
ATOM	331	O5'	DA	B	17	-8.787	1.572	-0.094	1.00	19.99		O
ANISOU	331	O5'	DA	B	17	2363	2225	3005	-3	158	298	O
ATOM	332	C5'	DA	B	17	-8.470	2.156	1.180	1.00	19.29		C
ANISOU	332	C5'	DA	B	17	2412	2049	2865	113	265	191	C
ATOM	333	C4'	DA	B	17	-7.340	3.135	0.999	1.00	18.87		C
ANISOU	333	C4'	DA	B	17	2286	1933	2949	238	249	267	C
ATOM	334	O4'	DA	B	17	-6.147	2.432	0.593	1.00	18.85		O
ANISOU	334	O4'	DA	B	17	2337	1937	2886	223	254	194	O
ATOM	335	C3'	DA	B	17	-7.584	4.188	-0.080	1.00	19.37		C
ANISOU	335	C3'	DA	B	17	2380	1815	3163	238	380	272	C

ATOM	336	O3'	DA	B	17	-7.017	5.418	0.430	1.00	20.63	O	
ANISOU	336	O3'	DA	B	17	2684	1723	3429	200	596	140	O
ATOM	337	C2'	DA	B	17	-6.885	3.614	-1.307	1.00	18.90	C	
ANISOU	337	C2'	DA	B	17	2333	1771	3077	208	203	212	C
ATOM	338	C1'	DA	B	17	-5.712	2.872	-0.702	1.00	17.85	C	
ANISOU	338	C1'	DA	B	17	2243	1683	2856	163	172	130	C
ATOM	339	N9	DA	B	17	-5.309	1.684	-1.439	1.00	17.40	N	
ANISOU	339	N9	DA	B	17	2284	1614	2712	0	186	206	N
ATOM	340	C8	DA	B	17	-6.124	0.642	-1.798	1.00	18.01	C	
ANISOU	340	C8	DA	B	17	2212	1866	2764	27	130	153	C
ATOM	341	N7	DA	B	17	-5.486	-0.359	-2.353	1.00	17.59	N	
ANISOU	341	N7	DA	B	17	2314	1681	2687	-19	140	159	N
ATOM	342	C5	DA	B	17	-4.156	0.031	-2.318	1.00	16.97	C	
ANISOU	342	C5	DA	B	17	2251	1542	2652	-97	65	146	C
ATOM	343	C6	DA	B	17	-2.974	-0.599	-2.743	1.00	16.69	C	
ANISOU	343	C6	DA	B	17	2336	1550	2455	64	92	114	C
ATOM	344	N6	DA	B	17	-2.932	-1.823	-3.290	1.00	17.33	N	
ANISOU	344	N6	DA	B	17	2408	1541	2635	1	30	40	N
ATOM	345	N1	DA	B	17	-1.814	0.077	-2.582	1.00	16.97	N	
ANISOU	345	N1	DA	B	17	2457	1347	2642	-7	153	155	N
ATOM	346	C2	DA	B	17	-1.848	1.292	-2.020	1.00	16.80	C	
ANISOU	346	C2	DA	B	17	2291	1470	2621	102	214	99	C
ATOM	347	N3	DA	B	17	-2.893	1.980	-1.567	1.00	16.78	N	
ANISOU	347	N3	DA	B	17	2203	1492	2678	98	197	192	N
ATOM	348	C4	DA	B	17	-4.031	1.288	-1.754	1.00	16.78	C	
ANISOU	348	C4	DA	B	17	2203	1602	2568	30	168	57	C
ATOM	349	P A	DA	B	18	-6.973	6.716	-0.455	0.80	21.71	P	
ANISOU	349	P A	DA	B	18	2701	1716	3831	375	459	196	P
ATOM	350	P B	DA	B	18	-7.014	6.862	-0.413	0.20	23.21	P	
ANISOU	350	P B	DA	B	18	2871	1940	4007	93	383	304	P
ATOM	351	OP1A	DA	B	18	-6.961	7.867	0.460	0.80	24.47	O	
ANISOU	351	OP1A	DA	B	18	3353	1705	4239	532	835	-131	O
ATOM	352	OP1B	DA	B	18	-6.985	7.995	0.560	0.20	26.26	O	
ANISOU	352	OP1B	DA	B	18	3379	2319	4280	66	427	-23	O
ATOM	353	OP2A	DA	B	18	-7.993	6.634	-1.528	0.80	23.59	O	
ANISOU	353	OP2A	DA	B	18	2990	1986	3987	300	341	348	O
ATOM	354	OP2B	DA	B	18	-8.071	6.823	-1.458	0.20	23.62	O	
ANISOU	354	OP2B	DA	B	18	3151	1777	4045	595	148	686	O
ATOM	355	O5'A	DA	B	18	-5.571	6.655	-1.191	0.80	20.14	O	
ANISOU	355	O5'A	DA	B	18	2782	1589	3280	137	397	100	O
ATOM	356	O5'B	DA	B	18	-5.608	6.872	-1.164	0.20	21.93	O	
ANISOU	356	O5'B	DA	B	18	2847	1990	3496	128	285	167	O
ATOM	357	C5'A	DA	B	18	-4.360	6.670	-0.449	0.80	19.88	C	
ANISOU	357	C5'A	DA	B	18	2659	1697	3194	-35	432	14	C
ATOM	358	C5'B	DA	B	18	-4.362	6.853	-0.449	0.20	21.44	C	
ANISOU	358	C5'B	DA	B	18	2815	2083	3246	246	378	10	C
ATOM	359	C4'A	DA	B	18	-3.211	6.593	-1.418	0.80	20.12	C	
ANISOU	359	C4'A	DA	B	18	3160	1160	3325	-1	716	27	C
ATOM	360	C4'B	DA	B	18	-3.225	6.832	-1.441	0.20	21.41	C	
ANISOU	360	C4'B	DA	B	18	3152	1772	3210	418	522	-87	C
ATOM	361	O4'A	DA	B	18	-3.190	5.295	-2.035	0.80	19.35	O	
ANISOU	361	O4'A	DA	B	18	3122	1202	3026	269	504	0	O
ATOM	362	O4'B	DA	B	18	-3.116	5.515	-2.021	0.20	20.69	O	
ANISOU	362	O4'B	DA	B	18	3120	1687	3053	336	437	17	O
ATOM	363	C3'A	DA	B	18	-3.249	7.615	-2.562	0.80	27.46	C	
ANISOU	363	C3'A	DA	B	18	5158	1849	3425	1195	1741	245	C

ATOM	364	C3'B	DA	B	18	-3.398	7.804	-2.611	0.20	28.34		C
ANISOU	364	C3'B	DA	B	18	5179	2142	3446	1490	841	113	C
ATOM	365	O3'A	DA	B	18	-2.021	8.360	-2.363	0.80	37.27		O
ANISOU	365	O3'A	DA	B	18	6728	898	6535	-1500	3792	-47	O
ATOM	366	O3'B	DA	B	18	-2.264	8.673	-2.648	0.20	38.21		O
ANISOU	366	O3'B	DA	B	18	8218	1664	4633	-241	1874	-505	O
ATOM	367	C2'A	DA	B	18	-3.380	6.757	-3.824	0.80	21.78		C
ANISOU	367	C2'A	DA	B	18	3057	1706	3509	547	726	464	C
ATOM	368	C2'B	DA	B	18	-3.442	6.921	-3.850	0.20	22.17		C
ANISOU	368	C2'B	DA	B	18	3039	1976	3408	356	509	385	C
ATOM	369	C1'A	DA	B	18	-2.791	5.432	-3.386	0.80	19.05		C
ANISOU	369	C1'A	DA	B	18	2845	1314	3079	327	611	143	C
ATOM	370	C1'B	DA	B	18	-2.777	5.642	-3.392	0.20	20.34		C
ANISOU	370	C1'B	DA	B	18	2798	1855	3072	301	476	89	C
ATOM	371	N9 A	DA	B	18	-3.263	4.249	-4.083	0.80	18.34		N
ANISOU	371	N9 A	DA	B	18	2757	1442	2767	499	353	155	N
ATOM	372	N9 B	DA	B	18	-3.253	4.440	-4.058	0.20	18.85		N
ANISOU	372	N9 B	DA	B	18	2638	1717	2805	478	288	254	N
ATOM	373	C8 A	DA	B	18	-4.568	3.846	-4.177	0.80	18.68		C
ANISOU	373	C8 A	DA	B	18	2584	1807	2707	580	326	207	C
ATOM	374	C8 B	DA	B	18	-4.558	4.032	-4.156	0.20	18.56		C
ANISOU	374	C8 B	DA	B	18	2659	1711	2679	386	214	316	C
ATOM	375	N7 A	DA	B	18	-4.720	2.684	-4.763	0.80	17.95		N
ANISOU	375	N7 A	DA	B	18	2321	1846	2651	397	238	219	N
ATOM	376	N7 B	DA	B	18	-4.705	2.869	-4.739	0.20	17.94		N
ANISOU	376	N7 B	DA	B	18	2299	1825	2691	213	238	236	N
ATOM	377	C5 A	DA	B	18	-3.424	2.278	-5.043	0.80	16.94		C
ANISOU	377	C5 A	DA	B	18	2274	1492	2669	266	162	330	C
ATOM	378	C5 B	DA	B	18	-3.405	2.469	-5.014	0.20	17.59		C
ANISOU	378	C5 B	DA	B	18	2323	1606	2752	267	145	411	C
ATOM	379	C6 A	DA	B	18	-2.904	1.111	-5.634	0.80	16.44		C
ANISOU	379	C6 A	DA	B	18	2303	1465	2478	151	3	188	C
ATOM	380	C6 B	DA	B	18	-2.881	1.308	-5.605	0.20	17.73		C
ANISOU	380	C6 B	DA	B	18	2344	1721	2669	127	96	203	C
ATOM	381	N6 A	DA	B	18	-3.660	0.097	-6.066	0.80	17.27		N
ANISOU	381	N6 A	DA	B	18	2289	1677	2596	125	170	272	N
ATOM	382	N6 B	DA	B	18	-3.633	0.299	-6.050	0.20	18.91		N
ANISOU	382	N6 B	DA	B	18	2327	2094	2763	51	98	56	N
ATOM	383	N1 A	DA	B	18	-1.566	1.028	-5.784	0.80	16.64		N
ANISOU	383	N1 A	DA	B	18	2430	1348	2542	219	19	130	N
ATOM	384	N1 B	DA	B	18	-1.540	1.225	-5.745	0.20	17.76		N
ANISOU	384	N1 B	DA	B	18	2361	1671	2715	246	11	204	N
ATOM	385	C2 A	DA	B	18	-0.806	2.028	-5.322	0.80	16.47		C
ANISOU	385	C2 A	DA	B	18	2366	1270	2620	204	59	130	C
ATOM	386	C2 B	DA	B	18	-0.785	2.229	-5.283	0.20	17.56		C
ANISOU	386	C2 B	DA	B	18	2317	1585	2770	203	112	251	C
ATOM	387	N3 A	DA	B	18	-1.176	3.165	-4.733	0.80	17.37		N
ANISOU	387	N3 A	DA	B	18	2583	1322	2693	213	198	52	N
ATOM	388	N3 B	DA	B	18	-1.161	3.365	-4.698	0.20	18.59		N
ANISOU	388	N3 B	DA	B	18	2577	1624	2862	269	187	235	N
ATOM	389	C4 A	DA	B	18	-2.515	3.230	-4.623	0.80	17.61		C
ANISOU	389	C4 A	DA	B	18	2590	1355	2744	340	167	98	C
ATOM	390	C4 B	DA	B	18	-2.500	3.423	-4.593	0.20	18.03		C
ANISOU	390	C4 B	DA	B	18	2561	1487	2800	299	214	227	C
ATOM	391	P A	DT	B	19	-1.541	9.518	-3.335	0.80	37.49		P
ANISOU	391	P A	DT	B	19	6209	1119	6913	431	1620	37	P

ATOM	392	P	B	DT	B	19	-2.057	9.603	-3.908	0.20	30.04	P	
ANISOU	392	P	B	DT	B	19	4515	2253	4645	534	1472	-77	P
ATOM	393	OP1A		DT	B	19	-0.693	10.471	-2.562	0.80	66.07	O	
ANISOU	393	OP1A		DT	B	19	10410	3780	10911	-889	1278	-2410	O
ATOM	394	OP1B		DT	B	19	-1.464	10.851	-3.390	0.20	21.48	O	
ANISOU	394	OP1B		DT	B	19	3829	1516	2815	1641	594	689	O
ATOM	395	OP2A		DT	B	19	-2.712	9.937	-4.105	0.80	59.66	O	
ANISOU	395	OP2A		DT	B	19	7409	6697	8561	902	391	-805	O
ATOM	396	OP2B		DT	B	19	-3.326	9.671	-4.683	0.20	21.09	O	
ANISOU	396	OP2B		DT	B	19	4167	351	3495	-593	2162	-457	O
ATOM	397	O5'A		DT	B	19	-0.728	8.748	-4.443	0.80	25.76	O	
ANISOU	397	O5'A		DT	B	19	3778	2013	3996	379	1258	685	O
ATOM	398	O5'B		DT	B	19	-0.999	8.800	-4.783	0.20	21.19	O	
ANISOU	398	O5'B		DT	B	19	3355	1503	3191	602	460	187	O
ATOM	399	C5'A		DT	B	19	0.469	8.079	-4.106	0.80	22.11	C	
ANISOU	399	C5'A		DT	B	19	3527	1598	3275	162	579	-20	C
ATOM	400	C5'B		DT	B	19	0.142	8.166	-4.191	0.20	17.66	C	
ANISOU	400	C5'B		DT	B	19	2742	1410	2556	67	311	-115	C
ATOM	401	C4'A		DT	B	19	0.918	7.292	-5.307	0.80	19.18	C	
ANISOU	401	C4'A		DT	B	19	2697	1620	2970	-112	73	-50	C
ATOM	402	C4'B		DT	B	19	0.827	7.367	-5.271	0.20	15.03	C	
ANISOU	402	C4'B		DT	B	19	2073	1059	2578	31	90	-121	C
ATOM	403	O4'A		DT	B	19	-0.042	6.275	-5.637	0.80	19.64	O	
ANISOU	403	O4'A		DT	B	19	2907	1446	3108	-203	-128	81	O
ATOM	404	O4'B		DT	B	19	-0.025	6.262	-5.629	0.20	14.40	O	
ANISOU	404	O4'B		DT	B	19	2034	951	2486	8	60	36	O
ATOM	405	C3'A		DT	B	19	1.056	8.140	-6.570	0.80	18.48	C	
ANISOU	405	C3'A		DT	B	19	2584	1483	2953	-186	145	-67	C
ATOM	406	C3'B		DT	B	19	1.048	8.162	-6.561	0.20	17.12	C	
ANISOU	406	C3'B		DT	B	19	2395	1566	2543	-150	114	-29	C
ATOM	407	O3'A		DT	B	19	2.450	8.345	-6.734	0.80	20.95	O	
ANISOU	407	O3'A		DT	B	19	2720	2181	3056	-496	269	-283	O
ATOM	408	O3'B		DT	B	19	2.450	8.340	-6.735	0.20	19.41	O	
ANISOU	408	O3'B		DT	B	19	2484	1823	3067	-275	210	-256	O
ATOM	409	C2'A		DT	B	19	0.376	7.328	-7.666	0.80	18.81	C	
ANISOU	409	C2'A		DT	B	19	2700	1634	2810	-89	73	6	C
ATOM	410	C2'B		DT	B	19	0.355	7.348	-7.647	0.20	15.70	C	
ANISOU	410	C2'B		DT	B	19	2185	1361	2419	-2	176	38	C
ATOM	411	C1'A		DT	B	19	0.158	5.982	-7.009	0.80	18.13	C	
ANISOU	411	C1'A		DT	B	19	2535	1382	2969	43	-153	-59	C
ATOM	412	C1'B		DT	B	19	0.163	5.993	-7.006	0.20	14.89	C	
ANISOU	412	C1'B		DT	B	19	2003	1242	2409	103	109	-6	C
ATOM	413	N1	A	DT	B	19	-0.993	5.204	-7.463	0.80	17.76	N	
ANISOU	413	N1	A	DT	B	19	2520	1414	2813	96	-179	73	N
ATOM	414	N1	B	DT	B	19	-0.992	5.212	-7.460	0.20	14.11	N	
ANISOU	414	N1	B	DT	B	19	1885	1104	2370	266	116	18	N
ATOM	415	C2	A	DT	B	19	-0.758	3.973	-8.032	0.80	17.56	C	
ANISOU	415	C2	A	DT	B	19	2494	1295	2883	-21	-171	91	C
ATOM	416	C2	B	DT	B	19	-0.760	3.979	-8.023	0.20	13.80	C	
ANISOU	416	C2	B	DT	B	19	1744	1080	2416	328	199	60	C
ATOM	417	O2	A	DT	B	19	0.357	3.542	-8.258	0.80	18.19	O	
ANISOU	417	O2	A	DT	B	19	2525	1394	2993	106	-87	-135	O
ATOM	418	O2	B	DT	B	19	0.358	3.548	-8.245	0.20	14.13	O	
ANISOU	418	O2	B	DT	B	19	1673	1136	2560	257	232	129	O
ATOM	419	N3	A	DT	B	19	-1.886	3.273	-8.358	0.80	17.05	N	
ANISOU	419	N3	A	DT	B	19	2429	1269	2780	-9	-233	19	N

ATOM	420	N3	B	DT	B	19	-1.892	3.281	-8.357	0.20	14.08		N
ANISOU	420	N3	B	DT	B	19	1773	1223	2353	208	185	206	N
ATOM	421	C4	A	DT	B	19	-3.195	3.664	-8.174	0.80	17.88		C
ANISOU	421	C4	A	DT	B	19	2415	1437	2941	93	-253	166	C
ATOM	422	C4	B	DT	B	19	-3.202	3.678	-8.173	0.20	13.56		C
ANISOU	422	C4	B	DT	B	19	1745	1129	2275	161	148	209	C
ATOM	423	O4	A	DT	B	19	-4.103	2.908	-8.501	0.80	18.22		O
ANISOU	423	O4	A	DT	B	19	2218	1578	3124	101	-246	35	O
ATOM	424	O4	B	DT	B	19	-4.117	2.932	-8.510	0.20	14.05		O
ANISOU	424	O4	B	DT	B	19	1911	1091	2334	50	207	137	O
ATOM	425	C5	A	DT	B	19	-3.373	4.983	-7.599	0.80	18.56		C
ANISOU	425	C5	A	DT	B	19	2458	1561	3032	178	-49	37	C
ATOM	426	C5	B	DT	B	19	-3.373	4.987	-7.580	0.20	14.19		C
ANISOU	426	C5	B	DT	B	19	1827	1234	2327	350	98	112	C
ATOM	427	C7	A	DT	B	19	-4.760	5.496	-7.370	0.80	18.90		C
ANISOU	427	C7	A	DT	B	19	2532	1734	2914	287	-61	-138	C
ATOM	428	C7	B	DT	B	19	-4.758	5.503	-7.346	0.20	14.48		C
ANISOU	428	C7	B	DT	B	19	1789	1526	2186	307	91	-105	C
ATOM	429	C6	A	DT	B	19	-2.275	5.670	-7.260	0.80	18.38		C
ANISOU	429	C6	A	DT	B	19	2634	1425	2924	234	-76	55	C
ATOM	430	C6	B	DT	B	19	-2.273	5.673	-7.247	0.20	14.57		C
ANISOU	430	C6	B	DT	B	19	1882	1274	2377	338	73	154	C
ATOM	431	P		DT	B	20	3.036	9.005	-8.063	1.00	23.35		P
ANISOU	431	P		DT	B	20	3632	1527	3712	-1010	506	-547	P
ATOM	432	OP1		DT	B	20	4.325	9.581	-7.694	1.00	30.87		O
ANISOU	432	OP1		DT	B	20	4382	3080	4264	-787	510	-330	O
ATOM	433	OP2		DT	B	20	2.084	9.773	-8.765	1.00	25.54		O
ANISOU	433	OP2		DT	B	20	4327	1244	4133	49	409	269	O
ATOM	434	O5'		DT	B	20	3.318	7.740	-8.990	1.00	19.35		O
ANISOU	434	O5'		DT	B	20	2986	1006	3359	-369	206	-141	O
ATOM	435	C5'		DT	B	20	4.091	6.648	-8.524	1.00	20.44		C
ANISOU	435	C5'		DT	B	20	2626	2094	3044	-341	37	-59	C
ATOM	436	C4'		DT	B	20	4.193	5.564	-9.574	1.00	18.85		C
ANISOU	436	C4'		DT	B	20	2653	1342	3164	-356	-31	-122	C
ATOM	437	O4'		DT	B	20	2.904	4.964	-9.816	1.00	18.38		O
ANISOU	437	O4'		DT	B	20	2628	1107	3248	-44	43	152	O
ATOM	438	C3'		DT	B	20	4.721	6.022	-10.937	1.00	20.07		C
ANISOU	438	C3'		DT	B	20	2601	1725	3298	-164	99	-202	C
ATOM	439	O3'		DT	B	20	5.718	5.067	-11.262	1.00	22.30		O
ANISOU	439	O3'		DT	B	20	2626	2342	3502	13	-152	-245	O
ATOM	440	C2'		DT	B	20	3.495	5.960	-11.834	1.00	17.95		C
ANISOU	440	C2'		DT	B	20	2782	801	3236	-11	127	36	C
ATOM	441	C1'		DT	B	20	2.687	4.834	-11.202	1.00	17.77		C
ANISOU	441	C1'		DT	B	20	2752	794	3205	-93	-25	88	C
ATOM	442	N1		DT	B	20	1.232	4.831	-11.406	1.00	16.82		N
ANISOU	442	N1		DT	B	20	2693	467	3229	66	105	132	N
ATOM	443	C2		DT	B	20	0.606	3.663	-11.790	1.00	16.13		C
ANISOU	443	C2		DT	B	20	2608	531	2986	-151	-18	120	C
ATOM	444	O2		DT	B	20	1.211	2.658	-12.120	1.00	17.23		O
ANISOU	444	O2		DT	B	20	2600	586	3359	205	-47	58	O
ATOM	445	N3		DT	B	20	-0.764	3.735	-11.823	1.00	16.14		N
ANISOU	445	N3		DT	B	20	2550	496	3084	-60	-71	103	N
ATOM	446	C4		DT	B	20	-1.557	4.817	-11.474	1.00	16.52		C
ANISOU	446	C4		DT	B	20	2600	548	3129	298	-76	166	C
ATOM	447	O4		DT	B	20	-2.770	4.714	-11.503	1.00	16.72		O
ANISOU	447	O4		DT	B	20	2573	646	3132	243	-113	61	O

ATOM	448	C5	DT	B	20	-0.831	5.998	-11.052	1.00	16.26		C
ANISOU	448	C5	DT	B	20	2713	262	3200	67	-124	144	C
ATOM	449	C7	DT	B	20	-1.597	7.230	-10.678	1.00	17.35		C
ANISOU	449	C7	DT	B	20	2748	663	3179	60	-54	0	C
ATOM	450	C6	DT	B	20	0.505	5.939	-11.029	1.00	16.49		C
ANISOU	450	C6	DT	B	20	2647	355	3263	110	-117	168	C
HETATM	451	P	5HC	B	21	6.694	5.225	-12.477	1.00	24.00		P
ANISOU	451	P	5HC	B	21	2655	2626	3838	-398	66	-527	P
HETATM	452	OP1	5HC	B	21	7.842	4.371	-12.214	1.00	27.67		O
ANISOU	452	OP1	5HC	B	21	2702	3594	4215	-121	-69	-954	O
HETATM	453	OP2	5HC	B	21	6.834	6.610	-12.784	1.00	27.44		O
ANISOU	453	OP2	5HC	B	21	3388	3242	3795	-1060	507	38	O
HETATM	454	O5'	5HC	B	21	5.884	4.644	-13.707	1.00	20.89		O
ANISOU	454	O5'	5HC	B	21	2638	1852	3447	-201	121	-290	O
HETATM	455	C5'	5HC	B	21	5.620	3.252	-13.748	1.00	20.57		C
ANISOU	455	C5'	5HC	B	21	2560	1628	3627	21	107	-257	C
HETATM	456	C4'	5HC	B	21	4.578	2.959	-14.798	1.00	21.24		C
ANISOU	456	C4'	5HC	B	21	2510	1946	3614	175	6	-280	C
HETATM	457	O4'	5HC	B	21	3.317	3.532	-14.439	1.00	19.62		O
ANISOU	457	O4'	5HC	B	21	2431	1633	3390	91	0	28	O
HETATM	458	C3'	5HC	B	21	4.875	3.524	-16.176	1.00	21.71		C
ANISOU	458	C3'	5HC	B	21	2647	1874	3725	211	35	-435	C
HETATM	459	O3'	5HC	B	21	5.438	2.419	-16.864	1.00	25.37		O
ANISOU	459	O3'	5HC	B	21	2658	3105	3873	754	-337	-623	O
HETATM	460	C2'	5HC	B	21	3.502	3.901	-16.723	1.00	21.90		C
ANISOU	460	C2'	5HC	B	21	2669	2326	3325	389	-88	-374	C
HETATM	461	C1'	5HC	B	21	2.548	3.507	-15.618	1.00	20.59		C
ANISOU	461	C1'	5HC	B	21	2455	1877	3490	234	-191	-117	C
HETATM	462	N1	5HC	B	21	1.381	4.370	-15.400	1.00	18.70		N
ANISOU	462	N1	5HC	B	21	2480	1304	3321	295	-93	52	N
HETATM	463	C2	5HC	B	21	0.108	3.820	-15.537	1.00	18.13		C
ANISOU	463	C2	5HC	B	21	2495	1254	3137	3	-77	-234	C
HETATM	464	O2	5HC	B	21	-0.001	2.656	-15.940	1.00	18.29		O
ANISOU	464	O2	5HC	B	21	2452	1078	3420	69	-79	-187	O
HETATM	465	N3	5HC	B	21	-0.969	4.572	-15.238	1.00	16.88		N
ANISOU	465	N3	5HC	B	21	2322	1020	3070	-13	-47	109	N
HETATM	466	C4	5HC	B	21	-0.811	5.825	-14.815	1.00	16.90		C
ANISOU	466	C4	5HC	B	21	2513	921	2987	78	-8	58	C
HETATM	467	N4	5HC	B	21	-1.903	6.521	-14.514	1.00	17.96		N
ANISOU	467	N4	5HC	B	21	2508	1105	3210	17	-38	95	N
HETATM	468	C5	5HC	B	21	0.477	6.421	-14.689	1.00	17.49		C
ANISOU	468	C5	5HC	B	21	2560	805	3280	-84	-23	360	C
HETATM	469	C5M	5HC	B	21	0.690	7.876	-14.210	1.00	18.79		C
ANISOU	469	C5M	5HC	B	21	2462	1763	2912	-81	-28	148	C
HETATM	470	O5	5HC	B	21	0.201	8.787	-15.220	1.00	20.00		O
ANISOU	470	O5	5HC	B	21	2760	1691	3148	121	-80	174	O
HETATM	471	C6	5HC	B	21	1.535	5.658	-14.968	1.00	17.74		C
ANISOU	471	C6	5HC	B	21	2518	1004	3219	245	-10	61	C
ATOM	472	P	DG	B	22	5.957	2.505	-18.307	1.00	29.70		P
ANISOU	472	P	DG	B	22	2495	4802	3987	179	-24	-916	P
ATOM	473	OP1	DG	B	22	6.885	1.386	-18.491	1.00	33.39		O
ANISOU	473	OP1	DG	B	22	2666	5472	4546	950	-410	-1860	O
ATOM	474	OP2	DG	B	22	6.319	3.881	-18.636	1.00	29.57		O
ANISOU	474	OP2	DG	B	22	2597	5036	3601	-368	417	-212	O
ATOM	475	O5'	DG	B	22	4.684	2.285	-19.230	1.00	27.62		O
ANISOU	475	O5'	DG	B	22	2380	4235	3879	238	-180	-964	O

ATOM	476	C5'	DG	B	22	4.041	1.033	-19.269	1.00	25.38		C
ANISOU	476	C5'	DG	B	22	2478	3560	3604	547	-109	-728	C
ATOM	477	C4'	DG	B	22	2.800	1.148	-20.118	1.00	21.58		C
ANISOU	477	C4'	DG	B	22	2557	2108	3534	415	-318	-873	C
ATOM	478	O4'	DG	B	22	1.813	1.979	-19.462	1.00	19.97		O
ANISOU	478	O4'	DG	B	22	2448	1682	3458	449	-123	-254	O
ATOM	479	C3'	DG	B	22	3.025	1.782	-21.488	1.00	20.21		C
ANISOU	479	C3'	DG	B	22	2191	1971	3515	214	-50	-683	C
ATOM	480	O3'	DG	B	22	2.262	1.031	-22.433	1.00	20.61		O
ANISOU	480	O3'	DG	B	22	2269	1932	3627	133	-105	-442	O
ATOM	481	C2'	DG	B	22	2.396	3.156	-21.352	1.00	19.75		C
ANISOU	481	C2'	DG	B	22	2472	1664	3366	-7	-58	-382	C
ATOM	482	C1'	DG	B	22	1.252	2.830	-20.434	1.00	18.52		C
ANISOU	482	C1'	DG	B	22	2424	1213	3400	325	-28	-185	C
ATOM	483	N9	DG	B	22	0.613	3.950	-19.753	1.00	17.92		N
ANISOU	483	N9	DG	B	22	2318	1303	3187	186	-168	-128	N
ATOM	484	C8	DG	B	22	1.194	5.119	-19.330	1.00	20.22		C
ANISOU	484	C8	DG	B	22	2494	1881	3308	127	-124	-372	C
ATOM	485	N7	DG	B	22	0.358	5.917	-18.732	1.00	18.58		N
ANISOU	485	N7	DG	B	22	2286	1570	3202	186	-63	-149	N
ATOM	486	C5	DG	B	22	-0.851	5.237	-18.766	1.00	16.85		C
ANISOU	486	C5	DG	B	22	2461	764	3175	-59	-124	-61	C
ATOM	487	C6	DG	B	22	-2.133	5.615	-18.294	1.00	17.29		C
ANISOU	487	C6	DG	B	22	2341	1160	3065	-16	-116	61	C
ATOM	488	O6	DG	B	22	-2.456	6.638	-17.702	1.00	17.63		O
ANISOU	488	O6	DG	B	22	2471	957	3267	212	-83	-42	O
ATOM	489	N1	DG	B	22	-3.083	4.639	-18.551	1.00	16.41		N
ANISOU	489	N1	DG	B	22	2246	966	3020	211	-17	-41	N
ATOM	490	C2	DG	B	22	-2.835	3.452	-19.185	1.00	16.45		C
ANISOU	490	C2	DG	B	22	2246	992	3009	204	-88	171	C
ATOM	491	N2	DG	B	22	-3.881	2.646	-19.351	1.00	16.72		N
ANISOU	491	N2	DG	B	22	2306	1027	3016	167	-56	126	N
ATOM	492	N3	DG	B	22	-1.646	3.086	-19.631	1.00	17.04		N
ANISOU	492	N3	DG	B	22	2258	1154	3062	90	-47	89	N
ATOM	493	C4	DG	B	22	-0.709	4.023	-19.396	1.00	17.26		C
ANISOU	493	C4	DG	B	22	2328	1080	3149	-15	-168	48	C
ATOM	494	P	DC	B	23	2.756	0.808	-23.886	1.00	19.80		P
ANISOU	494	P	DC	B	23	1990	2396	3136	52	48	-425	P
ATOM	495	OP1	DC	B	23	3.873	-0.136	-23.892	1.00	23.05		O
ANISOU	495	OP1	DC	B	23	2205	2822	3730	302	28	-727	O
ATOM	496	OP2	DC	B	23	2.907	2.108	-24.494	1.00	21.38		O
ANISOU	496	OP2	DC	B	23	2474	2333	3317	-312	90	-202	O
ATOM	497	O5'	DC	B	23	1.485	0.108	-24.533	1.00	19.03		O
ANISOU	497	O5'	DC	B	23	2148	2068	3012	83	-46	-327	O
ATOM	498	C5'	DC	B	23	0.972	-1.125	-24.013	1.00	19.03		C
ANISOU	498	C5'	DC	B	23	2107	1944	3178	140	-176	-298	C
ATOM	499	C4'	DC	B	23	-0.520	-1.031	-23.792	1.00	18.25		C
ANISOU	499	C4'	DC	B	23	2121	1919	2892	203	-92	-92	C
ATOM	500	O4'	DC	B	23	-0.820	-0.072	-22.761	1.00	18.05		O
ANISOU	500	O4'	DC	B	23	2136	1869	2851	212	-37	-139	O
ATOM	501	C3'	DC	B	23	-1.349	-0.568	-24.982	1.00	18.05		C
ANISOU	501	C3'	DC	B	23	2269	1705	2882	107	-179	-203	C
ATOM	502	O3'	DC	B	23	-1.636	-1.658	-25.842	1.00	19.30		O
ANISOU	502	O3'	DC	B	23	2450	1634	3248	254	-200	-323	O
ATOM	503	C2'	DC	B	23	-2.620	-0.106	-24.307	1.00	17.85		C
ANISOU	503	C2'	DC	B	23	2150	1754	2876	100	-106	-92	C

ATOM	504	C1'	DC	B	23	-2.087	0.529	-23.050	1.00	17.71		C
ANISOU	504	C1'	DC	B	23	2004	1751	2973	132	-84	-50	C
ATOM	505	N1	DC	B	23	-1.915	1.990	-23.046	1.00	17.09		N
ANISOU	505	N1	DC	B	23	2003	1687	2801	82	-101	-86	N
ATOM	506	C2	DC	B	23	-2.998	2.782	-22.663	1.00	16.37		C
ANISOU	506	C2	DC	B	23	2005	1694	2520	124	-95	16	C
ATOM	507	O2	DC	B	23	-4.094	2.246	-22.477	1.00	17.16		O
ANISOU	507	O2	DC	B	23	1978	1718	2824	172	-73	-68	O
ATOM	508	N3	DC	B	23	-2.815	4.112	-22.494	1.00	17.01		N
ANISOU	508	N3	DC	B	23	2025	1739	2698	59	-102	-23	N
ATOM	509	C4	DC	B	23	-1.621	4.656	-22.735	1.00	17.14		C
ANISOU	509	C4	DC	B	23	2004	1755	2752	53	3	-79	C
ATOM	510	N4	DC	B	23	-1.475	5.963	-22.518	1.00	17.90		N
ANISOU	510	N4	DC	B	23	2006	1811	2983	74	-20	-144	N
ATOM	511	C5	DC	B	23	-0.515	3.877	-23.176	1.00	17.98		C
ANISOU	511	C5	DC	B	23	2070	1873	2888	45	80	-238	C
ATOM	512	C6	DC	B	23	-0.700	2.561	-23.303	1.00	17.79		C
ANISOU	512	C6	DC	B	23	1996	1873	2888	64	0	-93	C
ATOM	513	P	DG	B	24	-2.027	-1.440	-27.343	1.00	19.52		P
ANISOU	513	P	DG	B	24	2302	1945	3169	198	-184	-386	P
ATOM	514	OP1	DG	B	24	-1.938	-2.767	-27.981	1.00	22.14		O
ANISOU	514	OP1	DG	B	24	2561	2042	3808	445	-462	-698	O
ATOM	515	OP2	DG	B	24	-1.251	-0.302	-27.899	1.00	20.89		O
ANISOU	515	OP2	DG	B	24	2455	2431	3050	53	-3	-418	O
ATOM	516	O5'	DG	B	24	-3.539	-0.949	-27.293	1.00	18.90		O
ANISOU	516	O5'	DG	B	24	2301	1915	2961	82	-157	-173	O
ATOM	517	C5'	DG	B	24	-4.611	-1.815	-26.891	1.00	17.95		C
ANISOU	517	C5'	DG	B	24	2247	1630	2942	53	-263	-103	C
ATOM	518	C4'	DG	B	24	-5.870	-1.000	-26.711	1.00	17.69		C
ANISOU	518	C4'	DG	B	24	2377	1411	2932	-2	-283	-91	C
ATOM	519	O4'	DG	B	24	-5.665	-0.029	-25.664	1.00	17.05		O
ANISOU	519	O4'	DG	B	24	2224	1476	2777	-22	-143	-86	O
ATOM	520	C3'	DG	B	24	-6.310	-0.178	-27.917	1.00	17.37		C
ANISOU	520	C3'	DG	B	24	2233	1538	2827	-46	-220	-155	C
ATOM	521	O3'	DG	B	24	-7.058	-1.008	-28.808	1.00	18.53		O
ANISOU	521	O3'	DG	B	24	2439	1531	3068	-43	-312	-224	O
ATOM	522	C2'	DG	B	24	-7.108	0.944	-27.279	1.00	17.50		C
ANISOU	522	C2'	DG	B	24	2248	1517	2883	-36	-100	-54	C
ATOM	523	C1'	DG	B	24	-6.357	1.186	-25.989	1.00	17.02		C
ANISOU	523	C1'	DG	B	24	2250	1418	2796	-106	-56	14	C
ATOM	524	N9	DG	B	24	-5.359	2.251	-26.043	1.00	16.84		N
ANISOU	524	N9	DG	B	24	2168	1485	2746	-98	-147	-72	N
ATOM	525	C8	DG	B	24	-4.063	2.145	-26.488	1.00	17.48		C
ANISOU	525	C8	DG	B	24	2292	1665	2683	-142	-62	-34	C
ATOM	526	N7	DG	B	24	-3.370	3.239	-26.316	1.00	17.33		N
ANISOU	526	N7	DG	B	24	2205	1656	2721	-102	-52	-82	N
ATOM	527	C5	DG	B	24	-4.269	4.128	-25.742	1.00	16.61		C
ANISOU	527	C5	DG	B	24	1996	1670	2642	-48	-140	-1	C
ATOM	528	C6	DG	B	24	-4.088	5.467	-25.307	1.00	16.94		C
ANISOU	528	C6	DG	B	24	2173	1637	2625	-256	-124	36	C
ATOM	529	O6	DG	B	24	-3.066	6.163	-25.358	1.00	18.23		O
ANISOU	529	O6	DG	B	24	2273	1692	2960	-330	-79	-51	O
ATOM	530	N1	DG	B	24	-5.262	5.999	-24.782	1.00	17.20		N
ANISOU	530	N1	DG	B	24	2265	1581	2689	-266	27	-72	N
ATOM	531	C2	DG	B	24	-6.458	5.334	-24.694	1.00	16.88		C
ANISOU	531	C2	DG	B	24	2297	1476	2639	-204	31	-108	C

ATOM	532	N2	DG	B	24	-7.485	6.014	-24.173	1.00	17.90		N	
ANISOU	532	N2	DG	B	24	2389	1516	2894	-171	175	-165	N	
ATOM	533	N3	DG	B	24	-6.632	4.076	-25.066	1.00	16.72		N	
ANISOU	533	N3	DG	B	24	2135	1535	2681	-170	-42	-143	N	
ATOM	534	C4	DG	B	24	-5.501	3.535	-25.570	1.00	16.50		C	
ANISOU	534	C4	DG	B	24	2214	1528	2527	-210	-93	-59	C	
TER	535		DG	B	24								
HETATM	536	MG		MG	A	101	-2.090	10.717	-18.618	1.00	19.21	MG	
ANISOU	536	MG		MG	A	101	2532	2214	2553	82	-100	-7	MG
HETATM	537	N1	SPK	B	101	3.058	13.484	-12.303	1.00	38.88		N	
ANISOU	537	N1	SPK	B	101	4846	4645	5280	-362	-350	-636	N	
HETATM	538	C2	SPK	B	101	2.338	13.371	-11.029	1.00	35.81		C	
ANISOU	538	C2	SPK	B	101	4971	4257	4376	-16	-185	-373	C	
HETATM	539	C3	SPK	B	101	3.293	12.989	-9.913	1.00	30.62		C	
ANISOU	539	C3	SPK	B	101	4522	3285	3826	364	193	-399	C	
HETATM	540	C4	SPK	B	101	2.535	12.987	-8.612	1.00	33.05		C	
ANISOU	540	C4	SPK	B	101	4292	3869	4397	-391	-165	331	C	
HETATM	541	N5	SPK	B	101	3.342	12.640	-7.440	1.00	32.00		N	
ANISOU	541	N5	SPK	B	101	4490	3570	4098	-253	238	134	N	
HETATM	542	C6	SPK	B	101	2.473	12.579	-6.256	1.00	37.35		C	
ANISOU	542	C6	SPK	B	101	5311	5164	3712	233	381	414	C	
HETATM	543	C7	SPK	B	101	3.267	12.078	-5.063	1.00	32.55		C	
ANISOU	543	C7	SPK	B	101	4172	4341	3852	6	33	-2	C	
HETATM	544	C8	SPK	B	101	2.320	11.846	-3.906	1.00	33.83		C	
ANISOU	544	C8	SPK	B	101	4600	4791	3461	-732	35	-96	C	
HETATM	545	C9	SPK	B	101	3.011	11.338	-2.676	1.00	37.83		C	
ANISOU	545	C9	SPK	B	101	4599	5260	4513	-792	939	1191	C	
HETATM	546	N10	SPK	B	101	2.252	11.856	-1.535	1.00	38.86		N	
ANISOU	546	N10	SPK	B	101	5459	3604	5703	-727	755	383	N	
HETATM	547	C11	SPK	B	101	2.948	11.432	-0.327	1.00	70.65		C	
ANISOU	547	C11	SPK	B	101	14443	11216	14481	-2296	-5727	2814	C	
HETATM	548	N1	ASPK	B	102	3.224	-6.111	4.543	0.50	30.43		N	
ANISOU	548	N1	ASPK	B	102	3750	3385	4425	211	-11	-18	N	
HETATM	549	N1	BSPK	B	102	3.498	-4.976	2.823	0.30	33.20		N	
ANISOU	549	N1	BSPK	B	102	4193	3925	4495	227	-95	60	N	
HETATM	550	C2	ASPK	B	102	3.134	-4.633	4.562	0.50	27.48		C	
ANISOU	550	C2	ASPK	B	102	3553	3273	3613	141	79	-3	C	
HETATM	551	C2	BSPK	B	102	3.170	-4.714	4.292	0.30	29.88		C	
ANISOU	551	C2	BSPK	B	102	3538	3557	4256	627	-479	386	C	
HETATM	552	C3	ASPK	B	102	4.449	-4.054	5.015	0.50	32.21		C	
ANISOU	552	C3	ASPK	B	102	3951	4325	3961	-149	-196	-275	C	
HETATM	553	C3	BSPK	B	102	4.374	-4.182	5.043	0.30	33.74		C	
ANISOU	553	C3	BSPK	B	102	4024	4514	4280	55	-283	-179	C	
HETATM	554	C4	ASPK	B	102	4.318	-2.673	5.567	0.50	29.32		C	
ANISOU	554	C4	ASPK	B	102	3401	3734	4003	-148	123	320	C	
HETATM	555	C4	BSPK	B	102	4.280	-2.739	5.455	0.30	31.94		C	
ANISOU	555	C4	BSPK	B	102	3467	4429	4240	167	-29	3	C	
HETATM	556	N5	ASPK	B	102	5.570	-1.933	5.760	0.50	31.92		N	
ANISOU	556	N5	ASPK	B	102	3612	4300	4216	-311	-138	-33	N	
HETATM	557	N5	BSPK	B	102	5.530	-2.012	5.745	0.30	31.37		N	
ANISOU	557	N5	BSPK	B	102	3674	4276	3968	69	39	-51	N	
HETATM	558	C6	ASPK	B	102	6.567	-2.158	4.741	0.50	32.61		C	
ANISOU	558	C6	ASPK	B	102	3649	4545	4196	643	-457	171	C	
HETATM	559	C6	BSPK	B	102	6.553	-2.175	4.736	0.30	25.06		C	
ANISOU	559	C6	BSPK	B	102	2603	3407	3511	254	-586	485	C	
HETATM	560	C7	ASPK	B	102	7.575	-1.044	4.737	0.50	36.82		C	

ANISOU	560	C7	ASPK	B	102	4652	4911	4424	138	-342	515	C
HETATM	561	C7	BSPK	B	102	7.544	-1.046	4.752	0.30	29.77		C
ANISOU	561	C7	BSPK	B	102	3304	4107	3898	-332	-215	360	C
HETATM	562	C8	ASPK	B	102	8.835	-1.685	4.254	0.50	38.26		C
ANISOU	562	C8	ASPK	B	102	4654	5107	4775	-127	-146	186	C
HETATM	563	C8	BSPK	B	102	8.820	-1.673	4.286	0.30	33.44		C
ANISOU	563	C8	BSPK	B	102	3414	4944	4348	147	-380	94	C
HETATM	564	C9	ASPK	B	102	9.757	-0.809	3.453	0.50	37.48		C
ANISOU	564	C9	ASPK	B	102	4887	4384	4966	-107	-354	144	C
HETATM	565	C9	BSPK	B	102	9.731	-0.809	3.459	0.30	38.30		C
ANISOU	565	C9	BSPK	B	102	4718	4826	5005	-13	-52	353	C
HETATM	566	N10	ASPK	B	102	10.279	0.219	4.320	0.50	33.65		N
ANISOU	566	N10	ASPK	B	102	3823	4509	4450	-848	-242	449	N
HETATM	567	N10	BSPK	B	102	10.276	0.226	4.304	0.30	38.11		N
ANISOU	567	N10	BSPK	B	102	4656	5177	4646	155	-116	339	N
HETATM	568	C11	ASPK	B	102	9.899	1.569	3.898	0.50	38.40		C
ANISOU	568	C11	ASPK	B	102	5004	4488	5096	-395	-44	343	C
HETATM	569	C11	BSPK	B	102	9.883	1.567	3.867	0.30	38.57		C
ANISOU	569	C11	BSPK	B	102	4897	4860	4897	-260	322	395	C
HETATM	570	C12	ASPK	B	102	10.427	1.801	2.514	0.50	37.63		C
ANISOU	570	C12	ASPK	B	102	4482	4950	4866	-177	32	51	C
HETATM	571	C12	BSPK	B	102	10.415	1.785	2.482	0.30	36.34		C
ANISOU	571	C12	BSPK	B	102	4199	4787	4822	-131	255	89	C
HETATM	572	C13	ASPK	B	102	10.114	3.126	1.874	0.50	39.84		C
ANISOU	572	C13	ASPK	B	102	5072	4906	5157	-194	93	137	C
HETATM	573	C13	BSPK	B	102	9.903	2.979	1.718	0.30	38.21		C
ANISOU	573	C13	BSPK	B	102	4740	4714	5064	-137	192	45	C
HETATM	574	N14	ASPK	B	102	10.183	2.948	0.411	0.50	38.49		N
ANISOU	574	N14	ASPK	B	102	4750	4733	5140	-373	99	89	N
HETATM	575	N14	BSPK	B	102	11.050	3.887	1.584	0.30	36.30		N
ANISOU	575	N14	BSPK	B	102	4259	4545	4988	236	510	-56	N
HETATM	576	O	HOH	A	201	5.846	-7.853	-14.025	1.00	21.22		O
ANISOU	576	O	HOH	A	201	3182	1741	3137	-44	332	-111	O
HETATM	577	O	HOH	A	202	2.577	-4.929	-0.769	1.00	26.57		O
ANISOU	577	O	HOH	A	202	4030	2357	3708	887	-159	-105	O
HETATM	578	O	HOH	A	203	2.952	2.709	-7.560	1.00	18.18		O
ANISOU	578	O	HOH	A	203	2248	1663	2997	45	10	86	O
HETATM	579	O	HOH	A	204	-1.508	3.996	-0.035	1.00	18.25		O
ANISOU	579	O	HOH	A	204	2450	1511	2970	-27	246	21	O
HETATM	580	O	HOH	A	205	-10.798	2.045	-13.263	1.00	31.40		O
ANISOU	580	O	HOH	A	205	3285	4050	4592	-510	933	-673	O
HETATM	581	O	HOH	A	206	-2.267	2.679	4.092	1.00	20.23		O
ANISOU	581	O	HOH	A	206	2738	1943	3005	172	27	-121	O
HETATM	582	O	HOH	A	207	-6.922	0.503	-10.931	1.00	27.30		O
ANISOU	582	O	HOH	A	207	3088	3739	3546	162	39	-501	O
HETATM	583	O	HOH	A	208	-1.957	9.193	-19.974	1.00	19.11		O
ANISOU	583	O	HOH	A	208	2565	1570	3125	-17	-190	101	O
HETATM	584	O	HOH	A	209	2.769	-6.156	-8.572	1.00	22.56		O
ANISOU	584	O	HOH	A	209	2926	1821	3823	672	445	512	O
HETATM	585	O	HOH	A	210	4.099	0.826	7.490	1.00	23.58		O
ANISOU	585	O	HOH	A	210	2554	2720	3683	-272	-259	47	O
HETATM	586	O	HOH	A	211	6.435	0.997	1.218	1.00	36.42		O
ANISOU	586	O	HOH	A	211	4374	3612	5850	940	-721	-261	O
HETATM	587	O	HOH	A	212	-4.084	-0.970	-17.389	1.00	21.97		O
ANISOU	587	O	HOH	A	212	3127	1964	3255	174	-16	-15	O
HETATM	588	O	HOH	A	213	-0.767	-5.435	-4.004	1.00	30.33		O

ANISOU	588	O	HOH A 213	5424	1742	4357	-406	844	-244	O
HETATM	589	O	HOH A 214	-1.733	-6.733	-11.114	1.00	24.67		O
ANISOU	589	O	HOH A 214	3046	1672	4655	140	-115	-151	O
HETATM	590	O	HOH A 215	-2.796	4.676	2.374	1.00	22.82		O
ANISOU	590	O	HOH A 215	3053	1953	3664	310	366	-9	O
HETATM	591	O	HOH A 216	-8.374	5.451	-12.923	1.00	25.51		O
ANISOU	591	O	HOH A 216	3009	2705	3979	503	21	-505	O
HETATM	592	O	HOH A 217	-0.165	-1.774	-15.968	1.00	31.40		O
ANISOU	592	O	HOH A 217	5557	2131	4241	482	996	8	O
HETATM	593	O	HOH A 218	-1.321	8.773	4.715	1.00	35.18		O
ANISOU	593	O	HOH A 218	5814	2277	5276	-569	-874	216	O
HETATM	594	O	HOH A 219	-3.958	-4.276	-6.741	1.00	28.51		O
ANISOU	594	O	HOH A 219	3443	2564	4822	-628	870	-729	O
HETATM	595	O	HOH A 220	-4.851	-3.816	-9.974	1.00	36.03		O
ANISOU	595	O	HOH A 220	2850	3864	6973	-556	84	1053	O
HETATM	596	O	HOH A 221	-15.519	4.872	-20.124	1.00	36.79		O
ANISOU	596	O	HOH A 221	2585	5781	5611	380	-358	254	O
HETATM	597	O	HOH A 222	-0.280	-5.917	-18.463	1.00	30.59		O
ANISOU	597	O	HOH A 222	5292	2197	4135	418	-184	23	O
HETATM	598	O	HOH A 223	-8.216	11.712	-17.811	1.00	26.78		O
ANISOU	598	O	HOH A 223	3520	2790	3863	-502	712	-212	O
HETATM	599	O	HOH A 224	-6.103	-1.516	-8.798	1.00	28.55		O
ANISOU	599	O	HOH A 224	3426	3361	4060	-613	371	-337	O
HETATM	600	O	HOH A 225	-4.251	3.582	12.794	1.00	27.78		O
ANISOU	600	O	HOH A 225	3656	2479	4418	399	315	706	O
HETATM	601	O	HOH A 226	-5.654	11.954	-25.879	1.00	24.74		O
ANISOU	601	O	HOH A 226	3458	2284	3657	-788	-10	210	O
HETATM	602	O	HOH A 227	6.060	-4.783	-5.268	1.00	36.77		O
ANISOU	602	O	HOH A 227	4834	3463	5673	1354	892	588	O
HETATM	603	O	HOH A 228	-5.742	9.863	-15.940	1.00	32.89		O
ANISOU	603	O	HOH A 228	5975	2598	3923	863	-154	-265	O
HETATM	604	O	HOH A 229	-0.067	2.401	11.646	1.00	35.80		O
ANISOU	604	O	HOH A 229	4458	4340	4803	-1673	215	-1165	O
HETATM	605	O	HOH A 230	6.061	-2.940	-3.236	1.00	34.36		O
ANISOU	605	O	HOH A 230	4231	3400	5424	1650	-789	-536	O
HETATM	606	O	HOH A 231	5.432	-0.922	2.438	1.00	25.28		O
ANISOU	606	O	HOH A 231	2882	3171	3552	587	-381	-97	O
HETATM	607	O	HOH A 232	-5.381	7.425	-13.438	1.00	28.30		O
ANISOU	607	O	HOH A 232	4590	2609	3554	1349	-203	-148	O
HETATM	608	O	HOH A 233	-8.708	10.080	-15.776	1.00	43.06		O
ANISOU	608	O	HOH A 233	4784	5110	6465	1451	-1040	-1009	O
HETATM	609	O	HOH A 234	-4.313	14.386	-26.084	1.00	41.33		O
ANISOU	609	O	HOH A 234	7568	4570	3564	-4477	1126	-539	O
HETATM	610	O	HOH A 235	-0.954	-3.255	-18.162	1.00	32.72		O
ANISOU	610	O	HOH A 235	5048	3045	4339	118	-588	-609	O
HETATM	611	O	HOH A 236	-0.305	9.442	-22.178	1.00	26.37		O
ANISOU	611	O	HOH A 236	3183	2874	3961	-643	195	-394	O
HETATM	612	O	HOH A 237	-1.199	12.650	-24.605	1.00	28.72		O
ANISOU	612	O	HOH A 237	3654	2552	4707	-297	-115	-503	O
HETATM	613	O	HOH A 238	-7.144	5.616	4.399	1.00	31.43		O
ANISOU	613	O	HOH A 238	4570	2735	4637	571	633	-223	O
HETATM	614	O	HOH A 239	-2.967	7.314	3.066	1.00	30.50		O
ANISOU	614	O	HOH A 239	4921	2268	4399	15	165	-517	O
HETATM	615	O	HOH A 240	-7.778	8.335	-13.432	1.00	38.02		O
ANISOU	615	O	HOH A 240	4855	4692	4898	1303	-488	-294	O
HETATM	616	O	HOH A 241	0.290	-8.273	-9.973	1.00	27.17		O

ANISOU	616	O	HOH A 241	3516	3113	3691	1182	313	277	O
HETATM	617	O	HOH A 242	2.347	9.016	6.032	1.00	43.86		O
ANISOU	617	O	HOH A 242	6442	3649	6572	-1681	628	-142	O
HETATM	618	O	HOH A 243	5.830	3.305	3.984	1.00	39.16		O
ANISOU	618	O	HOH A 243	3858	4989	6031	-895	-1710	952	O
HETATM	619	O	HOH A 244	7.765	5.493	1.254	1.00	64.61		O
ANISOU	619	O	HOH A 244	5470	5662	13417	1054	577	-2395	O
HETATM	620	O	HOH A 245	7.623	6.811	-1.183	1.00	55.02		O
ANISOU	620	O	HOH A 245	6938	6040	7925	-586	3284	-890	O
HETATM	621	O	HOH A 246	8.861	0.126	-9.839	1.00	45.23		O
ANISOU	621	O	HOH A 246	4136	6350	6699	560	1369	2188	O
HETATM	622	O	HOH A 247	9.514	3.058	-6.160	1.00	44.59		O
ANISOU	622	O	HOH A 247	6123	4693	6125	-204	-665	-1094	O
HETATM	623	O	HOH A 248	9.951	0.432	-5.269	1.00	45.44		O
ANISOU	623	O	HOH A 248	3065	7107	7093	-96	-576	-3114	O
HETATM	624	O	HOH A 249	-4.168	10.730	-18.627	1.00	20.60		O
ANISOU	624	O	HOH A 249	2855	1718	3252	221	-173	14	O
HETATM	625	O	HOH A 250	-3.180	11.072	-26.087	1.00	30.86		O
ANISOU	625	O	HOH A 250	4985	2517	4222	-560	408	258	O
HETATM	626	O	HOH A 251	-2.466	13.105	-28.138	1.00	46.90		O
ANISOU	626	O	HOH A 251	8693	3807	5319	1082	994	1116	O
HETATM	627	O	HOH A 252	-10.487	8.382	-15.000	0.50	32.50		O
ANISOU	627	O	HOH A 252	3781	4716	3848	552	604	1337	O
HETATM	628	O	HOH A 253	-13.058	-3.117	-17.601	1.00	40.37		O
ANISOU	628	O	HOH A 253	4272	3803	7261	-656	-151	-1232	O
HETATM	629	O	HOH A 254	-12.401	-3.541	-13.825	1.00	55.40		O
ANISOU	629	O	HOH A 254	9652	4972	6425	-855	53	1702	O
HETATM	630	O	HOH A 255	-9.737	0.565	-11.326	1.00	36.24		O
ANISOU	630	O	HOH A 255	3346	5962	4462	495	479	80	O
HETATM	631	O	HOH A 256	-9.996	-2.331	-11.645	1.00	51.95		O
ANISOU	631	O	HOH A 256	8425	6025	5287	-1779	1536	844	O
HETATM	632	O	HOH A 257	-10.303	3.992	9.471	1.00	33.34		O
ANISOU	632	O	HOH A 257	3606	2601	6458	-72	-910	-165	O
HETATM	633	O	HOH A 258	-10.265	2.496	11.682	1.00	36.71		O
ANISOU	633	O	HOH A 258	4112	3207	6627	945	-19	404	O
HETATM	634	O	HOH A 259	5.075	-5.162	-2.051	1.00	44.14		O
ANISOU	634	O	HOH A 259	5345	6083	5341	2915	1061	1118	O
HETATM	635	O	HOH A 260	7.799	-1.435	-1.628	1.00	44.95		O
ANISOU	635	O	HOH A 260	5505	6608	4964	2344	-1518	-806	O
HETATM	636	O	HOH A 261	-8.208	16.400	-20.331	1.00	36.90		O
ANISOU	636	O	HOH A 261	4113	4015	5892	-812	731	-1124	O
HETATM	637	O	HOH A 262	-10.191	15.734	-18.403	0.50	34.87		O
ANISOU	637	O	HOH A 262	3782	3441	6023	169	377	-418	O
HETATM	638	O	HOH A 263	-15.642	8.143	-19.912	1.00	43.10		O
ANISOU	638	O	HOH A 263	4192	6048	6136	-994	64	1385	O
HETATM	639	O	HOH A 264	-8.234	4.649	12.473	1.00	113.09		O
ANISOU	639	O	HOH A 264	26290	7910	8768	6416	7413	670	O
HETATM	640	O	HOH A 265	7.201	4.585	-7.653	1.00	45.43		O
ANISOU	640	O	HOH A 265	4940	8013	4305	-2529	266	-571	O
HETATM	641	O	HOH A 266	-11.054	-4.604	-17.070	0.50	33.63		O
ANISOU	641	O	HOH A 266	4674	2648	5454	-509	-889	-4	O
HETATM	642	O	HOH A 267	1.471	-6.433	-20.456	0.50	35.63		O
ANISOU	642	O	HOH A 267	5505	3538	4494	249	279	-259	O
HETATM	643	O	HOH A 268	6.072	-4.148	-14.926	1.00	41.40		O
ANISOU	643	O	HOH A 268	6838	3456	5436	1281	2206	562	O
HETATM	644	O	HOH A 269	5.869	8.208	-1.107	0.50	45.75		O

ANISOU	644	O	HOH A 269	6524	4822	6037	-935	-927	-276	O
HETATM	645	O	HOH A 270	-0.028	9.926	9.373	0.50	51.17		O
ANISOU	645	O	HOH A 270	3836	9268	6339	-2288	1767	-3587	O
HETATM	646	O	HOH A 271	-7.384	6.482	6.969	1.00	34.53		O
ANISOU	646	O	HOH A 271	4442	2938	5740	766	766	-253	O
HETATM	647	O	AHOH A 272	-6.243	-7.856	-18.372	0.50	37.97		O
ANISOU	647	O	AHOH A 272	6359	3629	4438	175	-138	-807	O
HETATM	648	O	BHOH A 272	-5.609	-8.461	-16.705	0.50	38.45		O
ANISOU	648	O	BHOH A 272	5311	3226	6070	-317	-40	747	O
HETATM	649	O	HOH A 273	2.287	3.921	9.668	0.50	49.09		O
ANISOU	649	O	HOH A 273	8460	4255	5936	79	-391	2175	O
HETATM	650	O	HOH A 274	-1.140	6.737	11.085	0.50	30.77		O
ANISOU	650	O	HOH A 274	4300	3575	3814	209	497	20	O
HETATM	651	O	HOH A 275	-0.510	-7.490	-5.475	1.00	52.18		O
ANISOU	651	O	HOH A 275	10433	2578	6814	-760	2553	-1512	O
HETATM	652	O	HOH A 276	-12.897	5.720	-14.135	0.50	27.97		O
ANISOU	652	O	HOH A 276	3568	3527	3532	178	167	-309	O
HETATM	653	O	HOH A 277	3.719	8.141	-2.619	0.50	32.92		O
ANISOU	653	O	HOH A 277	4463	2763	5280	-940	-672	-522	O
HETATM	654	O	HOH A 278	4.551	7.369	7.001	0.50	36.36		O
ANISOU	654	O	HOH A 278	4808	2798	6206	-774	402	247	O
HETATM	655	O	HOH A 279	1.787	8.779	10.502	0.50	37.48		O
ANISOU	655	O	HOH A 279	4443	4398	5400	-87	-94	102	O
HETATM	656	O	HOH A 280	-11.605	17.221	-21.811	0.50	40.39		O
ANISOU	656	O	HOH A 280	4953	3736	6658	-693	1537	-354	O
HETATM	657	O	HOH A 281	-1.539	8.821	1.241	1.00	35.87		O
ANISOU	657	O	HOH A 281	5334	3512	4783	30	484	42	O
HETATM	658	O	HOH A 282	6.541	-4.764	1.452	1.00	53.26		O
ANISOU	658	O	HOH A 282	5674	7651	6909	2279	-1107	-355	O
HETATM	659	O	HOH A 283	0.942	8.099	-0.326	1.00	30.53		O
ANISOU	659	O	HOH A 283	4699	1918	4980	-345	-645	-305	O
HETATM	660	O	HOH B 201	1.208	4.167	-3.482	1.00	18.99		O
ANISOU	660	O	HOH B 201	2613	1473	3128	62	103	90	O
HETATM	661	O	HOH B 202	1.032	11.287	-14.460	1.00	29.99		O
ANISOU	661	O	HOH B 202	4798	2030	4563	-38	-1182	302	O
HETATM	662	O	HOH B 203	-4.929	6.390	-10.991	1.00	23.33		O
ANISOU	662	O	HOH B 203	2754	2153	3955	503	5	457	O
HETATM	663	O	HOH B 204	-2.808	9.187	-13.976	1.00	30.03		O
ANISOU	663	O	HOH B 204	3322	1883	6202	333	368	511	O
HETATM	664	O	HOH B 205	0.236	5.707	-1.401	1.00	21.69		O
ANISOU	664	O	HOH B 205	3031	1549	3661	-64	722	-43	O
HETATM	665	O	HOH B 206	-2.107	9.244	-17.147	1.00	20.68		O
ANISOU	665	O	HOH B 206	2824	1832	3198	94	-131	-1	O
HETATM	666	O	HOH B 207	1.120	8.388	-17.725	1.00	19.98		O
ANISOU	666	O	HOH B 207	2582	1816	3190	33	-140	-12	O
HETATM	667	O	HOH B 208	-2.765	9.025	-7.569	1.00	23.32		O
ANISOU	667	O	HOH B 208	3262	1631	3967	425	-139	13	O
HETATM	668	O	HOH B 209	3.323	4.503	-5.373	1.00	21.06		O
ANISOU	668	O	HOH B 209	2437	2347	3219	-208	69	-114	O
HETATM	669	O	HOH B 210	5.514	-2.950	9.131	1.00	27.08		O
ANISOU	669	O	HOH B 210	2599	3651	4036	-650	-139	360	O
HETATM	670	O	HOH B 211	1.549	9.196	-11.289	1.00	24.52		O
ANISOU	670	O	HOH B 211	3663	1671	3980	-455	96	-98	O
HETATM	671	O	HOH B 212	-7.533	-3.787	-28.440	1.00	21.94		O
ANISOU	671	O	HOH B 212	2876	1826	3633	-267	-79	-89	O
HETATM	672	O	HOH B 213	-6.950	6.216	-4.271	1.00	25.45		O

ANISOU	672	O	HOH B 213	3068	2790	3809	850	68	233	O
HETATM	673	O	HOH B 214	-0.843	2.370	-27.422	1.00	22.00		O
ANISOU	673	O	HOH B 214	2526	2356	3475	55	83	-121	O
HETATM	674	O	HOH B 215	2.859	0.560	-11.571	1.00	19.88		O
ANISOU	674	O	HOH B 215	2375	1753	3422	176	234	146	O
HETATM	675	O	HOH B 216	-7.216	5.223	-10.481	1.00	27.23		O
ANISOU	675	O	HOH B 216	3226	3055	4064	429	-298	191	O
HETATM	676	O	HOH B 217	1.364	3.522	-26.298	1.00	24.69		O
ANISOU	676	O	HOH B 217	2968	2684	3728	-333	506	-134	O
HETATM	677	O	HOH B 218	-7.253	1.815	-5.266	1.00	26.95		O
ANISOU	677	O	HOH B 218	2879	4109	3249	264	87	145	O
HETATM	678	O	HOH B 219	-12.908	0.487	1.932	1.00	27.48		O
ANISOU	678	O	HOH B 219	2510	4243	3687	-108	-44	820	O
HETATM	679	O	HOH B 220	-4.881	8.664	-9.367	1.00	26.12		O
ANISOU	679	O	HOH B 220	3294	2689	3939	444	-173	-253	O
HETATM	680	O	HOH B 221	-6.855	-2.260	-3.880	1.00	30.59		O
ANISOU	680	O	HOH B 221	3920	3587	4115	-877	342	-1077	O
HETATM	681	O	HOH B 222	-0.764	10.780	-8.470	1.00	28.92		O
ANISOU	681	O	HOH B 222	3894	2097	4994	-204	493	367	O
HETATM	682	O	HOH B 223	4.567	1.638	-9.587	1.00	21.53		O
ANISOU	682	O	HOH B 223	2373	2332	3473	92	154	-3	O
HETATM	683	O	HOH B 224	-9.427	2.142	-3.659	1.00	29.63		O
ANISOU	683	O	HOH B 224	3099	4067	4092	329	416	983	O
HETATM	684	O	HOH B 225	1.318	6.994	-22.868	1.00	27.93		O
ANISOU	684	O	HOH B 225	3055	3135	4419	-955	436	-283	O
HETATM	685	O	HOH B 226	4.322	-2.546	-24.998	1.00	33.62		O
ANISOU	685	O	HOH B 226	3418	3595	5760	841	-383	-1419	O
HETATM	686	O	HOH B 227	-0.568	10.709	-12.365	1.00	38.66		O
ANISOU	686	O	HOH B 227	4687	2774	7227	-69	-1041	-59	O
HETATM	687	O	HOH B 228	-1.492	-5.561	-1.404	1.00	91.33		O
ANISOU	687	O	HOH B 228	20165	4937	9597	1784	2806	-1276	O
HETATM	688	O	HOH B 229	-6.143	8.785	-5.168	1.00	31.08		O
ANISOU	688	O	HOH B 229	3850	3117	4841	334	42	1170	O
HETATM	689	O	HOH B 230	-6.494	-0.685	-6.185	1.00	27.45		O
ANISOU	689	O	HOH B 230	2883	3521	4022	-25	85	-557	O
HETATM	690	O	HOH B 231	-0.963	0.258	-19.741	1.00	24.36		O
ANISOU	690	O	HOH B 231	2966	2255	4032	325	132	-24	O
HETATM	691	O	HOH B 232	-11.485	4.360	-0.265	1.00	31.52		O
ANISOU	691	O	HOH B 232	3198	2712	6066	628	409	297	O
HETATM	692	O	HOH B 233	0.761	0.345	-17.322	1.00	28.54		O
ANISOU	692	O	HOH B 233	4644	2172	4025	128	625	257	O
HETATM	693	O	HOH B 234	-6.582	2.748	-9.500	1.00	25.77		O
ANISOU	693	O	HOH B 234	2958	2659	4174	40	-537	285	O
HETATM	694	O	HOH B 235	-2.468	-14.775	5.778	1.00	60.04		O
ANISOU	694	O	HOH B 235	5572	6312	10927	687	624	-1562	O
HETATM	695	O	HOH B 236	-9.975	5.017	-2.626	1.00	32.96		O
ANISOU	695	O	HOH B 236	4050	4037	4436	72	297	713	O
HETATM	696	O	HOH B 237	-3.802	-8.987	0.610	1.00	40.56		O
ANISOU	696	O	HOH B 237	6819	3316	5276	-551	1407	285	O
HETATM	697	O	HOH B 238	5.576	8.867	-5.530	1.00	39.43		O
ANISOU	697	O	HOH B 238	5197	4878	4904	-1770	-985	27	O
HETATM	698	O	HOH B 239	0.480	-2.203	-20.508	1.00	37.54		O
ANISOU	698	O	HOH B 239	4418	4737	5107	1193	-91	-468	O
HETATM	699	O	HOH B 240	-12.279	-6.259	3.031	1.00	39.02		O
ANISOU	699	O	HOH B 240	4242	5382	5200	-1741	673	-493	O
HETATM	700	O	HOH B 241	-8.420	2.908	-7.461	1.00	34.05		O

ANISOU	700	O	HOH B 241	4404	4454	4077	1230	77	297	O
HETATM	701	O	HOH B 242	-10.273	-9.187	2.528	1.00	41.62		O
ANISOU	701	O	HOH B 242	5966	4686	5158	-2102	5	1009	O
HETATM	702	O	HOH B 243	4.413	5.754	-19.474	1.00	35.34		O
ANISOU	702	O	HOH B 243	3352	5036	5039	-1119	347	-457	O
HETATM	703	O	HOH B 244	-0.523	5.710	-26.410	1.00	28.37		O
ANISOU	703	O	HOH B 244	3209	3201	4366	-328	509	-110	O
HETATM	704	O	HOH B 245	-13.296	-3.589	2.389	1.00	43.39		O
ANISOU	704	O	HOH B 245	4011	6231	6243	177	-701	-1001	O
HETATM	705	O	HOH B 246	3.910	6.115	-22.191	1.00	31.12		O
ANISOU	705	O	HOH B 246	3318	3722	4785	-477	-369	474	O
HETATM	706	O	HOH B 247	5.326	6.311	-5.026	1.00	82.42		O
ANISOU	706	O	HOH B 247	11574	8469	11271	-3133	-5399	6007	O
HETATM	707	O	HOH B 248	4.443	7.607	-14.904	1.00	27.67		O
ANISOU	707	O	HOH B 248	2871	2857	4785	-351	-181	707	O
HETATM	708	O	HOH B 249	3.874	9.504	-12.859	1.00	33.03		O
ANISOU	708	O	HOH B 249	3890	3827	4831	-797	290	648	O
HETATM	709	O	HOH B 250	-10.648	6.203	1.297	0.50	30.90		O
ANISOU	709	O	HOH B 250	4705	2511	4523	274	255	360	O
HETATM	710	O	HOH B 251	-4.093	13.457	-15.884	1.00	33.85		O
ANISOU	710	O	HOH B 251	3413	4675	4770	720	-301	-1439	O
HETATM	711	O	HOH B 252	-8.480	5.016	-6.150	1.00	37.19		O
ANISOU	711	O	HOH B 252	4094	5567	4468	-609	198	-404	O
HETATM	712	O	HOH B 253	-8.275	6.732	-8.510	1.00	39.66		O
ANISOU	712	O	HOH B 253	4432	6236	4399	1298	-426	-1019	O
HETATM	713	O	HOH B 254	-7.029	8.894	-7.746	1.00	32.37		O
ANISOU	713	O	HOH B 254	4189	3701	4410	1087	-101	506	O
HETATM	714	O	HOH B 255	-5.872	7.775	3.047	1.00	32.68		O
ANISOU	714	O	HOH B 255	5065	2364	4988	669	500	15	O
HETATM	715	O	HOH B 256	0.102	-14.670	9.631	1.00	37.03		O
ANISOU	715	O	HOH B 256	4927	2523	6617	253	181	48	O
HETATM	716	O	HOH B 257	7.620	2.741	2.779	0.50	34.87		O
ANISOU	716	O	HOH B 257	2704	5122	5421	665	158	-897	O
HETATM	717	O	HOH B 258	7.157	2.257	-10.065	1.00	35.13		O
ANISOU	717	O	HOH B 258	2444	5144	5758	222	122	1879	O
HETATM	718	O	HOH B 259	8.052	7.180	-8.396	1.00	54.77		O
ANISOU	718	O	HOH B 259	6130	9365	5315	-1718	-676	-801	O
HETATM	719	O	HOH B 260	7.020	-2.654	-25.999	1.00	35.32		O
ANISOU	719	O	HOH B 260	3206	3170	7042	78	1084	-807	O
HETATM	720	O	HOH B 261	-0.005	10.737	-18.644	1.00	20.32		O
ANISOU	720	O	HOH B 261	2671	1772	3275	76	-208	-33	O
HETATM	721	O	HOH B 262	-2.116	12.137	-17.086	1.00	20.92		O
ANISOU	721	O	HOH B 262	2913	1813	3221	-15	-193	-147	O
HETATM	722	O	HOH B 263	3.917	8.126	-17.425	1.00	38.73		O
ANISOU	722	O	HOH B 263	2599	7142	4975	-894	45	-562	O
HETATM	723	O	HOH B 264	4.746	-1.496	-21.640	1.00	47.14		O
ANISOU	723	O	HOH B 264	7382	5315	5214	3120	-1025	-1646	O
HETATM	724	O	HOH B 265	4.739	4.057	-24.046	1.00	35.72		O
ANISOU	724	O	HOH B 265	4053	3999	5519	-1522	-737	421	O
HETATM	725	O	HOH B 266	4.040	11.534	-17.525	1.00	38.94		O
ANISOU	725	O	HOH B 266	3784	2887	8124	134	18	-790	O
HETATM	726	O	HOH B 267	6.503	9.438	-9.282	1.00	107.68		O
ANISOU	726	O	HOH B 267	7985	15830	17098	-4205	5000	-5534	O
HETATM	727	O	HOH B 268	7.885	12.176	-7.962	1.00	46.26		O
ANISOU	727	O	HOH B 268	4531	5048	7998	-1489	-141	1016	O
HETATM	728	O	HOH B 269	-2.797	8.488	-26.981	1.00	32.64		O

ANISOU	728	O	HOH B 269	4521	3306	4572	-339	-454	501	O
HETATM	729	O	HOH B 270	-2.080	12.157	-20.062	1.00	20.75		O
ANISOU	729	O	HOH B 270	2844	1634	3405	-4	-353	-21	O
HETATM	730	O	HOH B 271	1.392	-10.091	4.267	1.00	48.78		O
ANISOU	730	O	HOH B 271	6494	4383	7655	-109	2158	-1768	O
HETATM	731	O	HOH B 272	-1.220	-11.002	2.292	1.00	44.13		O
ANISOU	731	O	HOH B 272	5183	6601	4982	1182	1028	338	O
HETATM	732	O	HOH B 273	-4.077	-6.118	-1.618	1.00	91.16		O
ANISOU	732	O	HOH B 273	20103	2930	11603	3457	2426	-4366	O
HETATM	733	O	HOH B 274	-4.825	-4.059	-4.000	1.00	35.08		O
ANISOU	733	O	HOH B 274	4110	3385	5830	-1047	849	-1504	O
HETATM	734	O	HOH B 275	-9.477	4.959	3.159	1.00	30.52		O
ANISOU	734	O	HOH B 275	4038	2270	5289	583	1174	800	O
HETATM	735	O	HOH B 276	-2.091	-8.116	-0.892	1.00	115.08		O
ANISOU	735	O	HOH B 276	27773	3331	12621	-473	3351	1236	O
HETATM	736	O	HOH B 277	0.788	-7.488	0.563	1.00	100.48		O
ANISOU	736	O	HOH B 277	13724	6460	17994	1204	2824	-1127	O
HETATM	737	O	HOH B 278	-11.763	2.704	-5.070	0.50	33.48		O
ANISOU	737	O	HOH B 278	3318	4599	4803	1064	-89	890	O
HETATM	738	O	HOH B 279	-4.548	12.687	-5.905	1.00	50.61		O
ANISOU	738	O	HOH B 279	6072	4619	8539	433	-295	-2169	O
HETATM	739	O	HOH B 280	-3.699	14.543	-4.043	0.50	37.16		O
ANISOU	739	O	HOH B 280	5562	2876	5679	222	1514	-13	O
HETATM	740	O	HOH B 281	7.368	0.558	-21.270	0.50	116.86		O
ANISOU	740	O	HOH B 281	6066	29679	8654	5224	-1497	-5527	O
HETATM	741	O	HOH B 282	7.459	4.907	-20.980	1.00	31.27		O
ANISOU	741	O	HOH B 282	2612	5169	4099	141	82	-184	O
HETATM	742	O	HOH B 283	-11.007	4.398	-9.441	0.50	44.87		O
ANISOU	742	O	HOH B 283	4336	8746	3963	-2176	670	-1173	O
HETATM	743	O	HOH B 284	-8.065	-5.150	-2.303	0.50	33.22		O
ANISOU	743	O	HOH B 284	4126	4788	3706	-404	-737	-199	O
HETATM	744	O	AHOH B 285	3.281	-0.970	-14.016	0.00	30.46		O
ANISOU	744	O	AHOH B 285	4205	3413	3957	722	447	-238	O
HETATM	745	O	BHOH B 285	4.696	-0.403	-13.308	0.50	30.55		O
ANISOU	745	O	BHOH B 285	5017	2847	3741	1776	1611	1055	O
HETATM	746	O	HOH B 286	-7.390	10.652	-2.905	0.50	51.22		O
ANISOU	746	O	HOH B 286	4287	7663	7511	1005	-360	721	O
HETATM	747	O	AHOH B 287	-0.115	-4.964	-23.684	0.50	40.01		O
ANISOU	747	O	AHOH B 287	3313	4142	7745	325	950	479	O
HETATM	748	O	BHOH B 287	-1.617	-4.775	-25.825	0.50	33.51		O
ANISOU	748	O	BHOH B 287	5764	1417	5550	600	-762	224	O
HETATM	749	O	HOH B 288	2.883	4.930	-28.258	0.50	29.95		O
ANISOU	749	O	HOH B 288	3243	3275	4860	-485	244	727	O
HETATM	750	O	HOH B 289	1.349	7.221	-25.391	0.50	36.90		O
ANISOU	750	O	HOH B 289	3463	6711	3845	-1508	566	-358	O
HETATM	751	O	HOH B 290	-5.602	10.344	-0.148	1.00	88.23		O
ANISOU	751	O	HOH B 290	5543	10851	17129	703	2732	-1796	O
HETATM	752	O	AHOH B 291	-11.204	-1.946	-3.074	0.50	32.47		O
ANISOU	752	O	AHOH B 291	3794	4167	4374	-140	-287	197	O
HETATM	753	O	BHOH B 291	-9.727	-1.753	-3.519	0.50	28.64		O
ANISOU	753	O	BHOH B 291	3080	4214	3586	-226	-109	-367	O
HETATM	754	O	HOH B 292	3.035	10.214	-20.664	1.00	45.28		O
ANISOU	754	O	HOH B 292	5553	3829	7820	-603	292	-443	O
HETATM	755	O	HOH B 293	3.919	10.217	-22.849	1.00	51.99		O
ANISOU	755	O	HOH B 293	6933	4557	8264	-1564	1697	-1140	O
HETATM	756	O	HOH B 294	2.777	-4.135	-26.264	1.00	62.82		O

ANISOU	756	O	HOH	B	294	6572	6106	11190	383	454	-1432	O
HETATM	757	O	HOH	B	295	10.997	0.183	-19.547	1.00	43.78		O
ANISOU	757	O	HOH	B	295	5694	6380	4558	-2444	-910	306	O
HETATM	758	O	HOH	B	296	9.623	3.051	-18.066	1.00	54.33		O
ANISOU	758	O	HOH	B	296	3240	9812	7591	66	-1597	2395	O
HETATM	759	O	HOH	B	297	8.538	5.519	-17.276	1.00	36.67		O
ANISOU	759	O	HOH	B	297	3983	5411	4538	-1536	738	-460	O
HETATM	760	O	HOH	B	298	11.494	3.365	-14.553	1.00	47.74		O
ANISOU	760	O	HOH	B	298	5135	3548	9454	-334	2302	-663	O
HETATM	761	O	HOH	B	299	9.634	4.034	-15.804	1.00	58.14		O
ANISOU	761	O	HOH	B	299	4550	9345	8195	-989	1336	1572	O
HETATM	762	O	HOH	B	300	9.242	2.528	-13.567	1.00	39.07		O
ANISOU	762	O	HOH	B	300	4961	3977	5907	1141	1444	516	O
HETATM	763	O	HOH	B	301	8.060	7.240	-15.865	0.50	36.86		O
ANISOU	763	O	HOH	B	301	4912	4149	4942	379	-324	1125	O
HETATM	764	O	HOH	B	302	7.362	7.443	-21.063	0.50	32.97		O
ANISOU	764	O	HOH	B	302	3879	3990	4658	-884	565	-264	O
HETATM	765	O	HOH	B	303	7.048	-0.623	-17.035	1.00	41.84		O
ANISOU	765	O	HOH	B	303	5338	5546	5011	518	332	340	O
HETATM	766	O	HOH	B	304	7.023	10.304	-3.981	1.00	44.49		O
ANISOU	766	O	HOH	B	304	6078	5021	5804	-1367	-356	-626	O
CONECT	150	162										
CONECT	162	150	163	164	165							
CONECT	163	162										
CONECT	164	162										
CONECT	165	162	166									
CONECT	166	165	167									
CONECT	167	166	168	169								
CONECT	168	167	172									
CONECT	169	167	170	171								
CONECT	170	169	185									
CONECT	171	169	172									
CONECT	172	168	171	173								
CONECT	173	172	174	184								
CONECT	174	173	175	176								
CONECT	175	174										
CONECT	176	174	177									
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CONECT	178	177										
CONECT	179	177	180	181	184							
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CONECT	184	173	179									
CONECT	185	170										
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CONECT	457	456	461									
CONECT	458	456	459	460								
CONECT	459	458	472									

CONECT	460	458	461		
CONECT	461	457	460	462	
CONECT	462	461	463	471	
CONECT	463	462	464	465	
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CONECT	468	466	469	471	
CONECT	469	468	470		
CONECT	470	469			
CONECT	471	462	468		
CONECT	472	459			
CONECT	536	583	624	665	720
CONECT	536	721	729		
CONECT	537	538			
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CONECT	567	565	569		
CONECT	568	566	570		
CONECT	569	567	571		
CONECT	570	568	572		
CONECT	571	569	573		
CONECT	572	570	574		
CONECT	573	571	575		
CONECT	574	572			
CONECT	575	573			
CONECT	583	536			
CONECT	624	536			


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CONNECT 665 536
CONNECT 720 536
CONNECT 721 536
CONNECT 729 536
MASTER 319 0 5 0 0 0 6 6 703 2 95 2
END
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File A-3: PDB coordinates for the crystal structure of 5-formyl-2'-deoxycytidine in B-type DNA, DDD^f.

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REMARK 3
REMARK 3 REFINEMENT.
REMARK 3 PROGRAM : REFMAC 5.8.0049
REMARK 3 AUTHORS : MURSHUDOV,SKUBAK,LEBEDEV,PANNU,
REMARK 3 STEINER,NICHOLLS,WINN,LONG,VAGIN
REMARK 3
REMARK 3 REFINEMENT TARGET : MAXIMUM LIKELIHOOD
REMARK 3
REMARK 3 DATA USED IN REFINEMENT.
REMARK 3 RESOLUTION RANGE HIGH (ANGSTROMS) : 1.74
REMARK 3 RESOLUTION RANGE LOW (ANGSTROMS) : 35.06
REMARK 3 DATA CUTOFF (SIGMA(F)) : NONE
REMARK 3 COMPLETENESS FOR RANGE (%) : 98.98
REMARK 3 NUMBER OF REFLECTIONS : 7121
REMARK 3
REMARK 3 FIT TO DATA USED IN REFINEMENT.
REMARK 3 CROSS-VALIDATION METHOD : THROUGHOUT
REMARK 3 FREE R VALUE TEST SET SELECTION : RANDOM
REMARK 3 R VALUE (WORKING + TEST SET) : 0.23571
REMARK 3 R VALUE (WORKING SET) : 0.23315
REMARK 3 FREE R VALUE : 0.29312
REMARK 3 FREE R VALUE TEST SET SIZE (%) : 4.6
REMARK 3 FREE R VALUE TEST SET COUNT : 344
REMARK 3
REMARK 3 FIT IN THE HIGHEST RESOLUTION BIN.
REMARK 3 TOTAL NUMBER OF BINS USED : 20
REMARK 3 BIN RESOLUTION RANGE HIGH : 1.740
REMARK 3 BIN RESOLUTION RANGE LOW : 1.785
REMARK 3 REFLECTION IN BIN (WORKING SET) : 521
REMARK 3 BIN COMPLETENESS (WORKING+TEST) (%) : 99.28
REMARK 3 BIN R VALUE (WORKING SET) : 0.300
REMARK 3 BIN FREE R VALUE SET COUNT : 27
REMARK 3 BIN FREE R VALUE : 0.358
REMARK 3
REMARK 3 NUMBER OF NON-HYDROGEN ATOMS USED IN REFINEMENT.
REMARK 3 ALL ATOMS : 547
REMARK 3
REMARK 3 B VALUES.
REMARK 3 FROM WILSON PLOT (A**2) : NULL
REMARK 3 MEAN B VALUE (OVERALL, A**2) : 35.181
REMARK 3 OVERALL ANISOTROPIC B VALUE.
REMARK 3 B11 (A**2) : 2.03
REMARK 3 B22 (A**2) : 0.80
REMARK 3 B33 (A**2) : -2.82
REMARK 3 B12 (A**2) : 0.00
REMARK 3 B13 (A**2) : -0.00
REMARK 3 B23 (A**2) : -0.00
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REMARK 3
REMARK 3 ESTIMATED OVERALL COORDINATE ERROR.
REMARK 3 ESU BASED ON R VALUE (A): 0.138
REMARK 3 ESU BASED ON FREE R VALUE (A): 0.144
REMARK 3 ESU BASED ON MAXIMUM LIKELIHOOD (A): 0.117
REMARK 3 ESU FOR B VALUES BASED ON MAXIMUM LIKELIHOOD (A**2): 3.908
REMARK 3
REMARK 3 CORRELATION COEFFICIENTS.
REMARK 3 CORRELATION COEFFICIENT FO-FC : 0.959
REMARK 3 CORRELATION COEFFICIENT FO-FC FREE : 0.932
REMARK 3
REMARK 3 RMS DEVIATIONS FROM IDEAL VALUES COUNT RMS WEIGHT
REMARK 3 BOND LENGTHS REFINED ATOMS (A): 552 ; 0.009 ; 0.011
REMARK 3 BOND LENGTHS OTHERS (A): 268 ; 0.002 ; 0.020
REMARK 3 BOND ANGLES REFINED ATOMS (DEGREES): 850 ; 1.747 ; 1.182
REMARK 3 BOND ANGLES OTHERS (DEGREES): 634 ; 1.611 ; 3.000
REMARK 3 CHIRAL-CENTER RESTRAINTS (A**3): 72 ; 0.071 ; 0.200
REMARK 3 GENERAL PLANES REFINED ATOMS (A): 290 ; 0.023 ; 0.020
REMARK 3 GENERAL PLANES OTHERS (A): 122 ; 0.003 ; 0.020
REMARK 3
REMARK 3 ISOTROPIC THERMAL FACTOR RESTRAINTS. COUNT RMS WEIGHT
REMARK 3 SIDE-CHAIN BOND REFINED ATOMS (A**2): 552 ; 3.873 ; 3.639
REMARK 3 SIDE-CHAIN BOND OTHER ATOMS (A**2) : 551 ; 3.874 ; 3.640
REMARK 3 SIDE-CHAIN ANGLE OTHER ATOMS (A**2) : 850 ; 5.591 ; 5.470
REMARK 3 LONG RANGE B REFINED ATOMS (A**2) : 819 ; 5.904 ; 34.255
REMARK 3 LONG RANGE B OTHER ATOMS (A**2) : 804 ; 5.858 ; 34.257
REMARK 3
REMARK 3 NCS RESTRAINTS STATISTICS
REMARK 3 NUMBER OF NCS GROUPS : NULL
REMARK 3
REMARK 3 TWIN DETAILS
REMARK 3 NUMBER OF TWIN DOMAINS : NULL
REMARK 3
REMARK 3 TLS DETAILS
REMARK 3 NUMBER OF TLS GROUPS : NULL
REMARK 3
REMARK 3 BULK SOLVENT MODELLING.
REMARK 3 METHOD USED : MASK
REMARK 3 PARAMETERS FOR MASK CALCULATION
REMARK 3 VDW PROBE RADIUS : 1.20
REMARK 3 ION PROBE RADIUS : 0.80
REMARK 3 SHRINKAGE RADIUS : 0.80
REMARK 3
REMARK 3 OTHER REFINEMENT REMARKS:
REMARK 3 HYDROGENS HAVE BEEN ADDED IN THE RIDING POSITIONS
REMARK 3 U VALUES : REFINED INDIVIDUALLY
REMARK 3
CRYST1 25.316 41.470 65.669 90.00 90.00 90.00 P 21 21 21
SCALE1 0.039501 0.000000 0.000000 0.000000
SCALE2 -0.000000 0.024114 0.000000 0.000000
SCALE3 0.000000 -0.000000 0.015228 0.000000
ATOM 1 O5' DC A 1 7.063 14.256 22.460 1.00 53.02 A O
ATOM 2 C5' DC A 1 6.906 13.495 23.675 1.00 44.30 A C
ATOM 3 C4' DC A 1 7.388 12.081 23.461 1.00 41.26 A C

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ATOM	4	C3'	DC	A	1	7.300	11.584	22.015	1.00	47.01	A	C
ATOM	5	O3'	DC	A	1	8.504	10.848	21.790	1.00	49.71	A	O
ATOM	6	C2'	DC	A	1	6.101	10.652	22.051	1.00	45.11	A	C
ATOM	7	C1'	DC	A	1	6.300	10.047	23.426	1.00	38.25	A	C
ATOM	8	O4'	DC	A	1	6.583	11.159	24.234	1.00	38.74	A	O
ATOM	9	N1	DC	A	1	5.148	9.354	23.997	1.00	34.51	A	N
ATOM	10	C6	DC	A	1	3.915	9.933	23.917	1.00	38.58	A	C
ATOM	11	C5	DC	A	1	2.834	9.319	24.412	1.00	44.80	A	C
ATOM	12	C4	DC	A	1	3.020	8.032	25.005	1.00	38.41	A	C
ATOM	13	N4	DC	A	1	1.987	7.382	25.525	1.00	41.79	A	N
ATOM	14	N3	DC	A	1	4.223	7.469	25.108	1.00	30.93	A	N
ATOM	15	C2	DC	A	1	5.300	8.084	24.591	1.00	31.81	A	C
ATOM	16	O2	DC	A	1	6.422	7.562	24.639	1.00	33.15	A	O
ATOM	17	P	DG	A	2	9.192	10.792	20.363	1.00	59.87	A	P
ATOM	18	OP1	DG	A	2	10.029	12.012	20.229	1.00	60.91	A	O
ATOM	19	O5'	DG	A	2	10.177	9.547	20.515	1.00	49.54	A	O
ATOM	20	C5'	DG	A	2	10.843	9.268	21.759	1.00	51.72	A	C
ATOM	21	C4'	DG	A	2	11.008	7.777	21.926	1.00	45.38	A	C
ATOM	22	C3'	DG	A	2	11.207	6.999	20.620	1.00	43.40	A	C
ATOM	23	O3'	DG	A	2	12.385	6.204	20.655	1.00	44.27	A	O
ATOM	24	C2'	DG	A	2	10.037	6.046	20.535	1.00	42.59	A	C
ATOM	25	C1'	DG	A	2	9.387	6.058	21.902	1.00	42.86	A	C
ATOM	26	O4'	DG	A	2	9.833	7.232	22.564	1.00	45.70	A	O
ATOM	27	N9	DG	A	2	7.920	6.111	21.844	1.00	36.66	A	N
ATOM	28	C4	DG	A	2	7.057	5.129	22.254	1.00	30.40	A	C
ATOM	29	C5	DG	A	2	5.798	5.599	21.951	1.00	31.88	A	C
ATOM	30	N7	DG	A	2	5.866	6.879	21.423	1.00	34.20	A	N
ATOM	31	C8	DG	A	2	7.143	7.133	21.356	1.00	34.88	A	C
ATOM	32	N3	DG	A	2	7.408	3.922	22.745	1.00	32.05	A	N
ATOM	33	C2	DG	A	2	6.341	3.189	23.054	1.00	28.25	A	C
ATOM	34	N2	DG	A	2	6.500	1.973	23.572	1.00	30.09	A	N
ATOM	35	N1	DG	A	2	5.050	3.604	22.863	1.00	28.69	A	N
ATOM	36	C6	DG	A	2	4.676	4.821	22.311	1.00	28.46	A	C
ATOM	37	O6	DG	A	2	3.480	5.099	22.193	1.00	32.17	A	O
ATOM	38	OP2	DG	A	2	8.161	10.441	19.332	1.00	47.41	A	O
ATOM	39	P	DC	A	3	12.899	5.496	19.330	1.00	51.42	A	P
ATOM	40	OP1	DC	A	3	14.356	5.350	19.468	1.00	57.83	A	O
ATOM	41	O5'	DC	A	3	12.171	4.087	19.303	1.00	43.82	A	O
ATOM	42	C5'	DC	A	3	12.528	3.078	20.259	1.00	39.21	A	C
ATOM	43	C4'	DC	A	3	11.504	1.979	20.180	1.00	36.20	A	C
ATOM	44	C3'	DC	A	3	11.370	1.330	18.811	1.00	33.94	A	C
ATOM	45	O3'	DC	A	3	12.156	0.142	18.796	1.00	40.30	A	O
ATOM	46	C2'	DC	A	3	9.912	0.925	18.746	1.00	38.63	A	C
ATOM	47	C1'	DC	A	3	9.243	1.687	19.852	1.00	37.19	A	C
ATOM	48	O4'	DC	A	3	10.216	2.552	20.428	1.00	32.62	A	O
ATOM	49	N1	DC	A	3	8.152	2.530	19.404	1.00	32.54	A	N
ATOM	50	C6	DC	A	3	8.401	3.731	18.804	1.00	33.17	A	C
ATOM	51	C5	DC	A	3	7.404	4.560	18.487	1.00	37.56	A	C
ATOM	52	C4	DC	A	3	6.081	4.146	18.818	1.00	32.41	A	C
ATOM	53	N4	DC	A	3	5.040	4.918	18.504	1.00	33.84	A	N
ATOM	54	N3	DC	A	3	5.834	2.962	19.372	1.00	30.95	A	N
ATOM	55	C2	DC	A	3	6.856	2.170	19.747	1.00	27.59	A	C
ATOM	56	O2	DC	A	3	6.670	1.072	20.272	1.00	30.70	A	O
ATOM	57	OP2	DC	A	3	12.277	6.183	18.156	1.00	49.20	A	O
ATOM	58	P	DG	A	4	12.291	-0.677	17.462	1.00	41.60	A	P
ATOM	59	OP1	DG	A	4	13.447	-1.554	17.603	1.00	49.14	A	O

ATOM	60	O5'	DG	A	4	11.045	-1.655	17.536	1.00	38.14	A	O
ATOM	61	C5'	DG	A	4	10.945	-2.646	18.574	1.00	39.91	A	C
ATOM	62	C4'	DG	A	4	9.681	-3.453	18.370	1.00	47.85	A	C
ATOM	63	C3'	DG	A	4	9.611	-4.190	17.030	1.00	47.02	A	C
ATOM	64	O3'	DG	A	4	9.131	-5.521	17.175	1.00	57.79	A	O
ATOM	65	C2'	DG	A	4	8.662	-3.365	16.192	1.00	39.78	A	C
ATOM	66	C1'	DG	A	4	7.789	-2.663	17.216	1.00	40.09	A	C
ATOM	67	O4'	DG	A	4	8.521	-2.593	18.424	1.00	42.15	A	O
ATOM	68	N9	DG	A	4	7.374	-1.309	16.870	1.00	36.73	A	N
ATOM	69	C4	DG	A	4	6.105	-0.797	17.026	1.00	34.90	A	C
ATOM	70	C5	DG	A	4	6.163	0.478	16.531	1.00	30.05	A	C
ATOM	71	N7	DG	A	4	7.443	0.774	16.090	1.00	28.84	A	N
ATOM	72	C8	DG	A	4	8.112	-0.330	16.268	1.00	29.03	A	C
ATOM	73	N3	DG	A	4	5.045	-1.453	17.545	1.00	36.92	A	N
ATOM	74	C2	DG	A	4	3.945	-0.719	17.515	1.00	30.57	A	C
ATOM	75	N2	DG	A	4	2.801	-1.233	17.985	1.00	36.63	A	N
ATOM	76	N1	DG	A	4	3.895	0.572	17.062	1.00	31.16	A	N
ATOM	77	C6	DG	A	4	4.976	1.275	16.543	1.00	32.62	A	C
ATOM	78	O6	DG	A	4	4.833	2.456	16.205	1.00	30.36	A	O
ATOM	79	OP2	DG	A	4	12.137	0.267	16.340	1.00	44.37	A	O
ATOM	80	P	DA	A	5	8.920	-6.401	15.858	1.00	64.69	A	P
ATOM	81	OP1	DA	A	5	9.442	-7.754	16.152	1.00	68.70	A	O
ATOM	82	OP2	DA	A	5	9.398	-5.627	14.674	1.00	58.00	A	O
ATOM	83	O5'	DA	A	5	7.342	-6.370	15.661	1.00	48.79	A	O
ATOM	84	C5'	DA	A	5	6.480	-6.477	16.794	1.00	48.18	A	C
ATOM	85	C4'	DA	A	5	5.063	-6.253	16.341	1.00	44.71	A	C
ATOM	86	C3'	DA	A	5	4.685	-6.980	15.047	1.00	45.71	A	C
ATOM	87	O3'	DA	A	5	3.415	-7.585	15.263	1.00	48.22	A	O
ATOM	88	C2'	DA	A	5	4.628	-5.870	14.009	1.00	43.09	A	C
ATOM	89	C1'	DA	A	5	4.231	-4.677	14.844	1.00	34.40	A	C
ATOM	90	O4'	DA	A	5	4.888	-4.847	16.078	1.00	40.05	A	O
ATOM	91	N9	DA	A	5	4.663	-3.396	14.313	1.00	28.61	A	N
ATOM	92	C4	DA	A	5	3.851	-2.294	14.288	1.00	27.73	A	C
ATOM	93	C5	DA	A	5	4.613	-1.274	13.753	1.00	27.16	A	C
ATOM	94	N7	DA	A	5	5.901	-1.711	13.488	1.00	26.97	A	N
ATOM	95	C8	DA	A	5	5.878	-2.978	13.838	1.00	26.54	A	C
ATOM	96	N3	DA	A	5	2.556	-2.238	14.635	1.00	28.73	A	N
ATOM	97	C2	DA	A	5	2.064	-1.022	14.403	1.00	28.64	A	C
ATOM	98	N1	DA	A	5	2.682	0.062	13.934	1.00	28.07	A	N
ATOM	99	C6	DA	A	5	3.990	-0.024	13.612	1.00	26.54	A	C
ATOM	100	N6	DA	A	5	4.589	1.046	13.097	1.00	25.01	A	N
ATOM	101	P	DA	A	6	2.703	-8.413	14.078	1.00	49.41	A	P
ATOM	102	OP1	DA	A	6	2.130	-9.628	14.710	1.00	47.72	A	O
ATOM	103	OP2	DA	A	6	3.629	-8.506	12.958	1.00	42.25	A	O
ATOM	104	O5'	DA	A	6	1.509	-7.450	13.707	1.00	42.99	A	O
ATOM	105	C5'	DA	A	6	0.561	-7.126	14.713	1.00	38.95	A	C
ATOM	106	C4'	DA	A	6	-0.488	-6.219	14.131	1.00	38.25	A	C
ATOM	107	C3'	DA	A	6	-1.151	-6.805	12.893	1.00	36.76	A	C
ATOM	108	O3'	DA	A	6	-2.548	-6.616	13.142	1.00	40.26	A	O
ATOM	109	C2'	DA	A	6	-0.481	-6.053	11.753	1.00	35.95	A	C
ATOM	110	C1'	DA	A	6	-0.202	-4.684	12.375	1.00	35.61	A	C
ATOM	111	O4'	DA	A	6	0.076	-4.941	13.740	1.00	39.54	A	O
ATOM	112	N9	DA	A	6	0.951	-3.969	11.827	1.00	30.39	A	N
ATOM	113	C4	DA	A	6	1.024	-2.630	11.523	1.00	27.08	A	C
ATOM	114	C5	DA	A	6	2.314	-2.419	11.080	1.00	25.98	A	C
ATOM	115	N7	DA	A	6	3.060	-3.588	11.161	1.00	27.17	A	N

ATOM	116	C8	DA	A	6	2.223	-4.459	11.665	1.00	27.03	A	C
ATOM	117	N3	DA	A	6	0.012	-1.741	11.508	1.00	29.47	A	N
ATOM	118	C2	DA	A	6	0.416	-0.573	11.015	1.00	30.01	A	C
ATOM	119	N1	DA	A	6	1.641	-0.202	10.614	1.00	27.91	A	N
ATOM	120	C6	DA	A	6	2.633	-1.124	10.640	1.00	26.25	A	C
ATOM	121	N6	DA	A	6	3.846	-0.764	10.235	1.00	26.47	A	N
ATOM	122	P	DT	A	7	-3.675	-7.057	12.036	1.00	46.76	A	P
ATOM	123	OP1	DT	A	7	-4.956	-7.270	12.751	1.00	48.13	A	O
ATOM	124	O5'	DT	A	7	-3.824	-5.738	11.162	1.00	37.89	A	O
ATOM	125	C5'	DT	A	7	-4.333	-4.544	11.722	1.00	35.27	A	C
ATOM	126	C4'	DT	A	7	-4.242	-3.447	10.693	1.00	31.30	A	C
ATOM	127	C3'	DT	A	7	-4.928	-3.792	9.369	1.00	32.43	A	C
ATOM	128	O3'	DT	A	7	-6.028	-2.904	9.254	1.00	32.90	A	O
ATOM	129	C2'	DT	A	7	-3.857	-3.555	8.302	1.00	35.76	A	C
ATOM	130	C1'	DT	A	7	-2.834	-2.713	9.044	1.00	33.09	A	C
ATOM	131	O4'	DT	A	7	-2.867	-3.181	10.361	1.00	30.26	A	O
ATOM	132	N1	DT	A	7	-1.449	-2.799	8.590	1.00	29.02	A	N
ATOM	133	C6	DT	A	7	-0.777	-3.999	8.592	1.00	31.24	A	C
ATOM	134	C5	DT	A	7	0.496	-4.132	8.208	1.00	29.65	A	C
ATOM	135	C7	DT	A	7	1.213	-5.444	8.221	1.00	34.02	A	C
ATOM	136	C4	DT	A	7	1.199	-2.951	7.767	1.00	29.11	A	C
ATOM	137	N3	DT	A	7	0.477	-1.778	7.830	1.00	27.76	A	N
ATOM	138	C2	DT	A	7	-0.842	-1.639	8.188	1.00	29.00	A	C
ATOM	139	O2	DT	A	7	-1.419	-0.565	8.159	1.00	26.02	A	O
ATOM	140	O4	DT	A	7	2.364	-2.925	7.416	1.00	27.37	A	O
ATOM	141	OP2	DT	A	7	-3.073	-8.059	11.105	1.00	43.54	A	O
ATOM	142	P	DT	A	8	-6.995	-3.006	7.972	1.00	40.98	A	P
ATOM	143	OP1	DT	A	8	-8.370	-2.749	8.444	1.00	46.45	A	O
ATOM	144	O5'	DT	A	8	-6.500	-1.771	7.107	1.00	34.55	A	O
ATOM	145	C5'	DT	A	8	-6.228	-0.532	7.741	1.00	32.48	A	C
ATOM	146	C4'	DT	A	8	-5.556	0.445	6.809	1.00	31.80	A	C
ATOM	147	C3'	DT	A	8	-6.173	0.561	5.413	1.00	35.67	A	C
ATOM	148	O3'	DT	A	8	-6.408	1.935	5.163	1.00	37.98	A	O
ATOM	149	C2'	DT	A	8	-5.120	-0.020	4.479	1.00	34.16	A	C
ATOM	150	C1'	DT	A	8	-3.826	0.249	5.254	1.00	32.91	A	C
ATOM	151	O4'	DT	A	8	-4.173	0.068	6.620	1.00	30.98	A	O
ATOM	152	N1	DT	A	8	-2.716	-0.648	4.999	1.00	28.55	A	N
ATOM	153	C6	DT	A	8	-2.911	-1.998	5.183	1.00	26.93	A	C
ATOM	154	C5	DT	A	8	-1.958	-2.912	5.029	1.00	28.09	A	C
ATOM	155	C7	DT	A	8	-2.209	-4.375	5.222	1.00	34.22	A	C
ATOM	156	C4	DT	A	8	-0.631	-2.456	4.640	1.00	28.62	A	C
ATOM	157	N3	DT	A	8	-0.501	-1.091	4.494	1.00	25.97	A	N
ATOM	158	C2	DT	A	8	-1.482	-0.143	4.636	1.00	28.11	A	C
ATOM	159	O2	DT	A	8	-1.279	1.042	4.419	1.00	28.16	A	O
ATOM	160	O4	DT	A	8	0.355	-3.169	4.523	1.00	27.94	A	O
ATOM	161	OP2	DT	A	8	-6.664	-4.231	7.217	1.00	36.27	A	O
HETATM	162	P	5fC	A	9	-7.040	2.426	3.783	1.00	46.65	A	P
HETATM	163	OP1	5fC	A	9	-7.703	3.698	4.094	1.00	48.80	A	O
HETATM	164	O5'	5fC	A	9	-5.760	2.817	2.928	1.00	41.00	A	O
HETATM	165	C5'	5fC	A	9	-4.954	3.937	3.361	1.00	37.44	A	C
HETATM	166	C4'	5fC	A	9	-3.802	4.108	2.403	1.00	37.18	A	C
HETATM	167	C3'	5fC	A	9	-4.231	4.327	0.951	1.00	38.78	A	C
HETATM	168	O3'	5fC	A	9	-3.781	5.658	0.760	1.00	44.04	A	O
HETATM	169	C2'	5fC	A	9	-3.539	3.216	0.159	1.00	35.05	A	C
HETATM	170	C1'	5fC	A	9	-2.482	2.683	1.108	1.00	32.92	A	C
HETATM	171	O4'	5fC	A	9	-2.988	2.924	2.415	1.00	34.45	A	O

HETATM	172	N1	5fC	A	9	-2.122	1.261	1.061	1.00	32.89	A	N
HETATM	173	C6	5fC	A	9	-3.042	0.288	1.336	1.00	33.23	A	C
HETATM	174	C5A	5fC	A	9	-3.729	-1.976	1.678	1.00	34.45	A	C
HETATM	175	O5A	5fC	A	9	-3.537	-3.228	1.731	1.00	40.65	A	O
HETATM	176	C5	5fC	A	9	-2.686	-1.004	1.402	1.00	30.16	A	C
HETATM	177	C4	5fC	A	9	-1.312	-1.319	1.187	1.00	27.02	A	C
HETATM	178	N4	5fC	A	9	-0.893	-2.587	1.246	1.00	29.47	A	N
HETATM	179	N3	5fC	A	9	-0.406	-0.376	0.912	1.00	27.85	A	N
HETATM	180	C2	5fC	A	9	-0.779	0.926	0.866	1.00	29.08	A	C
HETATM	181	O2	5fC	A	9	0.017	1.823	0.584	1.00	29.99	A	O
HETATM	182	OP2	5fC	A	9	-7.746	1.286	3.154	1.00	46.31	A	O
ATOM	183	P	DG	A	10	-4.099	6.435	-0.551	1.00	53.68	A	P
ATOM	184	OP1	DG	A	10	-3.680	7.859	-0.329	1.00	56.56	A	O
ATOM	185	O5'	DG	A	10	-3.052	5.863	-1.605	1.00	48.51	A	O
ATOM	186	C5'	DG	A	10	-1.681	6.252	-1.555	1.00	44.58	A	C
ATOM	187	C4'	DG	A	10	-0.908	5.465	-2.590	1.00	37.26	A	C
ATOM	188	C3'	DG	A	10	-1.520	5.524	-3.990	1.00	35.18	A	C
ATOM	189	O3'	DG	A	10	-0.450	5.472	-4.930	1.00	41.27	A	O
ATOM	190	C2'	DG	A	10	-2.327	4.254	-4.033	1.00	34.05	A	C
ATOM	191	C1'	DG	A	10	-1.350	3.345	-3.394	1.00	30.38	A	C
ATOM	192	O4'	DG	A	10	-0.939	4.066	-2.223	1.00	33.85	A	O
ATOM	193	N9	DG	A	10	-1.724	1.985	-3.009	1.00	31.02	A	N
ATOM	194	C4	DG	A	10	-0.823	0.958	-2.854	1.00	27.56	A	C
ATOM	195	C5	DG	A	10	-1.572	-0.135	-2.483	1.00	27.59	A	C
ATOM	196	N7	DG	A	10	-2.910	0.217	-2.326	1.00	28.63	A	N
ATOM	197	C8	DG	A	10	-2.956	1.476	-2.673	1.00	29.82	A	C
ATOM	198	N3	DG	A	10	0.501	1.036	-3.089	1.00	26.34	A	N
ATOM	199	C2	DG	A	10	1.115	-0.110	-2.867	1.00	25.50	A	C
ATOM	200	N2	DG	A	10	2.430	-0.189	-3.066	1.00	25.55	A	N
ATOM	201	N1	DG	A	10	0.468	-1.266	-2.480	1.00	28.74	A	N
ATOM	202	C6	DG	A	10	-0.908	-1.374	-2.260	1.00	26.64	A	C
ATOM	203	O6	DG	A	10	-1.389	-2.454	-1.892	1.00	27.40	A	O
ATOM	204	OP2	DG	A	10	-5.417	6.007	-1.059	1.00	46.61	A	O
ATOM	205	P	DC	A	11	-0.005	6.780	-5.749	1.00	47.60	A	P
ATOM	206	OP1	DC	A	11	-0.140	7.939	-4.826	1.00	49.58	A	O
ATOM	207	O5'	DC	A	11	1.542	6.552	-5.988	1.00	39.08	A	O
ATOM	208	C5'	DC	A	11	2.461	6.429	-4.899	1.00	40.60	A	C
ATOM	209	C4'	DC	A	11	3.584	5.492	-5.279	1.00	42.97	A	C
ATOM	210	C3'	DC	A	11	4.142	5.651	-6.691	1.00	38.22	A	C
ATOM	211	O3'	DC	A	11	5.571	5.550	-6.612	1.00	43.05	A	O
ATOM	212	C2'	DC	A	11	3.512	4.507	-7.460	1.00	36.70	A	C
ATOM	213	C1'	DC	A	11	3.342	3.429	-6.403	1.00	36.16	A	C
ATOM	214	O4'	DC	A	11	3.145	4.122	-5.179	1.00	35.36	A	O
ATOM	215	N1	DC	A	11	2.191	2.543	-6.538	1.00	34.57	A	N
ATOM	216	C6	DC	A	11	0.933	3.061	-6.656	1.00	34.02	A	C
ATOM	217	C5	DC	A	11	-0.141	2.269	-6.624	1.00	32.64	A	C
ATOM	218	C4	DC	A	11	0.071	0.885	-6.369	1.00	32.57	A	C
ATOM	219	N4	DC	A	11	-0.963	0.051	-6.294	1.00	31.31	A	N
ATOM	220	N3	DC	A	11	1.294	0.374	-6.228	1.00	29.10	A	N
ATOM	221	C2	DC	A	11	2.366	1.194	-6.226	1.00	30.81	A	C
ATOM	222	O2	DC	A	11	3.510	0.769	-6.041	1.00	29.32	A	O
ATOM	223	OP2	DC	A	11	-0.685	6.733	-7.085	1.00	47.16	A	O
ATOM	224	P	DG	A	12	6.494	5.956	-7.860	1.00	46.38	A	P
ATOM	225	OP1	DG	A	12	7.893	6.126	-7.365	1.00	48.60	A	O
ATOM	226	O5'	DG	A	12	6.420	4.678	-8.818	1.00	46.73	A	O
ATOM	227	C5'	DG	A	12	7.268	3.517	-8.601	1.00	44.27	A	C

ATOM	228	C4'	DG	A	12	6.852	2.399	-9.527	1.00	39.03	A	C
ATOM	229	C3'	DG	A	12	6.779	2.780	-11.005	1.00	40.02	A	C
ATOM	230	O3'	DG	A	12	8.019	2.512	-11.636	1.00	39.96	A	O
ATOM	231	C2'	DG	A	12	5.662	1.914	-11.559	1.00	40.49	A	C
ATOM	232	C1'	DG	A	12	5.043	1.263	-10.317	1.00	39.93	A	C
ATOM	233	O4'	DG	A	12	5.523	1.998	-9.198	1.00	38.45	A	O
ATOM	234	N9	DG	A	12	3.590	1.272	-10.246	1.00	33.33	A	N
ATOM	235	C4	DG	A	12	2.805	0.223	-9.849	1.00	29.93	A	C
ATOM	236	C5	DG	A	12	1.513	0.708	-9.856	1.00	34.72	A	C
ATOM	237	N7	DG	A	12	1.493	2.055	-10.195	1.00	35.39	A	N
ATOM	238	C8	DG	A	12	2.747	2.354	-10.397	1.00	38.74	A	C
ATOM	239	N3	DG	A	12	3.237	-1.024	-9.558	1.00	27.18	A	N
ATOM	240	C2	DG	A	12	2.230	-1.844	-9.290	1.00	29.13	A	C
ATOM	241	N2	DG	A	12	2.475	-3.139	-9.034	1.00	30.35	A	N
ATOM	242	N1	DG	A	12	0.910	-1.461	-9.283	1.00	28.64	A	N
ATOM	243	C6	DG	A	12	0.447	-0.178	-9.546	1.00	30.47	A	C
ATOM	244	O6	DG	A	12	-0.763	0.071	-9.464	1.00	33.53	A	O
ATOM	245	OP2	DG	A	12	5.803	7.035	-8.586	1.00	50.37	A	O
TER	246		DG	A	12							
ATOM	246	O5'	DC	B	13	-3.942	-8.932	-9.675	1.00	60.68	B	O
ATOM	247	C5'	DC	B	13	-2.729	-9.569	-10.124	1.00	54.83	B	C
ATOM	248	C4'	DC	B	13	-1.569	-9.016	-9.328	1.00	51.71	B	C
ATOM	249	C3'	DC	B	13	-1.699	-9.163	-7.811	1.00	50.71	B	C
ATOM	250	O3'	DC	B	13	-0.454	-9.373	-7.174	1.00	55.06	B	O
ATOM	251	C2'	DC	B	13	-2.033	-7.765	-7.348	1.00	52.73	B	C
ATOM	252	C1'	DC	B	13	-1.231	-6.959	-8.340	1.00	45.42	B	C
ATOM	253	O4'	DC	B	13	-1.468	-7.595	-9.576	1.00	47.77	B	O
ATOM	254	N1	DC	B	13	-1.657	-5.569	-8.454	1.00	40.50	B	N
ATOM	255	C6	DC	B	13	-2.976	-5.243	-8.328	1.00	44.08	B	C
ATOM	256	C5	DC	B	13	-3.381	-3.973	-8.398	1.00	47.50	B	C
ATOM	257	C4	DC	B	13	-2.383	-2.977	-8.582	1.00	42.21	B	C
ATOM	258	N4	DC	B	13	-2.733	-1.699	-8.683	1.00	43.64	B	N
ATOM	259	N3	DC	B	13	-1.094	-3.289	-8.735	1.00	34.79	B	N
ATOM	260	C2	DC	B	13	-0.698	-4.570	-8.645	1.00	31.99	B	C
ATOM	261	O2	DC	B	13	0.497	-4.891	-8.719	1.00	35.45	B	O
ATOM	262	P	DG	B	14	0.158	-10.839	-7.027	1.00	61.99	B	P
ATOM	263	OP1	DG	B	14	0.415	-11.402	-8.368	1.00	51.57	B	O
ATOM	264	O5'	DG	B	14	1.630	-10.517	-6.538	1.00	53.67	B	O
ATOM	265	C5'	DG	B	14	2.532	-9.990	-7.520	1.00	48.43	B	C
ATOM	266	C4'	DG	B	14	3.541	-9.084	-6.868	1.00	43.78	B	C
ATOM	267	C3'	DG	B	14	3.853	-9.428	-5.416	1.00	40.04	B	C
ATOM	268	O3'	DG	B	14	5.259	-9.425	-5.278	1.00	43.30	B	O
ATOM	269	C2'	DG	B	14	3.206	-8.320	-4.610	1.00	41.74	B	C
ATOM	270	C1'	DG	B	14	3.194	-7.165	-5.582	1.00	43.03	B	C
ATOM	271	O4'	DG	B	14	3.021	-7.745	-6.864	1.00	41.00	B	O
ATOM	272	N9	DG	B	14	2.078	-6.259	-5.376	1.00	35.89	B	N
ATOM	273	C4	DG	B	14	2.091	-4.899	-5.570	1.00	33.28	B	C
ATOM	274	C5	DG	B	14	0.802	-4.485	-5.326	1.00	33.87	B	C
ATOM	275	N7	DG	B	14	-0.008	-5.563	-5.000	1.00	32.49	B	N
ATOM	276	C8	DG	B	14	0.783	-6.596	-5.076	1.00	35.17	B	C
ATOM	277	N3	DG	B	14	3.174	-4.153	-5.879	1.00	32.12	B	N
ATOM	278	C2	DG	B	14	2.882	-2.863	-5.927	1.00	30.96	B	C
ATOM	279	N2	DG	B	14	3.837	-1.973	-6.211	1.00	32.19	B	N
ATOM	280	N1	DG	B	14	1.634	-2.354	-5.698	1.00	25.52	B	N
ATOM	281	C6	DG	B	14	0.509	-3.099	-5.373	1.00	30.41	B	C
ATOM	282	O6	DG	B	14	-0.587	-2.529	-5.208	1.00	31.80	B	O

ATOM	283	OP2	DG	B	14	-0.608	-11.574	-5.988	1.00	65.24	B	O
ATOM	284	P	DC	B	15	5.895	-9.786	-3.878	1.00	47.66	B	P
ATOM	285	OP1	DC	B	15	7.083	-10.582	-4.175	1.00	45.77	B	O
ATOM	286	O5'	DC	B	15	6.352	-8.357	-3.345	1.00	38.19	B	O
ATOM	287	C5'	DC	B	15	7.265	-7.568	-4.121	1.00	36.87	B	C
ATOM	288	C4'	DC	B	15	7.129	-6.128	-3.706	1.00	32.10	B	C
ATOM	289	C3'	DC	B	15	7.394	-5.931	-2.225	1.00	32.83	B	C
ATOM	290	O3'	DC	B	15	8.727	-5.527	-2.135	1.00	33.30	B	O
ATOM	291	C2'	DC	B	15	6.503	-4.770	-1.844	1.00	36.15	B	C
ATOM	292	C1'	DC	B	15	5.503	-4.652	-2.974	1.00	33.11	B	C
ATOM	293	O4'	DC	B	15	5.773	-5.703	-3.895	1.00	34.52	B	O
ATOM	294	N1	DC	B	15	4.118	-4.776	-2.546	1.00	28.61	B	N
ATOM	295	C6	DC	B	15	3.615	-5.979	-2.141	1.00	32.82	B	C
ATOM	296	C5	DC	B	15	2.333	-6.105	-1.783	1.00	32.74	B	C
ATOM	297	C4	DC	B	15	1.535	-4.923	-1.781	1.00	31.98	B	C
ATOM	298	N4	DC	B	15	0.251	-4.983	-1.426	1.00	33.10	B	N
ATOM	299	N3	DC	B	15	2.033	-3.736	-2.138	1.00	27.49	B	N
ATOM	300	C2	DC	B	15	3.329	-3.626	-2.486	1.00	25.72	B	C
ATOM	301	O2	DC	B	15	3.814	-2.552	-2.866	1.00	26.37	B	O
ATOM	302	OP2	DC	B	15	4.825	-10.254	-2.978	1.00	46.52	B	O
ATOM	303	P	DG	B	16	9.438	-5.339	-0.751	1.00	35.28	B	P
ATOM	304	OP1	DG	B	16	10.835	-5.682	-0.951	1.00	43.92	B	O
ATOM	305	O5'	DG	B	16	9.265	-3.779	-0.498	1.00	33.04	B	O
ATOM	306	C5'	DG	B	16	9.931	-2.839	-1.302	1.00	30.23	B	C
ATOM	307	C4'	DG	B	16	9.400	-1.467	-0.991	1.00	30.82	B	C
ATOM	308	C3'	DG	B	16	9.796	-1.009	0.403	1.00	30.57	B	C
ATOM	309	O3'	DG	B	16	10.181	0.363	0.256	1.00	33.66	B	O
ATOM	310	C2'	DG	B	16	8.583	-1.370	1.223	1.00	30.16	B	C
ATOM	311	C1'	DG	B	16	7.452	-1.147	0.247	1.00	27.65	B	C
ATOM	312	O4'	DG	B	16	7.943	-1.501	-1.015	1.00	28.73	B	O
ATOM	313	N9	DG	B	16	6.243	-1.902	0.502	1.00	25.37	B	N
ATOM	314	C4	DG	B	16	4.970	-1.405	0.395	1.00	26.60	B	C
ATOM	315	C5	DG	B	16	4.144	-2.429	0.790	1.00	26.09	B	C
ATOM	316	N7	DG	B	16	4.881	-3.550	1.145	1.00	26.78	B	N
ATOM	317	C8	DG	B	16	6.117	-3.203	0.931	1.00	24.67	B	C
ATOM	318	N3	DG	B	16	4.637	-0.163	-0.016	1.00	29.64	B	N
ATOM	319	C2	DG	B	16	3.339	0.046	0.065	1.00	25.65	B	C
ATOM	320	N2	DG	B	16	2.841	1.218	-0.295	1.00	26.47	B	N
ATOM	321	N1	DG	B	16	2.428	-0.910	0.436	1.00	26.79	B	N
ATOM	322	C6	DG	B	16	2.751	-2.197	0.840	1.00	27.95	B	C
ATOM	323	O6	DG	B	16	1.866	-2.969	1.164	1.00	27.91	B	O
ATOM	324	OP2	DG	B	16	8.669	-6.002	0.297	1.00	39.64	B	O
ATOM	325	P	DA	B	17	10.234	1.424	1.446	1.00	38.13	B	P
ATOM	326	OP1	DA	B	17	11.183	2.496	1.064	1.00	37.61	B	O
ATOM	327	OP2	DA	B	17	10.339	0.727	2.754	1.00	33.58	B	O
ATOM	328	O5'	DA	B	17	8.805	2.135	1.373	1.00	34.32	B	O
ATOM	329	C5'	DA	B	17	8.384	2.861	0.191	1.00	33.02	B	C
ATOM	330	C4'	DA	B	17	7.231	3.758	0.549	1.00	31.13	B	C
ATOM	331	C3'	DA	B	17	7.468	4.649	1.766	1.00	35.18	B	C
ATOM	332	O3'	DA	B	17	6.770	5.856	1.499	1.00	39.66	B	O
ATOM	333	C2'	DA	B	17	6.786	3.905	2.901	1.00	33.80	B	C
ATOM	334	C1'	DA	B	17	5.651	3.181	2.213	1.00	30.96	B	C
ATOM	335	O4'	DA	B	17	6.077	2.951	0.876	1.00	31.53	B	O
ATOM	336	N9	DA	B	17	5.324	1.875	2.773	1.00	28.32	B	N
ATOM	337	C4	DA	B	17	4.072	1.380	3.037	1.00	28.30	B	C
ATOM	338	C5	DA	B	17	4.266	0.085	3.485	1.00	27.61	B	C

ATOM	339	N7	DA	B	17	5.612	-0.249	3.450	1.00	26.12	B	N
ATOM	340	C8	DA	B	17	6.195	0.847	3.028	1.00	26.53	B	C
ATOM	341	N3	DA	B	17	2.902	2.032	2.941	1.00	27.16	B	N
ATOM	342	C2	DA	B	17	1.899	1.254	3.342	1.00	27.23	B	C
ATOM	343	N1	DA	B	17	1.934	-0.019	3.779	1.00	27.91	B	N
ATOM	344	C6	DA	B	17	3.125	-0.651	3.825	1.00	24.10	B	C
ATOM	345	N6	DA	B	17	3.166	-1.906	4.269	1.00	27.66	B	N
ATOM	346	P	DA	B	18	6.778	7.058	2.537	1.00	38.20	B	P
ATOM	347	OP1	DA	B	18	6.645	8.308	1.721	1.00	43.32	B	O
ATOM	348	OP2	DA	B	18	7.855	6.823	3.538	1.00	41.19	B	O
ATOM	349	O5'	DA	B	18	5.373	6.914	3.234	1.00	32.27	B	O
ATOM	350	C5'	DA	B	18	4.210	6.789	2.412	1.00	36.14	B	C
ATOM	351	C4'	DA	B	18	3.009	6.675	3.311	1.00	34.86	B	C
ATOM	352	C3'	DA	B	18	3.032	7.587	4.538	1.00	36.89	B	C
ATOM	353	O3'	DA	B	18	1.779	8.296	4.506	1.00	42.86	B	O
ATOM	354	C2'	DA	B	18	3.231	6.625	5.702	1.00	32.51	B	C
ATOM	355	C1'	DA	B	18	2.673	5.311	5.175	1.00	32.89	B	C
ATOM	356	O4'	DA	B	18	2.963	5.323	3.807	1.00	32.09	B	O
ATOM	357	N9	DA	B	18	3.244	4.077	5.700	1.00	29.18	B	N
ATOM	358	C4	DA	B	18	2.559	2.937	6.064	1.00	30.18	B	C
ATOM	359	C5	DA	B	18	3.526	2.020	6.425	1.00	28.20	B	C
ATOM	360	N7	DA	B	18	4.795	2.534	6.210	1.00	27.86	B	N
ATOM	361	C8	DA	B	18	4.573	3.748	5.767	1.00	29.38	B	C
ATOM	362	N3	DA	B	18	1.224	2.772	6.150	1.00	27.11	B	N
ATOM	363	C2	DA	B	18	0.926	1.555	6.607	1.00	28.91	B	C
ATOM	364	N1	DA	B	18	1.752	0.571	6.992	1.00	27.71	B	N
ATOM	365	C6	DA	B	18	3.082	0.759	6.863	1.00	27.13	B	C
ATOM	366	N6	DA	B	18	3.906	-0.231	7.198	1.00	25.58	B	N
ATOM	367	P	DT	B	19	1.354	9.281	5.750	1.00	46.96	B	P
ATOM	368	OP1	DT	B	19	0.586	10.374	5.187	1.00	59.97	B	O
ATOM	369	O5'	DT	B	19	0.397	8.373	6.659	1.00	42.24	B	O
ATOM	370	C5'	DT	B	19	-0.683	7.564	6.172	1.00	36.68	B	C
ATOM	371	C4'	DT	B	19	-1.136	6.605	7.254	1.00	36.07	B	C
ATOM	372	C3'	DT	B	19	-1.392	7.236	8.631	1.00	34.72	B	C
ATOM	373	O3'	DT	B	19	-2.781	7.079	8.891	1.00	38.41	B	O
ATOM	374	C2'	DT	B	19	-0.522	6.437	9.588	1.00	35.10	B	C
ATOM	375	C1'	DT	B	19	-0.285	5.138	8.810	1.00	32.49	B	C
ATOM	376	O4'	DT	B	19	-0.138	5.588	7.478	1.00	32.49	B	O
ATOM	377	N1	DT	B	19	0.929	4.370	9.164	1.00	30.17	B	N
ATOM	378	C6	DT	B	19	2.157	4.978	9.025	1.00	29.58	B	C
ATOM	379	C5	DT	B	19	3.323	4.363	9.254	1.00	29.57	B	C
ATOM	380	C7	DT	B	19	4.646	5.036	9.074	1.00	33.09	B	C
ATOM	381	C4	DT	B	19	3.293	2.978	9.664	1.00	30.05	B	C
ATOM	382	N3	DT	B	19	2.022	2.430	9.805	1.00	27.48	B	N
ATOM	383	C2	DT	B	19	0.823	3.045	9.537	1.00	28.01	B	C
ATOM	384	O2	DT	B	19	-0.245	2.475	9.666	1.00	30.76	B	O
ATOM	385	O4	DT	B	19	4.288	2.318	9.958	1.00	27.38	B	O
ATOM	386	OP2	DT	B	19	2.562	9.524	6.601	1.00	46.52	B	O
ATOM	387	P	DT	B	20	-3.425	7.545	10.310	1.00	43.75	B	P
ATOM	388	OP1	DT	B	20	-4.785	8.037	9.992	1.00	56.62	B	O
ATOM	389	O5'	DT	B	20	-3.569	6.173	11.090	1.00	37.85	B	O
ATOM	390	C5'	DT	B	20	-4.267	5.115	10.469	1.00	34.81	B	C
ATOM	391	C4'	DT	B	20	-4.106	3.895	11.331	1.00	38.13	B	C
ATOM	392	C3'	DT	B	20	-4.581	4.176	12.750	1.00	36.83	B	C
ATOM	393	O3'	DT	B	20	-5.444	3.092	13.028	1.00	40.63	B	O
ATOM	394	C2'	DT	B	20	-3.301	4.222	13.568	1.00	32.79	B	C

ATOM	395	C1'	DT	B	20	-2.440	3.233	12.801	1.00	35.72	B	C
ATOM	396	O4'	DT	B	20	-2.710	3.522	11.436	1.00	34.54	B	O
ATOM	397	N1	DT	B	20	-0.990	3.317	12.967	1.00	31.05	B	N
ATOM	398	C6	DT	B	20	-0.373	4.523	12.722	1.00	30.10	B	C
ATOM	399	C5	DT	B	20	0.952	4.690	12.736	1.00	32.95	B	C
ATOM	400	C7	DT	B	20	1.607	6.008	12.473	1.00	33.96	B	C
ATOM	401	C4	DT	B	20	1.781	3.523	12.983	1.00	29.71	B	C
ATOM	402	N3	DT	B	20	1.093	2.349	13.210	1.00	29.85	B	N
ATOM	403	C2	DT	B	20	-0.266	2.174	13.209	1.00	28.53	B	C
ATOM	404	O2	DT	B	20	-0.782	1.098	13.450	1.00	30.71	B	O
ATOM	405	O4	DT	B	20	3.003	3.527	13.003	1.00	29.18	B	O
ATOM	406	OP2	DT	B	20	-2.417	8.367	11.028	1.00	42.86	B	O
HETATM	407	P	5fC	B	21	-6.397	3.156	14.316	1.00	45.81	B	P
HETATM	408	OP1	5fC	B	21	-7.641	2.441	13.949	1.00	53.55	B	O
HETATM	409	O5'	5fC	B	21	-5.621	2.260	15.383	1.00	38.52	B	O
HETATM	410	C5'	5fC	B	21	-5.305	0.887	15.106	1.00	39.92	B	C
HETATM	411	C4'	5fC	B	21	-4.219	0.396	16.028	1.00	38.57	B	C
HETATM	412	C3'	5fC	B	21	-4.499	0.629	17.518	1.00	37.82	B	C
HETATM	413	O3'	5fC	B	21	-4.904	-0.636	18.058	1.00	42.45	B	O
HETATM	414	C2'	5fC	B	21	-3.181	1.147	18.091	1.00	41.97	B	C
HETATM	415	C1'	5fC	B	21	-2.196	0.881	16.975	1.00	39.77	B	C
HETATM	416	O4'	5fC	B	21	-2.950	1.029	15.764	1.00	35.58	B	O
HETATM	417	N1	5fC	B	21	-1.046	1.765	16.879	1.00	35.23	B	N
HETATM	418	C6	5fC	B	21	-1.196	3.086	16.567	1.00	33.99	B	C
HETATM	419	C5A	5fC	B	21	-0.352	5.299	16.088	1.00	40.73	B	C
HETATM	420	O5A	5fC	B	21	0.557	6.134	15.970	1.00	46.68	B	O
HETATM	421	C5	5fC	B	21	-0.130	3.874	16.374	1.00	35.86	B	C
HETATM	422	C4	5fC	B	21	1.157	3.267	16.467	1.00	28.43	B	C
HETATM	423	N4	5fC	B	21	2.253	3.995	16.299	1.00	32.91	B	N
HETATM	424	N3	5fC	B	21	1.310	1.984	16.786	1.00	30.81	B	N
HETATM	425	C2	5fC	B	21	0.227	1.200	16.963	1.00	28.51	B	C
HETATM	426	O2	5fC	B	21	0.329	0.005	17.255	1.00	29.85	B	O
HETATM	427	OP2	5fC	B	21	-6.418	4.564	14.764	1.00	42.85	B	O
ATOM	428	P	DG	B	22	-5.590	-0.642	19.507	1.00	48.14	B	P
ATOM	429	OP1	DG	B	22	-6.566	-1.738	19.540	1.00	48.52	B	O
ATOM	430	O5'	DG	B	22	-4.361	-0.922	20.495	1.00	45.62	B	O
ATOM	431	C5'	DG	B	22	-3.551	-2.115	20.397	1.00	43.78	B	C
ATOM	432	C4'	DG	B	22	-2.314	-1.925	21.246	1.00	43.62	B	C
ATOM	433	C3'	DG	B	22	-2.620	-1.421	22.657	1.00	46.92	B	C
ATOM	434	O3'	DG	B	22	-1.778	-2.098	23.563	1.00	52.80	B	O
ATOM	435	C2'	DG	B	22	-2.269	0.058	22.612	1.00	43.47	B	C
ATOM	436	C1'	DG	B	22	-1.080	-0.020	21.682	1.00	37.63	B	C
ATOM	437	O4'	DG	B	22	-1.450	-0.915	20.666	1.00	36.05	B	O
ATOM	438	N9	DG	B	22	-0.584	1.200	21.066	1.00	33.86	B	N
ATOM	439	C4	DG	B	22	0.746	1.411	20.810	1.00	31.62	B	C
ATOM	440	C5	DG	B	22	0.823	2.674	20.281	1.00	35.96	B	C
ATOM	441	N7	DG	B	22	-0.438	3.262	20.225	1.00	35.29	B	N
ATOM	442	C8	DG	B	22	-1.239	2.351	20.707	1.00	32.23	B	C
ATOM	443	N3	DG	B	22	1.726	0.501	20.961	1.00	30.03	B	N
ATOM	444	C2	DG	B	22	2.910	0.991	20.638	1.00	27.87	B	C
ATOM	445	N2	DG	B	22	3.988	0.234	20.784	1.00	27.26	B	N
ATOM	446	N1	DG	B	22	3.099	2.219	20.071	1.00	29.46	B	N
ATOM	447	C6	DG	B	22	2.099	3.162	19.863	1.00	28.03	B	C
ATOM	448	O6	DG	B	22	2.381	4.253	19.367	1.00	31.42	B	O
ATOM	449	OP2	DG	B	22	-5.925	0.752	19.835	1.00	43.73	B	O
ATOM	450	P A	DC	B	23	-2.411	-2.835	24.828	0.70	63.05	B	P

ATOM	451	P	B	DC	B	23	-2.226	-2.215	25.065	0.30	57.21	B	P
ATOM	452	OP1A		DC	B	23	-2.905	-4.172	24.371	0.70	53.56	B	O
ATOM	453	OP1B		DC	B	23	-3.531	-2.928	25.082	0.30	56.27	B	O
ATOM	454	O5'		DC	B	23	-1.129	-3.177	25.709	1.00	55.63	B	O
ATOM	455	C5'		DC	B	23	-0.062	-3.886	25.031	1.00	53.67	B	C
ATOM	456	C4'		DC	B	23	1.282	-3.291	25.374	1.00	51.17	B	C
ATOM	457	C3'		DC	B	23	1.413	-2.797	26.816	1.00	47.47	B	C
ATOM	458	O3'		DC	B	23	2.262	-3.653	27.576	1.00	45.15	B	O
ATOM	459	C2'		DC	B	23	2.098	-1.453	26.691	1.00	46.78	B	C
ATOM	460	C1'		DC	B	23	2.368	-1.279	25.215	1.00	44.28	B	C
ATOM	461	O4'		DC	B	23	1.482	-2.132	24.549	1.00	46.14	B	O
ATOM	462	N1		DC	B	23	2.081	0.078	24.767	1.00	37.76	B	N
ATOM	463	C6		DC	B	23	0.809	0.571	24.805	1.00	37.20	B	C
ATOM	464	C5		DC	B	23	0.546	1.824	24.436	1.00	32.17	B	C
ATOM	465	C4		DC	B	23	1.629	2.605	23.957	1.00	31.87	B	C
ATOM	466	N4		DC	B	23	1.421	3.851	23.563	1.00	33.91	B	N
ATOM	467	N3		DC	B	23	2.880	2.136	23.927	1.00	32.45	B	N
ATOM	468	C2		DC	B	23	3.135	0.879	24.351	1.00	32.11	B	C
ATOM	469	O2		DC	B	23	4.286	0.408	24.374	1.00	32.87	B	O
ATOM	470	OP2A		DC	B	23	-3.323	-1.855	25.504	0.70	55.11	B	O
ATOM	471	OP2B		DC	B	23	-2.146	-0.861	25.675	0.30	53.52	B	O
ATOM	472	P		DG	B	24	2.435	-3.376	29.155	1.00	50.50	B	P
ATOM	473	OP1		DG	B	24	2.710	-4.660	29.813	1.00	52.60	B	O
ATOM	474	O5'		DG	B	24	3.821	-2.588	29.291	1.00	34.59	B	O
ATOM	475	C5'		DG	B	24	5.059	-3.083	28.827	1.00	32.11	B	C
ATOM	476	C4'		DG	B	24	5.991	-1.913	28.629	1.00	29.05	B	C
ATOM	477	C3'		DG	B	24	6.190	-1.013	29.843	1.00	31.74	B	C
ATOM	478	O3'		DG	B	24	7.106	-1.670	30.743	1.00	33.44	B	O
ATOM	479	C2'		DG	B	24	6.699	0.265	29.213	1.00	31.87	B	C
ATOM	480	C1'		DG	B	24	6.008	0.285	27.852	1.00	32.26	B	C
ATOM	481	O4'		DG	B	24	5.458	-1.032	27.634	1.00	32.57	B	O
ATOM	482	N9		DG	B	24	4.911	1.240	27.746	1.00	30.42	B	N
ATOM	483	C4		DG	B	24	4.965	2.488	27.160	1.00	29.08	B	C
ATOM	484	C5		DG	B	24	3.684	2.989	27.250	1.00	28.01	B	C
ATOM	485	N7		DG	B	24	2.841	2.078	27.883	1.00	28.82	B	N
ATOM	486	C8		DG	B	24	3.610	1.061	28.161	1.00	29.69	B	C
ATOM	487	N3		DG	B	24	6.070	3.084	26.664	1.00	28.49	B	N
ATOM	488	C2		DG	B	24	5.799	4.269	26.145	1.00	28.65	B	C
ATOM	489	N2		DG	B	24	6.778	4.986	25.584	1.00	30.60	B	N
ATOM	490	N1		DG	B	24	4.554	4.836	26.145	1.00	30.24	B	N
ATOM	491	C6		DG	B	24	3.415	4.272	26.707	1.00	30.60	B	C
ATOM	492	O6		DG	B	24	2.337	4.883	26.657	1.00	33.90	B	O
ATOM	493	OP2		DG	B	24	1.340	-2.476	29.617	1.00	46.76	B	O
TER	495			DG	B	24							
HETATM	494	O	HOH	D		1	-1.235	3.732	4.967	1.00	33.32		O
HETATM	495	O	HOH	D		2	7.433	1.866	6.580	1.00	35.46		O
HETATM	496	O	HOH	D		3	-3.248	3.702	7.088	1.00	31.96		O
HETATM	497	O	HOH	D		4	-0.686	5.471	3.037	1.00	33.80		O
HETATM	498	O	HOH	D		5	1.299	3.980	1.638	1.00	29.48		O
HETATM	499	O	HOH	D		6	-2.796	1.657	9.007	1.00	32.07		O
HETATM	500	O	HOH	D		7	-2.389	-1.131	12.704	1.00	34.16		O
HETATM	501	O	HOH	D		8	-4.168	0.271	10.872	1.00	32.19		O
HETATM	502	O	HOH	D		9	-5.494	5.247	6.748	1.00	41.05		O
HETATM	503	O	HOH	D		10	-7.753	6.312	3.107	0.50	34.38		O
HETATM	504	O	HOH	D		11	2.364	5.060	-0.380	1.00	41.29		O
HETATM	505	O	HOH	D		12	2.242	3.285	-2.738	1.00	28.85		O

HETATM	506	O	HOH	D	13	4.898	4.355	-1.815	0.50	25.06	O
HETATM	507	O	HOH	D	14	6.853	-0.930	7.245	1.00	38.82	O
HETATM	508	O	HOH	D	15	6.843	-2.303	4.608	1.00	38.51	O
HETATM	509	O	HOH	D	16	4.607	-4.679	7.406	0.50	28.91	O
HETATM	510	O	HOH	D	17	0.869	-5.582	4.449	1.00	34.87	O
HETATM	511	O	HOH	D	18	1.448	-5.739	1.661	1.00	43.62	O
HETATM	512	O	HOH	D	19	1.415	7.262	21.771	0.50	30.85	O
HETATM	513	O	HOH	D	21	16.991	-2.215	16.301	0.50	34.08	O
HETATM	514	O	HOH	D	22	10.557	-0.035	13.154	0.50	34.41	O
HETATM	515	O	HOH	D	23	11.050	-0.243	6.784	0.50	36.17	O
HETATM	516	O	HOH	D	24	5.586	-4.164	10.221	0.50	27.55	O
HETATM	517	O	HOH	D	25	0.292	-8.834	9.617	0.50	28.42	O
HETATM	518	O	HOH	D	26	-6.129	0.688	0.238	0.50	26.28	O
HETATM	519	O	HOH	D	27	-3.753	1.339	-6.468	1.00	40.42	O
HETATM	520	O	HOH	D	28	-3.342	-3.599	-4.471	0.50	38.93	O
HETATM	521	O	HOH	D	29	-3.410	11.131	11.508	0.50	36.07	O
HETATM	522	O	HOH	D	30	6.549	-2.220	9.846	0.50	24.58	O
HETATM	523	O	HOH	D	31	-5.264	-0.871	-5.608	0.50	37.78	O
HETATM	524	O	HOH	D	32	1.654	-2.427	21.810	0.50	30.64	O
HETATM	525	O	HOH	D	33	13.668	-4.092	15.692	0.50	37.66	O
HETATM	526	O	HOH	D	34	4.002	8.540	19.726	0.50	29.90	O
HETATM	527	O	HOH	D	35	7.435	1.468	12.226	0.50	32.35	O
HETATM	528	O	HOH	D	36	4.958	5.388	12.916	0.50	24.22	O
HETATM	529	O	HOH	D	37	7.379	4.989	12.247	0.50	31.89	O
HETATM	530	O	HOH	D	38	2.911	8.402	9.179	0.50	32.48	O
HETATM	532	O	HOH	D	40	-1.224	5.098	23.716	0.50	32.01	O
HETATM	533	O	HOH	D	41	0.575	9.124	-2.874	0.50	37.11	O
HETATM	534	O	HOH	D	42	-5.646	-3.658	4.484	0.50	33.30	O
HETATM	535	O	HOH	D	43	9.996	-9.555	-0.480	0.50	42.10	O
HETATM	536	O	HOH	D	45	6.760	6.132	6.242	0.50	24.09	O
HETATM	537	O	HOH	D	46	0.474	9.434	11.141	0.50	27.15	O
HETATM	538	O	HOH	D	47	-2.819	-5.561	-3.598	0.50	35.36	O
HETATM	539	O	HOH	D	48	-9.076	5.897	-1.749	0.50	38.31	O
HETATM	540	O	HOH	D	49	-2.498	4.972	-8.464	0.50	44.09	O
HETATM	541	O	HOH	D	50	-3.105	11.098	6.472	0.50	47.78	O
HETATM	542	O	HOH	D	51	-4.028	4.461	17.622	0.50	35.08	O
HETATM	543	O	HOH	D	52	-10.123	-1.526	20.655	0.50	46.82	O
HETATM	544	O	HOH	D	53	0.143	1.387	29.067	0.50	39.83	O
HETATM	545	O	HOH	D	54	-1.704	-10.349	11.681	0.50	29.01	O
HETATM	546	O	HOH	D	55	-6.503	-5.881	-8.818	0.50	41.85	O
HETATM	547	O	HOH	D	56	-0.821	11.591	-7.683	1.00	55.79	O

END

File A-3: Dictionary file of 5-formyl-2'-deoxycytidine.

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# --- DESCRIPTION OF MONOMERS ---
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  _chem_comp_atom.y
  _chem_comp_atom.z
5fC      P      P      P      0.000      0.001      -0.540      0.119
5fC      OP3    O      OP     -0.660      0.227      -1.987      0.516
5fC      OP1    O      OP     -0.660      0.381      0.441      1.194
5fC      "O5'"  O      O2     0.000      -1.613     -0.460      0.002
5fC      "C5'"  C      CH2    0.000      -2.412     0.695     -0.366
5fC      "H5'"  H      H      0.000      -2.004     1.206     -1.241
5fC      "H5'"  H      H      0.000      -2.500     1.405     0.459
5fC      "C4'"  C      CH1    0.000      -3.783     0.131     -0.701
5fC      "H4'"  H      H      0.000      -3.606     -0.705     -1.391
5fC      "C3'"  C      CH1    0.000      -4.862     0.991     -1.350
5fC      "H3'"  H      H      0.000      -4.664     2.061     -1.199
5fC      "O3'"  O      OH1    0.000      -4.928     0.664     -2.754
5fC      "HO3'" H      H      0.000      -5.661     1.144     -3.165
5fC      "C2'"  C      CH2    0.000      -6.185     0.558     -0.660
5fC      "H2'"  H      H      0.000      -6.865     0.104     -1.383
5fC      "H2'"  H      H      0.000      -6.673     1.415     -0.192
5fC      "C1'"  C      CH1    0.000      -5.815     -0.478     0.421
5fC      "H1'"  H      H      0.000      -6.089     -1.462     0.015
5fC      "O4'"  O      O2     0.000      -4.377     -0.452     0.488

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5fC	N1	N	NR6	0.000	-6.434	-0.388	1.735
5fC	C6	C	CR16	0.000	-7.038	-1.463	2.305
5fC	H6	H	H	0.000	-7.005	-2.406	1.774
5fC	H5C	H	H	-0.500	3.069	7.403	0.644
5fC	C5A	C	C	0.000	1.797	7.837	0.471
5fC	O5A	O	OC	-0.500	1.352	8.985	0.585
5fC	C5	C	CR16	0.000	-7.709	-1.424	3.556
5fC	H5	H	H	0.000	-8.160	-2.314	3.979
5fC	C4	C	CR6	0.000	-7.767	-0.214	4.213
5fC	N4	N	NH2	0.000	-8.312	-0.031	5.446
5fC	H42	H	H	0.000	-8.712	-0.813	5.946
5fC	H41	H	H	0.000	-8.318	0.889	5.866
5fC	N3	N	NRD6	0.000	-6.825	0.681	3.769
5fC	C2	C	CR6	0.000	-6.022	0.556	2.670
5fC	O2	O	O	0.000	-4.961	1.193	2.528
5fC	OP2	O	OP	-0.660	0.506	-0.216	-1.268

loop_

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5fC      P        "O5'"      OP2  .
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5fC      OP1      P          .      .
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5fC      "H5'"      "C5'"      .      .
5fC      "H5'"      "C5'"      .      .
5fC      "C4'"      "C5'"      "C3'"  .
5fC      "H4'"      "C4'"      .      .
5fC      "C3'"      "C4'"      "C2'"  .
5fC      "H3'"      "C3'"      .      .
5fC      "O3'"      "C3'"      "HO3'" .
5fC      "HO3'"     "O3'"      .      .
5fC      "C2'"      "C3'"      "C1'"  .
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5fC      "H2'"      "C2'"      .      .
5fC      "C1'"      "C2'"      N1     .
5fC      "H1'"      "C1'"      .      .
5fC      "O4'"      "C1'"      .      .
5fC      N1        "C1'"      C6     .
5fC      C5A      H5C      C5     .
5fC      O5A      C5A      .      .
5fC      C6        N1        C5     .
5fC      H6        C6        .      .
5fC      C5        C6        C4     .
5fC      H5        C5        .      .
5fC      C4        C5        N3     .
5fC      N4        C4        H41    .
5fC      H42      N4        .      .
5fC      H41      N4        .      .
5fC      N3        C4        C2     .
5fC      C2        N3        O2     .
5fC      O2        C2        .      END
5fC      OP2      P          .      .

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5fC      OP1    P       deloc    1.485    0.017
5fC      OP2    P       deloc    1.485    0.017
5fC      "O5'" P       single   1.593    0.010
5fC      "C5'" "O5'" single   1.440    0.016
5fC      "C4'" "C5'" single   1.511    0.008
5fC      "C4'" "O4'" single   1.446    0.011
5fC      "C3'" "C4'" single   1.528    0.010
5fC      "O4'" "C1'" single   1.420    0.013
5fC      "O3'" "C3'" single   1.431    0.013
5fC      "C2'" "C3'" single   1.518    0.010
5fC      "C1'" "C2'" single   1.521    0.014
5fC      N1     "C1'" single   1.470    0.012
5fC      N1     C2     single   1.397    0.010
5fC      C6     N1     single   1.367    0.006
5fC      O2     C2     double   1.240    0.009
5fC      C2     N3     single   1.353    0.008
5fC      N3     C4     single   1.335    0.007
5fC      N4     C4     single   1.335    0.009
5fC      C4     C5     single   1.425    0.008
5fC      C5     C5A    single   1.452    0.020
5fC      O5A    C5A    double   1.227    0.020
5fC      C5     C6     single   1.339    0.008
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5fC      "H5'" "C5'" single   1.092    0.020
5fC      "H4'" "C4'" single   1.099    0.020
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5fC      "H2'" "C2'" single   1.092    0.020
5fC      "H1'" "C1'" single   1.099    0.020
5fC      H41    N4     single   1.010    0.020
5fC      H42    N4     single   1.010    0.020
5fC      H6     C6     single   1.083    0.020
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5fC      OP3    P       "O5'" 104.000    1.900
5fC      OP1    P       "O5'" 108.100    2.900
5fC      OP3    P       OP2    108.300    3.200
5fC      OP1    P       OP2    119.600    1.500
5fC      "O5'" P       OP2    108.300    2.700

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5fc	P	"O5'"	"C5'"	120.900	1.600
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5fc	"O5'"	"C5'"	"H5' '"	109.470	3.000
5fc	"O5'"	"C5'"	"C4'"	110.200	1.400
5fc	"H5'"	"C5'"	"H5' '"	107.900	3.000
5fc	"H5'"	"C5'"	"C4'"	109.470	3.000
5fc	"H5' '"	"C5'"	"C4'"	109.470	3.000
5fc	"C5'"	"C4'"	"H4'"	108.340	3.000
5fc	"C5'"	"C4'"	"C3'"	114.700	1.500
5fc	"C5'"	"C4'"	"O4'"	109.400	1.600
5fc	"H4'"	"C4'"	"C3'"	108.340	3.000
5fc	"H4'"	"C4'"	"O4'"	109.470	3.000
5fc	"C3'"	"C4'"	"O4'"	105.600	1.000
5fc	"C4'"	"C3'"	"H3'"	108.340	3.000
5fc	"C4'"	"C3'"	"O3'"	110.300	2.200
5fc	"C4'"	"C3'"	"C2'"	103.200	1.000
5fc	"H3'"	"C3'"	"O3'"	109.470	3.000
5fc	"H3'"	"C3'"	"C2'"	108.340	3.000
5fc	"O3'"	"C3'"	"C2'"	110.600	2.700
5fc	"C3'"	"O3'"	"HO3'"	109.470	3.000
5fc	"C3'"	"C2'"	"H2' "	109.470	3.000
5fc	"C3'"	"C2'"	"H2' '"	109.470	3.000
5fc	"C3'"	"C2'"	"C1'"	102.700	1.400
5fc	"H2' "	"C2'"	"H2' '"	107.900	3.000
5fc	"H2' "	"C2'"	"C1'"	109.470	3.000
5fc	"H2' '"	"C2'"	"C1'"	109.470	3.000
5fc	"C2'"	"C1'"	"H1'"	108.340	3.000
5fc	"C2'"	"C1'"	"O4'"	106.100	1.100
5fc	"C2'"	"C1'"	N1	114.200	1.600
5fc	"H1'"	"C1'"	"O4'"	109.470	3.000
5fc	"H1'"	"C1'"	N1	109.470	3.000
5fc	"O4'"	"C1'"	N1	107.800	0.800
5fc	"C1'"	"O4'"	"C4'"	109.700	1.400
5fc	"C1'"	N1	C6	120.800	1.200
5fc	"C1'"	N1	C2	118.800	1.100
5fc	C6	N1	C2	120.300	0.400
5fc	N1	C6	H6	120.000	3.000
5fc	N1	C6	C5	121.000	0.500
5fc	H6	C6	C5	120.000	3.000
5fc	H5C	C5A	O5A	119.783	3.000
5fc	H5C	C5A	C5	114.626	3.000
5fc	O5A	C5A	C5	125.591	3.000
5fc	C5A	C5	C4	124.414	3.000
5fc	C5A	C5	C6	119.940	3.000
5fc	C6	C5	H5	120.000	3.000
5fc	C6	C5	C4	117.400	0.500
5fc	H5	C5	C4	120.000	3.000
5fc	C5	C4	N4	120.200	0.700
5fc	C5	C4	N3	121.900	0.400
5fc	N4	C4	N3	118.000	0.700
5fc	C4	N4	H42	120.000	3.000
5fc	C4	N4	H41	120.000	3.000
5fc	H42	N4	H41	120.000	3.000
5fc	C4	N3	C2	119.900	0.500
5fc	N3	C2	O2	121.900	0.700
5fc	N3	C2	N1	119.200	0.700

```

5fC      O2      C2      N1      118.900      0.600
loop_
_chem_comp_tor.comp_id
_chem_comp_tor.id
_chem_comp_tor.atom_id_1
_chem_comp_tor.atom_id_2
_chem_comp_tor.atom_id_3
_chem_comp_tor.atom_id_4
_chem_comp_tor.value_angle
_chem_comp_tor.value_angle_esd
_chem_comp_tor.period
5fC      var_1    OP2      P        "O5'" "C5'"    -60.014  20.000  1
5fC      var_2    P        "O5'" "C5'" "C4'"    179.972  20.000  1
5fC      var_3    "O5'" "C5'" "C4'" "C3'"    -179.996  20.000  3
5fC      var_4    "C5'" "C4'" "O4'" "C1'"    150.000  20.000  1
5fC      var_5    "C5'" "C4'" "C3'" "C2'"    -150.000  20.000  3
5fC      var_6    "C4'" "C3'" "O3'" "HO3'"    175.000  20.000  1
5fC      var_7    "C4'" "C3'" "C2'" "C1'"     30.000  20.000  3
5fC      var_8    "C3'" "C2'" "C1'" N1       120.000  20.000  3
5fC      var_9    "C2'" "C1'" "O4'" "C4'"    -30.000  20.000  1
5fC      var_10   "C2'" "C1'" N1      C6       120.298  20.000  1
5fC      CONST_1  "C1'" N1      C2      N3       180.000  0.000  0
5fC      CONST_2  "C1'" N1      C6      C5       180.000  0.000  0
5fC      CONST_3  N1      C6      C5      C4        0.000  0.000  0
5fC      CONST_4  C6      C5      C4      N3        0.000  0.000  0
5fC      CONST_5  C5      C4      N4      H41      180.000  0.000  0
5fC      CONST_6  C5      C4      N3      C2        0.000  0.000  0
5fC      CONST_7  C4      N3      C2      O2       180.000  0.000  0
loop_
_chem_comp_chir.comp_id
_chem_comp_chir.id
_chem_comp_chir.atom_id_centre
_chem_comp_chir.atom_id_1
_chem_comp_chir.atom_id_2
_chem_comp_chir.atom_id_3
_chem_comp_chir.volume_sign
5fC      chir_01  "C4'" "C5'" "O4'" "C3'"    negativ
5fC      chir_02  "C3'" "C4'" "O3'" "C2'"    negativ
5fC      chir_03  "C1'" "O4'" "C2'" N1       positiv
loop_
_chem_comp_plane_atom.comp_id
_chem_comp_plane_atom.plane_id
_chem_comp_plane_atom.atom_id
_chem_comp_plane_atom.dist_esd
5fC      plan-1  N1      0.020
5fC      plan-1  "C1'"  0.020
5fC      plan-1  C2      0.020
5fC      plan-1  C6      0.020
5fC      plan-1  N3      0.020
5fC      plan-1  C4      0.020
5fC      plan-1  C5      0.020
5fC      plan-1  O2      0.020
5fC      plan-1  N4      0.020
5fC      plan-1  H5      0.020
5fC      plan-1  H6      0.020
5fC      plan-1  H42     0.020

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5fC	plan-1	H41	0.020
5fC	plan-1	C5A	0.020
5fC	plan-2	N4	0.020
5fC	plan-2	C4	0.020
5fC	plan-2	H41	0.020
5fC	plan-2	H42	0.020

File A-4: PDB coordinates for the crystal structure of 5-carboxyl-2'-deoxycytidine in B-type DNA, DDD^{ca}.

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HEADER      ----
COMPND      ---
REMARK      3
REMARK      3 REFINEMENT.
REMARK      3   PROGRAM      : REFMAC 5.8.0049
REMARK      3   AUTHORS      : MURSHUDOV, SKUBAK, LEBEDEV, PANNU,
REMARK      3                       STEINER, NICHOLLS, WINN, LONG, VAGIN
REMARK      3
REMARK      3   REFINEMENT TARGET : MAXIMUM LIKELIHOOD
REMARK      3
REMARK      3 DATA USED IN REFINEMENT.
REMARK      3   RESOLUTION RANGE HIGH (ANGSTROMS) : 1.95
REMARK      3   RESOLUTION RANGE LOW  (ANGSTROMS) : 25.89
REMARK      3   DATA CUTOFF          (SIGMA(F)) : NONE
REMARK      3   COMPLETENESS FOR RANGE          (%) : 97.54
REMARK      3   NUMBER OF REFLECTIONS          : 4704
REMARK      3
REMARK      3 FIT TO DATA USED IN REFINEMENT.
REMARK      3   CROSS-VALIDATION METHOD          : THROUGHOUT
REMARK      3   FREE R VALUE TEST SET SELECTION : RANDOM
REMARK      3   R VALUE      (WORKING + TEST SET) : 0.22492
REMARK      3   R VALUE      (WORKING SET)       : 0.22129
REMARK      3   FREE R VALUE                    : 0.26721
REMARK      3   FREE R VALUE TEST SET SIZE      (%) : 8.0
REMARK      3   FREE R VALUE TEST SET COUNT     : 409
REMARK      3
REMARK      3 FIT IN THE HIGHEST RESOLUTION BIN.
REMARK      3   TOTAL NUMBER OF BINS USED          : 20
REMARK      3   BIN RESOLUTION RANGE HIGH          : 1.950
REMARK      3   BIN RESOLUTION RANGE LOW           : 2.000
REMARK      3   REFLECTION IN BIN (WORKING SET)    : 352
REMARK      3   BIN COMPLETENESS (WORKING+TEST) (%) : 99.48
REMARK      3   BIN R VALUE (WORKING SET)          : 0.331
REMARK      3   BIN FREE R VALUE SET COUNT         : 30
REMARK      3   BIN FREE R VALUE                   : 0.261
REMARK      3
REMARK      3 NUMBER OF NON-HYDROGEN ATOMS USED IN REFINEMENT.
REMARK      3   ALL ATOMS                          : 508
REMARK      3
REMARK      3 B VALUES.
REMARK      3   FROM WILSON PLOT (A**2) : NULL
REMARK      3   MEAN B VALUE (OVERALL, A**2) : 45.709
REMARK      3 OVERALL ANISOTROPIC B VALUE.
REMARK      3   B11 (A**2) : 0.97
REMARK      3   B22 (A**2) : 1.63
REMARK      3   B33 (A**2) : -2.60
REMARK      3   B12 (A**2) : 0.00
REMARK      3   B13 (A**2) : -0.00
REMARK      3   B23 (A**2) : -0.00
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REMARK 3
REMARK 3 ESTIMATED OVERALL COORDINATE ERROR.
REMARK 3 ESU BASED ON R VALUE (A): 0.195
REMARK 3 ESU BASED ON FREE R VALUE (A): 0.178
REMARK 3 ESU BASED ON MAXIMUM LIKELIHOOD (A): 0.129
REMARK 3 ESU FOR B VALUES BASED ON MAXIMUM LIKELIHOOD (A**2): 4.712
REMARK 3
REMARK 3 CORRELATION COEFFICIENTS.
REMARK 3 CORRELATION COEFFICIENT FO-FC : 0.967
REMARK 3 CORRELATION COEFFICIENT FO-FC FREE : 0.949
REMARK 3
REMARK 3 RMS DEVIATIONS FROM IDEAL VALUES COUNT RMS WEIGHT
REMARK 3 BOND LENGTHS REFINED ATOMS (A): 555 ; 0.009 ; 0.012
REMARK 3 BOND LENGTHS OTHERS (A): 266 ; 0.002 ; 0.020
REMARK 3 BOND ANGLES REFINED ATOMS (DEGREES): 855 ; 2.209 ; 1.307
REMARK 3 BOND ANGLES OTHERS (DEGREES): 632 ; 3.717 ; 3.000
REMARK 3 CHIRAL-CENTER RESTRAINTS (A**3): 72 ; 0.164 ; 0.200
REMARK 3 GENERAL PLANES REFINED ATOMS (A): 290 ; 0.022 ; 0.020
REMARK 3 GENERAL PLANES OTHERS (A): 112 ; 0.003 ; 0.020
REMARK 3
REMARK 3 ISOTROPIC THERMAL FACTOR RESTRAINTS. COUNT RMS WEIGHT
REMARK 3 SIDE-CHAIN BOND REFINED ATOMS (A**2): 555 ; 5.348 ; 4.691
REMARK 3 SIDE-CHAIN BOND OTHER ATOMS (A**2) : 554 ; 5.346 ; 4.692
REMARK 3 SIDE-CHAIN ANGLE OTHER ATOMS (A**2) : 856 ; 7.305 ; 7.043
REMARK 3 LONG RANGE B REFINED ATOMS (A**2) : 777 ; 8.229 ; 42.681
REMARK 3 LONG RANGE B OTHER ATOMS (A**2) : 777 ; 8.218 ; 42.680
REMARK 3
REMARK 3 NCS RESTRAINTS STATISTICS
REMARK 3 NUMBER OF NCS GROUPS : NULL
REMARK 3
REMARK 3 TWIN DETAILS
REMARK 3 NUMBER OF TWIN DOMAINS : NULL
REMARK 3
REMARK 3 TLS DETAILS
REMARK 3 NUMBER OF TLS GROUPS : NULL
REMARK 3
REMARK 3 BULK SOLVENT MODELLING.
REMARK 3 METHOD USED : MASK
REMARK 3 PARAMETERS FOR MASK CALCULATION
REMARK 3 VDW PROBE RADIUS : 1.20
REMARK 3 ION PROBE RADIUS : 0.80
REMARK 3 SHRINKAGE RADIUS : 0.80
REMARK 3
REMARK 3 OTHER REFINEMENT REMARKS:
REMARK 3 HYDROGENS HAVE BEEN ADDED IN THE RIDING POSITIONS
REMARK 3 U VALUES : REFINED INDIVIDUALLY
REMARK 3
LINK O3' DT A 8 P 5CC A 9 p
LINK O3' 5CC A 9 P DG A 10 p
LINK O3' DT B 20 P 5CC B 21 p
LINK O3' 5CC B 21 P DG B 22 p
CRYST1 24.250 41.340 66.410 90.00 90.00 90.00 P 21 21 21
SCALE1 0.041237 0.000000 0.000000 0.000000
SCALE2 -0.000000 0.024190 0.000000 0.000000

```

SCALE3	0.000000	-0.000000	0.015058	0.000000								
ATOM	1	O5'	DC	A	1	-7.052	-14.063	24.484	1.00	73.36	A	O
ATOM	2	C5'	DC	A	1	-5.905	-13.257	24.182	1.00	62.09	A	C
ATOM	3	C4'	DC	A	1	-6.351	-11.836	23.928	1.00	63.76	A	C
ATOM	4	C3'	DC	A	1	-6.443	-11.252	22.509	1.00	61.30	A	C
ATOM	5	O3'	DC	A	1	-7.673	-10.550	22.278	1.00	65.33	A	O
ATOM	6	C2'	DC	A	1	-5.444	-10.113	22.580	1.00	61.59	A	C
ATOM	7	C1'	DC	A	1	-5.574	-9.641	24.048	1.00	53.10	A	C
ATOM	8	O4'	DC	A	1	-5.848	-10.802	24.824	1.00	53.30	A	O
ATOM	9	N1	DC	A	1	-4.355	-9.007	24.567	1.00	48.71	A	N
ATOM	10	C6	DC	A	1	-3.134	-9.581	24.349	1.00	58.64	A	C
ATOM	11	C5	DC	A	1	-2.007	-8.978	24.749	1.00	60.56	A	C
ATOM	12	C4	DC	A	1	-2.131	-7.709	25.388	1.00	51.04	A	C
ATOM	13	N4	DC	A	1	-1.042	-7.051	25.785	1.00	54.76	A	N
ATOM	14	N3	DC	A	1	-3.321	-7.145	25.619	1.00	45.32	A	N
ATOM	15	C2	DC	A	1	-4.447	-7.752	25.191	1.00	47.51	A	C
ATOM	16	O2	DC	A	1	-5.566	-7.259	25.389	1.00	45.69	A	O
ATOM	17	P	ADG	A	2	-8.351	-10.327	20.812	0.60	49.70	A	P
ATOM	18	P	BDG	A	2	-9.153	-11.222	22.408	0.40	59.25	A	P
ATOM	19	OP1ADG	A	2	-9.000	-11.582	20.404	0.60	50.59	A	O	
ATOM	20	OP1BDG	A	2	-9.335	-11.666	23.809	0.40	58.44	A	O	
ATOM	21	O5'ADG	A	2	-9.518	-9.334	21.278	0.60	54.51	A	O	
ATOM	22	O5'BDG	A	2	-10.039	-9.931	22.140	0.40	58.67	A	O	
ATOM	23	C5'	DG	A	2	-9.415	-8.734	22.621	1.00	62.56	A	C
ATOM	24	C4'	DG	A	2	-10.317	-7.546	22.797	1.00	55.32	A	C
ATOM	25	C3'	DG	A	2	-10.782	-6.859	21.508	1.00	51.37	A	C
ATOM	26	O3'	DG	A	2	-11.857	-5.953	21.672	1.00	52.05	A	O
ATOM	27	C2'	DG	A	2	-9.616	-5.975	21.144	1.00	42.77	A	C
ATOM	28	C1'	DG	A	2	-8.949	-5.668	22.484	1.00	47.50	A	C
ATOM	29	O4'	DG	A	2	-9.434	-6.605	23.442	1.00	54.26	A	O
ATOM	30	N9	DG	A	2	-7.481	-5.744	22.467	1.00	36.58	A	N
ATOM	31	C4	DG	A	2	-6.637	-4.739	22.850	1.00	32.32	A	C
ATOM	32	C5	DG	A	2	-5.363	-5.227	22.631	1.00	40.20	A	C
ATOM	33	N7	DG	A	2	-5.407	-6.489	22.051	1.00	36.88	A	N
ATOM	34	C8	DG	A	2	-6.684	-6.752	21.966	1.00	43.75	A	C
ATOM	35	N3	DG	A	2	-7.009	-3.574	23.423	1.00	32.55	A	N
ATOM	36	C2	DG	A	2	-5.970	-2.825	23.730	1.00	34.57	A	C
ATOM	37	N2	DG	A	2	-6.163	-1.606	24.252	1.00	35.05	A	N
ATOM	38	N1	DG	A	2	-4.665	-3.214	23.532	1.00	32.46	A	N
ATOM	39	C6	DG	A	2	-4.266	-4.432	22.979	1.00	31.98	A	C
ATOM	40	O6	DG	A	2	-3.066	-4.703	22.873	1.00	35.68	A	O
ATOM	41	OP2ADG	A	2	-7.363	-9.714	19.886	0.60	48.06	A	O	
ATOM	42	OP2BDG	A	2	-9.354	-12.172	21.289	0.40	61.54	A	O	
ATOM	43	P	DC	A	3	-12.543	-5.279	20.386	1.00	61.14	A	P
ATOM	44	OP1	DC	A	3	-13.982	-5.060	20.730	1.00	60.08	A	O
ATOM	45	O5'	DC	A	3	-11.833	-3.871	20.239	1.00	47.64	A	O
ATOM	46	C5'	DC	A	3	-12.087	-2.899	21.230	1.00	42.68	A	C
ATOM	47	C4'	DC	A	3	-11.068	-1.816	21.039	1.00	40.14	A	C
ATOM	48	C3'	DC	A	3	-11.129	-1.226	19.639	1.00	38.58	A	C
ATOM	49	O3'	DC	A	3	-11.931	-0.043	19.680	1.00	52.48	A	O
ATOM	50	C2'	DC	A	3	-9.673	-0.960	19.280	1.00	43.61	A	C
ATOM	51	C1'	DC	A	3	-8.866	-1.593	20.411	1.00	33.96	A	C
ATOM	52	O4'	DC	A	3	-9.770	-2.425	21.129	1.00	34.90	A	O
ATOM	53	N1	DC	A	3	-7.760	-2.446	19.979	1.00	36.21	A	N
ATOM	54	C6	DC	A	3	-8.011	-3.649	19.379	1.00	39.52	A	C
ATOM	55	C5	DC	A	3	-7.016	-4.475	19.051	1.00	42.17	A	C

ATOM	56	C4	DC	A	3	-5.690	-4.053	19.354	1.00	37.92	A	C
ATOM	57	N4	DC	A	3	-4.652	-4.820	19.026	1.00	39.08	A	N
ATOM	58	N3	DC	A	3	-5.439	-2.886	19.939	1.00	31.81	A	N
ATOM	59	C2	DC	A	3	-6.454	-2.070	20.283	1.00	36.23	A	C
ATOM	60	O2	DC	A	3	-6.254	-0.969	20.820	1.00	33.44	A	O
ATOM	61	OP2	DC	A	3	-12.066	-5.955	19.157	1.00	53.75	A	O
ATOM	62	P	DG	A	4	-12.355	0.658	18.288	1.00	57.75	A	P
ATOM	63	OP1	DG	A	4	-13.630	1.365	18.529	1.00	61.40	A	O
ATOM	64	O5'	DG	A	4	-11.231	1.770	18.124	1.00	53.26	A	O
ATOM	65	C5'	DG	A	4	-11.068	2.713	19.204	1.00	52.38	A	C
ATOM	66	C4'	DG	A	4	-9.860	3.589	18.975	1.00	56.18	A	C
ATOM	67	C3'	DG	A	4	-9.904	4.367	17.663	1.00	51.80	A	C
ATOM	68	O3'	DG	A	4	-9.301	5.645	17.866	1.00	62.96	A	O
ATOM	69	C2'	DG	A	4	-9.116	3.493	16.709	1.00	52.63	A	C
ATOM	70	C1'	DG	A	4	-8.069	2.874	17.630	1.00	46.75	A	C
ATOM	71	O4'	DG	A	4	-8.652	2.787	18.919	1.00	46.49	A	O
ATOM	72	N9	DG	A	4	-7.628	1.542	17.268	1.00	44.50	A	N
ATOM	73	C4	DG	A	4	-6.351	1.057	17.419	1.00	43.27	A	C
ATOM	74	C5	DG	A	4	-6.405	-0.254	16.994	1.00	35.39	A	C
ATOM	75	N7	DG	A	4	-7.676	-0.572	16.539	1.00	39.85	A	N
ATOM	76	C8	DG	A	4	-8.370	0.520	16.726	1.00	47.00	A	C
ATOM	77	N3	DG	A	4	-5.294	1.745	17.897	1.00	40.90	A	N
ATOM	78	C2	DG	A	4	-4.188	1.017	17.875	1.00	36.18	A	C
ATOM	79	N2	DG	A	4	-3.039	1.533	18.309	1.00	35.32	A	N
ATOM	80	N1	DG	A	4	-4.137	-0.285	17.458	1.00	33.84	A	N
ATOM	81	C6	DG	A	4	-5.216	-1.017	16.987	1.00	37.56	A	C
ATOM	82	O6	DG	A	4	-5.059	-2.196	16.656	1.00	37.54	A	O
ATOM	83	OP2	DG	A	4	-12.244	-0.357	17.217	1.00	52.40	A	O
ATOM	84	P	DA	A	5	-9.212	6.689	16.649	1.00	73.45	A	P
ATOM	85	OP1	DA	A	5	-9.300	8.063	17.231	1.00	78.48	A	O
ATOM	86	OP2	DA	A	5	-10.136	6.219	15.562	1.00	64.08	A	O
ATOM	87	O5'	DA	A	5	-7.707	6.556	16.153	1.00	57.34	A	O
ATOM	88	C5'	DA	A	5	-6.685	6.812	17.124	1.00	60.56	A	C
ATOM	89	C4'	DA	A	5	-5.339	6.490	16.528	1.00	61.87	A	C
ATOM	90	C3'	DA	A	5	-5.085	7.143	15.171	1.00	60.08	A	C
ATOM	91	O3'	DA	A	5	-3.762	7.635	15.348	1.00	62.97	A	O
ATOM	92	C2'	DA	A	5	-5.181	5.986	14.195	1.00	55.49	A	C
ATOM	93	C1'	DA	A	5	-4.641	4.842	15.041	1.00	50.61	A	C
ATOM	94	O4'	DA	A	5	-5.183	5.068	16.324	1.00	53.78	A	O
ATOM	95	N9	DA	A	5	-5.028	3.496	14.610	1.00	39.67	A	N
ATOM	96	C4	DA	A	5	-4.212	2.386	14.555	1.00	35.52	A	C
ATOM	97	C5	DA	A	5	-5.016	1.348	14.116	1.00	33.44	A	C
ATOM	98	N7	DA	A	5	-6.311	1.794	13.877	1.00	36.91	A	N
ATOM	99	C8	DA	A	5	-6.272	3.067	14.207	1.00	43.13	A	C
ATOM	100	N3	DA	A	5	-2.907	2.313	14.874	1.00	34.17	A	N
ATOM	101	C2	DA	A	5	-2.439	1.076	14.666	1.00	35.80	A	C
ATOM	102	N1	DA	A	5	-3.087	-0.007	14.210	1.00	32.37	A	N
ATOM	103	C6	DA	A	5	-4.407	0.093	13.949	1.00	33.95	A	C
ATOM	104	N6	DA	A	5	-5.059	-0.986	13.516	1.00	31.51	A	N
ATOM	105	P	DA	A	6	-3.126	8.618	14.308	1.00	65.36	A	P
ATOM	106	OP1	DA	A	6	-2.767	9.852	15.053	1.00	65.12	A	O
ATOM	107	OP2	DA	A	6	-3.990	8.639	13.091	1.00	55.72	A	O
ATOM	108	O5'	DA	A	6	-1.745	7.905	13.937	1.00	54.98	A	O
ATOM	109	C5'	DA	A	6	-0.952	7.280	14.947	1.00	50.01	A	C
ATOM	110	C4'	DA	A	6	0.056	6.374	14.281	1.00	49.55	A	C
ATOM	111	C3'	DA	A	6	0.647	6.970	13.003	1.00	46.82	A	C

ATOM	112	O3'	DA	A	6	2.049	6.773	13.203	1.00	53.59	A	O
ATOM	113	C2'	DA	A	6	-0.045	6.196	11.885	1.00	44.53	A	C
ATOM	114	C1'	DA	A	6	-0.330	4.833	12.529	1.00	41.49	A	C
ATOM	115	O4'	DA	A	6	-0.562	5.119	13.897	1.00	46.07	A	O
ATOM	116	N9	DA	A	6	-1.506	4.113	12.019	1.00	35.31	A	N
ATOM	117	C4	DA	A	6	-1.593	2.766	11.740	1.00	34.49	A	C
ATOM	118	C5	DA	A	6	-2.905	2.543	11.355	1.00	29.46	A	C
ATOM	119	N7	DA	A	6	-3.642	3.720	11.419	1.00	35.23	A	N
ATOM	120	C8	DA	A	6	-2.775	4.614	11.838	1.00	36.22	A	C
ATOM	121	N3	DA	A	6	-0.609	1.854	11.801	1.00	36.17	A	N
ATOM	122	C2	DA	A	6	-1.047	0.661	11.408	1.00	36.35	A	C
ATOM	123	N1	DA	A	6	-2.267	0.303	10.995	1.00	31.68	A	N
ATOM	124	C6	DA	A	6	-3.238	1.238	10.961	1.00	30.29	A	C
ATOM	125	N6	DA	A	6	-4.456	0.867	10.580	1.00	31.37	A	N
ATOM	126	P	DT	A	7	3.154	7.216	12.109	1.00	58.14	A	P
ATOM	127	OP1	DT	A	7	4.426	7.459	12.845	1.00	53.73	A	O
ATOM	128	O5'	DT	A	7	3.392	5.884	11.240	1.00	50.01	A	O
ATOM	129	C5'	DT	A	7	3.829	4.630	11.835	1.00	48.02	A	C
ATOM	130	C4'	DT	A	7	3.712	3.520	10.814	1.00	43.46	A	C
ATOM	131	C3'	DT	A	7	4.361	3.864	9.472	1.00	48.85	A	C
ATOM	132	O3'	DT	A	7	5.419	2.927	9.344	1.00	63.44	A	O
ATOM	133	C2'	DT	A	7	3.259	3.716	8.426	1.00	42.75	A	C
ATOM	134	C1'	DT	A	7	2.252	2.831	9.141	1.00	39.37	A	C
ATOM	135	O4'	DT	A	7	2.329	3.226	10.512	1.00	43.92	A	O
ATOM	136	N1	DT	A	7	0.846	2.931	8.730	1.00	36.71	A	N
ATOM	137	C6	DT	A	7	0.187	4.141	8.759	1.00	40.24	A	C
ATOM	138	C5	DT	A	7	-1.103	4.287	8.446	1.00	38.65	A	C
ATOM	139	C5M	DT	A	7	-1.798	5.613	8.470	1.00	40.91	A	C
ATOM	140	C4	DT	A	7	-1.852	3.100	8.086	1.00	36.66	A	C
ATOM	141	N3	DT	A	7	-1.133	1.925	8.108	1.00	31.96	A	N
ATOM	142	C2	DT	A	7	0.202	1.775	8.365	1.00	34.29	A	C
ATOM	143	O2	DT	A	7	0.749	0.684	8.354	1.00	34.03	A	O
ATOM	144	O4	DT	A	7	-3.052	3.067	7.849	1.00	36.53	A	O
ATOM	145	OP2	DT	A	7	2.552	8.248	11.185	1.00	43.35	A	O
ATOM	146	P	DT	A	8	6.412	3.030	8.138	1.00	67.95	A	P
ATOM	147	OP1	DT	A	8	7.785	2.884	8.702	1.00	74.13	A	O
ATOM	148	O5'	DT	A	8	5.998	1.769	7.268	1.00	62.03	A	O
ATOM	149	C5'	DT	A	8	5.835	0.484	7.892	1.00	56.42	A	C
ATOM	150	C4'	DT	A	8	5.075	-0.405	6.938	1.00	57.99	A	C
ATOM	151	C3'	DT	A	8	5.658	-0.469	5.526	1.00	52.21	A	C
ATOM	152	O3'	DT	A	8	5.937	-1.856	5.336	1.00	60.60	A	O
ATOM	153	C2'	DT	A	8	4.553	0.071	4.621	1.00	46.93	A	C
ATOM	154	C1'	DT	A	8	3.302	-0.182	5.427	1.00	45.70	A	C
ATOM	155	O4'	DT	A	8	3.716	0.052	6.783	1.00	46.28	A	O
ATOM	156	N1	DT	A	8	2.137	0.708	5.164	1.00	41.19	A	N
ATOM	157	C6	DT	A	8	2.322	2.064	5.322	1.00	37.35	A	C
ATOM	158	C5	DT	A	8	1.344	2.968	5.199	1.00	37.20	A	C
ATOM	159	C7	DT	A	8	1.573	4.438	5.358	1.00	41.50	A	C
ATOM	160	C4	DT	A	8	0.021	2.496	4.886	1.00	38.03	A	C
ATOM	161	N3	DT	A	8	-0.102	1.123	4.737	1.00	34.39	A	N
ATOM	162	C2	DT	A	8	0.893	0.184	4.871	1.00	35.72	A	C
ATOM	163	O2	DT	A	8	0.689	-1.007	4.681	1.00	34.97	A	O
ATOM	164	O4	DT	A	8	-0.962	3.211	4.760	1.00	35.24	A	O
ATOM	165	OP2	DT	A	8	6.040	4.213	7.308	1.00	73.03	A	O
HETATM	166	O5C	5CC	A	9	4.207	1.575	1.530	1.00	54.21	A	O
HETATM	167	C5A	5CC	A	9	3.049	1.984	1.695	1.00	52.02	A	C

HETATM	168	O5A	5CC	A	9	2.734	3.175	1.909	1.00	46.46	A	O
HETATM	169	C5	5CC	A	9	2.042	0.921	1.541	1.00	43.46	A	C
HETATM	170	C4	5CC	A	9	0.638	1.279	1.382	1.00	37.98	A	C
HETATM	171	N4	5CC	A	9	0.172	2.555	1.437	1.00	38.12	A	N
HETATM	172	N3	5CC	A	9	-0.222	0.275	1.164	1.00	37.14	A	N
HETATM	173	C2	5CC	A	9	0.213	-1.003	1.155	1.00	38.84	A	C
HETATM	174	O2	5CC	A	9	-0.622	-1.888	0.954	1.00	40.39	A	O
HETATM	175	C6	5CC	A	9	2.476	-0.414	1.487	1.00	40.05	A	C
HETATM	176	N1	5CC	A	9	1.571	-1.383	1.250	1.00	45.45	A	N
HETATM	177	C1'	5CC	A	9	1.891	-2.820	1.316	1.00	41.48	A	C
HETATM	178	O4'	5CC	A	9	2.191	-2.944	2.699	1.00	45.28	A	O
HETATM	179	C2'	5CC	A	9	3.122	-3.361	0.619	1.00	48.49	A	C
HETATM	180	C3'	5CC	A	9	3.628	-4.428	1.591	1.00	53.39	A	C
HETATM	181	O3'	5CC	A	9	3.155	-5.730	1.276	1.00	62.46	A	O
HETATM	182	C4'	5CC	A	9	3.071	-4.057	2.947	1.00	57.68	A	C
HETATM	183	C5'	5CC	A	9	4.233	-3.674	3.865	1.00	57.58	A	C
HETATM	184	O5'	5CC	A	9	5.211	-2.881	3.178	1.00	57.19	A	O
HETATM	185	P	5CC	A	9	6.539	-2.386	3.956	1.00	62.68	A	P
HETATM	186	OP2	5CC	A	9	7.308	-3.617	4.286	1.00	62.34	A	O
HETATM	187	OP1	5CC	A	9	7.099	-1.200	3.191	1.00	60.71	A	O
ATOM	188	P	DG	A	10	3.669	-6.457	-0.061	1.00	69.97	A	P
ATOM	189	OP1	DG	A	10	3.588	-7.931	0.147	1.00	75.89	A	O
ATOM	190	O5'	DG	A	10	2.586	-6.046	-1.160	1.00	63.96	A	O
ATOM	191	C5'	DG	A	10	1.183	-6.326	-1.005	1.00	65.19	A	C
ATOM	192	C4'	DG	A	10	0.434	-5.640	-2.122	1.00	59.41	A	C
ATOM	193	C3'	DG	A	10	1.061	-5.778	-3.515	1.00	55.44	A	C
ATOM	194	O3'	DG	A	10	-0.019	-5.802	-4.431	1.00	64.52	A	O
ATOM	195	C2'	DG	A	10	1.776	-4.452	-3.725	1.00	53.34	A	C
ATOM	196	C1'	DG	A	10	0.769	-3.549	-3.045	1.00	46.38	A	C
ATOM	197	O4'	DG	A	10	0.427	-4.221	-1.855	1.00	50.21	A	O
ATOM	198	N9	DG	A	10	1.160	-2.189	-2.694	1.00	43.07	A	N
ATOM	199	C4	DG	A	10	0.280	-1.150	-2.522	1.00	35.88	A	C
ATOM	200	C5	DG	A	10	1.056	-0.065	-2.195	1.00	39.24	A	C
ATOM	201	N7	DG	A	10	2.393	-0.428	-2.091	1.00	41.95	A	N
ATOM	202	C8	DG	A	10	2.407	-1.696	-2.408	1.00	42.06	A	C
ATOM	203	N3	DG	A	10	-1.053	-1.207	-2.711	1.00	34.13	A	N
ATOM	204	C2	DG	A	10	-1.631	-0.024	-2.551	1.00	34.68	A	C
ATOM	205	N2	DG	A	10	-2.954	0.091	-2.730	1.00	29.21	A	N
ATOM	206	N1	DG	A	10	-0.951	1.118	-2.193	1.00	33.88	A	N
ATOM	207	C6	DG	A	10	0.427	1.183	-1.969	1.00	40.73	A	C
ATOM	208	O6	DG	A	10	0.950	2.248	-1.637	1.00	38.33	A	O
ATOM	209	OP2	DG	A	10	4.918	-5.793	-0.545	1.00	61.30	A	O
ATOM	210	P	DC	A	11	-0.388	-7.136	-5.177	1.00	67.48	A	P
ATOM	211	OP1	DC	A	11	-0.244	-8.256	-4.200	1.00	77.79	A	O
ATOM	212	O5'	DC	A	11	-1.956	-6.989	-5.434	1.00	66.75	A	O
ATOM	213	C5'	DC	A	11	-2.882	-6.528	-4.416	1.00	60.75	A	C
ATOM	214	C4'	DC	A	11	-3.916	-5.614	-5.041	1.00	60.57	A	C
ATOM	215	C3'	DC	A	11	-4.246	-5.907	-6.508	1.00	52.98	A	C
ATOM	216	O3'	DC	A	11	-5.669	-5.751	-6.541	1.00	51.48	A	O
ATOM	217	C2'	DC	A	11	-3.410	-4.877	-7.270	1.00	47.97	A	C
ATOM	218	C1'	DC	A	11	-3.413	-3.676	-6.324	1.00	44.23	A	C
ATOM	219	O4'	DC	A	11	-3.469	-4.230	-5.011	1.00	46.22	A	O
ATOM	220	N1	DC	A	11	-2.262	-2.747	-6.338	1.00	40.13	A	N
ATOM	221	C6	DC	A	11	-0.989	-3.243	-6.375	1.00	43.73	A	C
ATOM	222	C5	DC	A	11	0.073	-2.434	-6.267	1.00	40.04	A	C
ATOM	223	C4	DC	A	11	-0.174	-1.054	-6.023	1.00	39.18	A	C

ATOM	224	N4	DC	A	11	0.849	-0.212	-5.888	1.00	36.48	A	N
ATOM	225	N3	DC	A	11	-1.418	-0.554	-5.962	1.00	33.83	A	N
ATOM	226	C2	DC	A	11	-2.484	-1.380	-6.082	1.00	40.39	A	C
ATOM	227	O2	DC	A	11	-3.661	-0.962	-5.964	1.00	39.89	A	O
ATOM	228	OP2	DC	A	11	0.389	-7.125	-6.458	1.00	69.13	A	O
ATOM	229	P	DG	A	12	-6.554	-6.181	-7.813	1.00	59.59	A	P
ATOM	230	OP1	DG	A	12	-7.981	-6.293	-7.340	1.00	55.36	A	O
ATOM	231	O5'	DG	A	12	-6.457	-4.924	-8.788	1.00	62.07	A	O
ATOM	232	C5'	DG	A	12	-7.284	-3.759	-8.598	1.00	54.25	A	C
ATOM	233	C4'	DG	A	12	-6.816	-2.673	-9.534	1.00	52.34	A	C
ATOM	234	C3'	DG	A	12	-6.676	-3.089	-10.995	1.00	47.72	A	C
ATOM	235	O3'	DG	A	12	-7.864	-2.672	-11.658	1.00	49.53	A	O
ATOM	236	C2'	DG	A	12	-5.445	-2.332	-11.481	1.00	49.88	A	C
ATOM	237	C1'	DG	A	12	-4.947	-1.577	-10.250	1.00	46.38	A	C
ATOM	238	O4'	DG	A	12	-5.496	-2.265	-9.142	1.00	46.34	A	O
ATOM	239	N9	DG	A	12	-3.503	-1.554	-10.067	1.00	43.47	A	N
ATOM	240	C4	DG	A	12	-2.731	-0.493	-9.658	1.00	37.26	A	C
ATOM	241	C5	DG	A	12	-1.439	-0.984	-9.604	1.00	40.22	A	C
ATOM	242	N7	DG	A	12	-1.410	-2.338	-9.905	1.00	38.98	A	N
ATOM	243	C8	DG	A	12	-2.652	-2.634	-10.181	1.00	45.61	A	C
ATOM	244	N3	DG	A	12	-3.169	0.761	-9.394	1.00	32.26	A	N
ATOM	245	C2	DG	A	12	-2.177	1.560	-9.036	1.00	36.77	A	C
ATOM	246	N2	DG	A	12	-2.413	2.849	-8.772	1.00	30.64	A	N
ATOM	247	N1	DG	A	12	-0.865	1.155	-8.923	1.00	35.53	A	N
ATOM	248	C6	DG	A	12	-0.395	-0.120	-9.211	1.00	40.62	A	C
ATOM	249	O6	DG	A	12	0.805	-0.374	-9.093	1.00	38.59	A	O
ATOM	250	OP2	DG	A	12	-5.843	-7.286	-8.556	1.00	53.88	A	O
TER												
ATOM	251	O5'	DC	B	13	4.030	9.050	-9.830	1.00	76.36	B	O
ATOM	252	C5'	DC	B	13	3.184	9.719	-8.875	1.00	73.06	B	C
ATOM	253	C4'	DC	B	13	2.101	8.783	-8.380	1.00	66.23	B	C
ATOM	254	C3'	DC	B	13	1.734	8.711	-6.898	1.00	56.80	B	C
ATOM	255	O3'	DC	B	13	0.341	8.972	-6.761	1.00	72.67	B	O
ATOM	256	C2'	DC	B	13	1.929	7.244	-6.579	1.00	61.22	B	C
ATOM	257	C1'	DC	B	13	1.448	6.620	-7.892	1.00	55.30	B	C
ATOM	258	O4'	DC	B	13	2.004	7.436	-8.925	1.00	60.23	B	O
ATOM	259	N1	DC	B	13	1.870	5.223	-8.095	1.00	50.03	B	N
ATOM	260	C6	DC	B	13	3.185	4.867	-7.971	1.00	53.03	B	C
ATOM	261	C5	DC	B	13	3.564	3.587	-8.069	1.00	52.21	B	C
ATOM	262	C4	DC	B	13	2.543	2.611	-8.276	1.00	46.93	B	C
ATOM	263	N4	DC	B	13	2.859	1.328	-8.401	1.00	48.27	B	N
ATOM	264	N3	DC	B	13	1.255	2.944	-8.368	1.00	43.23	B	N
ATOM	265	C2	DC	B	13	0.882	4.236	-8.260	1.00	39.47	B	C
ATOM	266	O2	DC	B	13	-0.310	4.578	-8.318	1.00	42.49	B	O
ATOM	267	P	DG	B	14	-0.165	10.283	-5.990	1.00	74.71	B	P
ATOM	268	OP1	DG	B	14	-0.062	11.416	-6.949	1.00	65.40	B	O
ATOM	269	O5'	DG	B	14	-1.714	10.008	-5.768	1.00	68.31	B	O
ATOM	270	C5'	DG	B	14	-2.603	9.942	-6.887	1.00	61.31	B	C
ATOM	271	C4'	DG	B	14	-3.718	9.007	-6.512	1.00	56.78	B	C
ATOM	272	C3'	DG	B	14	-4.193	9.221	-5.078	1.00	55.19	B	C
ATOM	273	O3'	DG	B	14	-5.608	9.320	-4.977	1.00	65.23	B	O
ATOM	274	C2'	DG	B	14	-3.671	8.018	-4.322	1.00	59.93	B	C
ATOM	275	C1'	DG	B	14	-3.502	6.963	-5.387	1.00	53.16	B	C
ATOM	276	O4'	DG	B	14	-3.204	7.664	-6.579	1.00	51.71	B	O
ATOM	277	N9	DG	B	14	-2.396	6.047	-5.146	1.00	43.09	B	N
ATOM	278	C4	DG	B	14	-2.412	4.684	-5.314	1.00	35.28	B	C

ATOM	279	C5	DG	B	14	-1.135	4.262	-4.996	1.00	35.51	B	C
ATOM	280	N7	DG	B	14	-0.328	5.343	-4.668	1.00	38.78	B	N
ATOM	281	C8	DG	B	14	-1.110	6.380	-4.796	1.00	39.46	B	C
ATOM	282	N3	DG	B	14	-3.482	3.943	-5.661	1.00	30.87	B	N
ATOM	283	C2	DG	B	14	-3.190	2.650	-5.706	1.00	33.78	B	C
ATOM	284	N2	DG	B	14	-4.128	1.779	-6.037	1.00	30.07	B	N
ATOM	285	N1	DG	B	14	-1.946	2.128	-5.423	1.00	31.50	B	N
ATOM	286	C6	DG	B	14	-0.823	2.884	-5.085	1.00	39.15	B	C
ATOM	287	O6	DG	B	14	0.282	2.324	-4.900	1.00	33.23	B	O
ATOM	288	OP2	DG	B	14	0.441	10.297	-4.620	1.00	66.01	B	O
ATOM	289	P	DC	B	15	-6.271	9.590	-3.539	1.00	63.47	B	P
ATOM	290	OP1	DC	B	15	-7.522	10.358	-3.728	1.00	67.33	B	O
ATOM	291	O5'	DC	B	15	-6.811	8.158	-3.100	1.00	54.40	B	O
ATOM	292	C5'	DC	B	15	-7.562	7.326	-3.995	1.00	51.25	B	C
ATOM	293	C4'	DC	B	15	-7.441	5.901	-3.509	1.00	46.61	B	C
ATOM	294	C3'	DC	B	15	-7.792	5.746	-2.040	1.00	44.34	B	C
ATOM	295	O3'	DC	B	15	-9.179	5.459	-2.064	1.00	50.96	B	O
ATOM	296	C2'	DC	B	15	-6.931	4.590	-1.581	1.00	45.36	B	C
ATOM	297	C1'	DC	B	15	-5.858	4.469	-2.634	1.00	39.26	B	C
ATOM	298	O4'	DC	B	15	-6.069	5.472	-3.597	1.00	42.42	B	O
ATOM	299	N1	DC	B	15	-4.475	4.571	-2.179	1.00	40.04	B	N
ATOM	300	C6	DC	B	15	-3.914	5.750	-1.774	1.00	42.25	B	C
ATOM	301	C5	DC	B	15	-2.623	5.812	-1.414	1.00	44.06	B	C
ATOM	302	C4	DC	B	15	-1.867	4.599	-1.467	1.00	41.06	B	C
ATOM	303	N4	DC	B	15	-0.573	4.598	-1.137	1.00	41.98	B	N
ATOM	304	N3	DC	B	15	-2.401	3.454	-1.899	1.00	32.64	B	N
ATOM	305	C2	DC	B	15	-3.708	3.403	-2.226	1.00	38.81	B	C
ATOM	306	O2	DC	B	15	-4.246	2.358	-2.609	1.00	37.99	B	O
ATOM	307	OP2	DC	B	15	-5.184	10.006	-2.608	1.00	70.00	B	O
ATOM	308	P	DG	B	16	-9.973	5.257	-0.697	1.00	52.67	B	P
ATOM	309	OP1	DG	B	16	-11.411	5.558	-1.018	1.00	56.56	B	O
ATOM	310	O5'	DG	B	16	-9.765	3.692	-0.440	1.00	47.18	B	O
ATOM	311	C5'	DG	B	16	-10.466	2.729	-1.242	1.00	40.66	B	C
ATOM	312	C4'	DG	B	16	-9.991	1.342	-0.889	1.00	40.99	B	C
ATOM	313	C3'	DG	B	16	-10.429	0.853	0.490	1.00	50.57	B	C
ATOM	314	O3'	DG	B	16	-10.613	-0.547	0.310	1.00	53.07	B	O
ATOM	315	C2'	DG	B	16	-9.255	1.237	1.375	1.00	47.97	B	C
ATOM	316	C1'	DG	B	16	-8.093	0.971	0.448	1.00	46.26	B	C
ATOM	317	O4'	DG	B	16	-8.543	1.338	-0.859	1.00	44.71	B	O
ATOM	318	N9	DG	B	16	-6.887	1.731	0.755	1.00	43.12	B	N
ATOM	319	C4	DG	B	16	-5.599	1.271	0.673	1.00	37.18	B	C
ATOM	320	C5	DG	B	16	-4.808	2.327	1.069	1.00	40.97	B	C
ATOM	321	N7	DG	B	16	-5.580	3.449	1.334	1.00	42.08	B	N
ATOM	322	C8	DG	B	16	-6.808	3.036	1.181	1.00	44.90	B	C
ATOM	323	N3	DG	B	16	-5.229	0.015	0.344	1.00	43.74	B	N
ATOM	324	C2	DG	B	16	-3.921	-0.141	0.392	1.00	40.21	B	C
ATOM	325	N2	DG	B	16	-3.384	-1.332	0.079	1.00	35.64	B	N
ATOM	326	N1	DG	B	16	-3.045	0.851	0.766	1.00	34.17	B	N
ATOM	327	C6	DG	B	16	-3.411	2.149	1.123	1.00	39.04	B	C
ATOM	328	O6	DG	B	16	-2.543	2.974	1.440	1.00	33.50	B	O
ATOM	329	OP2	DG	B	16	-9.275	5.985	0.403	1.00	43.89	B	O
ATOM	330	P	DA	B	17	-10.873	-1.550	1.543	1.00	60.01	B	P
ATOM	331	OP1	DA	B	17	-12.021	-2.422	1.129	1.00	59.36	B	O
ATOM	332	OP2	DA	B	17	-10.942	-0.775	2.802	1.00	58.30	B	O
ATOM	333	O5'	DA	B	17	-9.497	-2.354	1.620	1.00	58.53	B	O
ATOM	334	C5'	DA	B	17	-8.789	-2.780	0.431	1.00	55.14	B	C

ATOM	335	C4'	DA	B	17	-7.675	-3.722	0.823	1.00	56.49	B	C
ATOM	336	C3'	DA	B	17	-8.026	-4.605	2.021	1.00	53.29	B	C
ATOM	337	O3'	DA	B	17	-7.341	-5.822	1.756	1.00	59.36	B	O
ATOM	338	C2'	DA	B	17	-7.490	-3.810	3.199	1.00	50.22	B	C
ATOM	339	C1'	DA	B	17	-6.229	-3.160	2.612	1.00	46.71	B	C
ATOM	340	O4'	DA	B	17	-6.455	-3.011	1.204	1.00	47.99	B	O
ATOM	341	N9	DA	B	17	-5.911	-1.842	3.151	1.00	37.32	B	N
ATOM	342	C4	DA	B	17	-4.658	-1.354	3.436	1.00	34.38	B	C
ATOM	343	C5	DA	B	17	-4.846	-0.048	3.848	1.00	35.45	B	C
ATOM	344	N7	DA	B	17	-6.197	0.263	3.882	1.00	39.75	B	N
ATOM	345	C8	DA	B	17	-6.783	-0.819	3.433	1.00	37.86	B	C
ATOM	346	N3	DA	B	17	-3.482	-1.977	3.263	1.00	29.34	B	N
ATOM	347	C2	DA	B	17	-2.472	-1.188	3.616	1.00	31.34	B	C
ATOM	348	N1	DA	B	17	-2.501	0.079	4.045	1.00	35.22	B	N
ATOM	349	C6	DA	B	17	-3.699	0.681	4.193	1.00	31.34	B	C
ATOM	350	N6	DA	B	17	-3.725	1.944	4.610	1.00	33.90	B	N
ATOM	351	P	DA	B	18	-7.303	-6.994	2.827	1.00	63.46	B	P
ATOM	352	OP1	DA	B	18	-7.241	-8.255	2.071	1.00	64.68	B	O
ATOM	353	OP2	DA	B	18	-8.370	-6.728	3.844	1.00	53.26	B	O
ATOM	354	O5'	DA	B	18	-5.901	-6.785	3.565	1.00	54.25	B	O
ATOM	355	C5'	DA	B	18	-4.671	-6.774	2.835	1.00	54.30	B	C
ATOM	356	C4'	DA	B	18	-3.551	-6.636	3.834	1.00	60.05	B	C
ATOM	357	C3'	DA	B	18	-3.744	-7.511	5.078	1.00	57.72	B	C
ATOM	358	O3'	DA	B	18	-2.496	-8.194	5.192	1.00	67.11	B	O
ATOM	359	C2'	DA	B	18	-4.102	-6.521	6.176	1.00	51.89	B	C
ATOM	360	C1'	DA	B	18	-3.362	-5.260	5.720	1.00	48.44	B	C
ATOM	361	O4'	DA	B	18	-3.453	-5.265	4.305	1.00	53.97	B	O
ATOM	362	N9	DA	B	18	-3.910	-3.989	6.189	1.00	35.82	B	N
ATOM	363	C4	DA	B	18	-3.190	-2.870	6.544	1.00	32.88	B	C
ATOM	364	C5	DA	B	18	-4.128	-1.902	6.843	1.00	28.49	B	C
ATOM	365	N7	DA	B	18	-5.412	-2.389	6.666	1.00	32.98	B	N
ATOM	366	C8	DA	B	18	-5.227	-3.623	6.257	1.00	31.86	B	C
ATOM	367	N3	DA	B	18	-1.852	-2.730	6.570	1.00	34.29	B	N
ATOM	368	C2	DA	B	18	-1.513	-1.502	6.968	1.00	37.34	B	C
ATOM	369	N1	DA	B	18	-2.302	-0.481	7.312	1.00	30.43	B	N
ATOM	370	C6	DA	B	18	-3.644	-0.644	7.228	1.00	32.56	B	C
ATOM	371	N6	DA	B	18	-4.435	0.371	7.544	1.00	32.31	B	N
ATOM	372	P	DT	B	19	-2.141	-9.149	6.471	1.00	74.44	B	P
ATOM	373	OP1	DT	B	19	-1.272	-10.207	5.964	1.00	64.28	B	O
ATOM	374	O5'	DT	B	19	-1.178	-8.232	7.345	1.00	65.35	B	O
ATOM	375	C5'	DT	B	19	-0.039	-7.620	6.735	1.00	55.18	B	C
ATOM	376	C4'	DT	B	19	0.486	-6.590	7.703	1.00	55.11	B	C
ATOM	377	C3'	DT	B	19	0.799	-7.186	9.080	1.00	51.36	B	C
ATOM	378	O3'	DT	B	19	2.185	-6.891	9.230	1.00	59.86	B	O
ATOM	379	C2'	DT	B	19	-0.143	-6.459	10.032	1.00	47.78	B	C
ATOM	380	C1'	DT	B	19	-0.350	-5.145	9.304	1.00	42.93	B	C
ATOM	381	O4'	DT	B	19	-0.470	-5.522	7.937	1.00	44.44	B	O
ATOM	382	N1	DT	B	19	-1.539	-4.383	9.648	1.00	36.97	B	N
ATOM	383	C6	DT	B	19	-2.776	-4.967	9.486	1.00	37.81	B	C
ATOM	384	C5	DT	B	19	-3.927	-4.333	9.728	1.00	32.22	B	C
ATOM	385	C5M	DT	B	19	-5.263	-4.992	9.583	1.00	35.85	B	C
ATOM	386	C4	DT	B	19	-3.862	-2.926	10.092	1.00	33.44	B	C
ATOM	387	N3	DT	B	19	-2.589	-2.398	10.209	1.00	29.18	B	N
ATOM	388	C2	DT	B	19	-1.411	-3.045	9.974	1.00	33.38	B	C
ATOM	389	O2	DT	B	19	-0.341	-2.496	10.117	1.00	31.82	B	O
ATOM	390	O4	DT	B	19	-4.822	-2.221	10.313	1.00	32.29	B	O

ATOM	391	OP2	DT	B	19	-3.370	-9.421	7.271	1.00	58.04	B	O
ATOM	392	P	DT	B	20	2.987	-7.355	10.533	1.00	73.37	B	P
ATOM	393	OP1	DT	B	20	4.336	-7.803	10.068	1.00	74.65	B	O
ATOM	394	O5'	DT	B	20	3.175	-5.996	11.345	1.00	61.52	B	O
ATOM	395	C5'	DT	B	20	3.754	-4.841	10.707	1.00	55.26	B	C
ATOM	396	C4'	DT	B	20	3.680	-3.682	11.670	1.00	53.84	B	C
ATOM	397	C3'	DT	B	20	4.217	-3.996	13.068	1.00	50.90	B	C
ATOM	398	O3'	DT	B	20	5.170	-2.965	13.327	1.00	62.87	B	O
ATOM	399	C2'	DT	B	20	2.975	-4.020	13.957	1.00	49.74	B	C
ATOM	400	C1'	DT	B	20	2.052	-3.048	13.241	1.00	44.10	B	C
ATOM	401	O4'	DT	B	20	2.305	-3.283	11.860	1.00	43.92	B	O
ATOM	402	N1	DT	B	20	0.606	-3.214	13.424	1.00	40.32	B	N
ATOM	403	C6	DT	B	20	0.025	-4.437	13.167	1.00	40.76	B	C
ATOM	404	C5	DT	B	20	-1.297	-4.648	13.177	1.00	39.81	B	C
ATOM	405	C7	DT	B	20	-1.902	-6.001	12.979	1.00	41.83	B	C
ATOM	406	C4	DT	B	20	-2.166	-3.516	13.405	1.00	35.39	B	C
ATOM	407	N3	DT	B	20	-1.519	-2.316	13.646	1.00	35.41	B	N
ATOM	408	C2	DT	B	20	-0.160	-2.088	13.624	1.00	35.76	B	C
ATOM	409	O2	DT	B	20	0.319	-0.984	13.824	1.00	38.20	B	O
ATOM	410	O4	DT	B	20	-3.395	-3.562	13.433	1.00	34.95	B	O
ATOM	411	OP2	DT	B	20	2.102	-8.234	11.359	1.00	57.95	B	O
HETATM	412	O5C	5CC	B	21	1.187	-5.336	16.493	1.00	46.32	B	O
HETATM	413	C5A	5CC	B	21	0.033	-4.955	16.609	1.00	43.90	B	C
HETATM	414	O5A	5CC	B	21	-0.946	-5.721	16.523	1.00	42.54	B	O
HETATM	415	C5	5CC	B	21	-0.146	-3.499	16.873	1.00	36.64	B	C
HETATM	416	C4	5CC	B	21	-1.454	-2.881	16.914	1.00	33.87	B	C
HETATM	417	N4	5CC	B	21	-2.580	-3.525	16.750	1.00	38.01	B	N
HETATM	418	N3	5CC	B	21	-1.587	-1.559	17.167	1.00	38.01	B	N
HETATM	419	C2	5CC	B	21	-0.490	-0.827	17.355	1.00	39.91	B	C
HETATM	420	O2	5CC	B	21	-0.602	0.420	17.593	1.00	41.01	B	O
HETATM	421	C6	5CC	B	21	0.959	-2.689	16.990	1.00	34.81	B	C
HETATM	422	N1	5CC	B	21	0.805	-1.414	17.288	1.00	40.67	B	N
HETATM	423	C1'	5CC	B	21	1.965	-0.507	17.400	1.00	42.36	B	C
HETATM	424	O4'	5CC	B	21	2.527	-0.579	16.053	1.00	46.37	B	O
HETATM	425	C2'	5CC	B	21	3.135	-0.872	18.301	1.00	48.67	B	C
HETATM	426	C3'	5CC	B	21	4.289	-0.241	17.532	1.00	50.17	B	C
HETATM	427	O3'	5CC	B	21	4.546	1.048	18.023	1.00	54.36	B	O
HETATM	428	C4'	5CC	B	21	3.871	-0.002	16.097	1.00	52.40	B	C
HETATM	429	C5'	5CC	B	21	4.900	-0.599	15.145	1.00	51.53	B	C
HETATM	430	O5'	5CC	B	21	5.208	-1.941	15.597	1.00	50.23	B	O
HETATM	431	P	5CC	B	21	6.014	-2.994	14.670	1.00	59.94	B	P
HETATM	432	OP2	5CC	B	21	7.400	-2.453	14.349	1.00	63.64	B	O
HETATM	433	OP1	5CC	B	21	5.844	-4.384	15.242	1.00	60.90	B	O
ATOM	434	P	DG	B	22	5.523	1.234	19.278	1.00	63.24	B	P
ATOM	435	OP1	DG	B	22	6.228	2.511	19.089	1.00	53.84	B	O
ATOM	436	O5'	DG	B	22	4.524	1.407	20.504	1.00	58.34	B	O
ATOM	437	C5'	DG	B	22	3.641	2.538	20.528	1.00	57.93	B	C
ATOM	438	C4'	DG	B	22	2.565	2.263	21.544	1.00	55.34	B	C
ATOM	439	C3'	DG	B	22	3.099	1.703	22.866	1.00	48.75	B	C
ATOM	440	O3'	DG	B	22	2.257	2.239	23.883	1.00	58.60	B	O
ATOM	441	C2'	DG	B	22	2.807	0.220	22.754	1.00	50.28	B	C
ATOM	442	C1'	DG	B	22	1.461	0.311	22.046	1.00	45.51	B	C
ATOM	443	O4'	DG	B	22	1.660	1.253	21.020	1.00	49.05	B	O
ATOM	444	N9	DG	B	22	0.921	-0.908	21.460	1.00	42.92	B	N
ATOM	445	C4	DG	B	22	-0.410	-1.118	21.205	1.00	35.01	B	C
ATOM	446	C5	DG	B	22	-0.493	-2.405	20.735	1.00	34.09	B	C

ATOM	447	N7	DG	B	22	0.764	-2.998	20.690	1.00	37.78	B	N
ATOM	448	C8	DG	B	22	1.568	-2.077	21.143	1.00	37.91	B	C
ATOM	449	N3	DG	B	22	-1.402	-0.225	21.409	1.00	32.49	B	N
ATOM	450	C2	DG	B	22	-2.583	-0.727	21.096	1.00	32.72	B	C
ATOM	451	N2	DG	B	22	-3.672	0.011	21.259	1.00	29.60	B	N
ATOM	452	N1	DG	B	22	-2.769	-1.994	20.611	1.00	32.87	B	N
ATOM	453	C6	DG	B	22	-1.759	-2.926	20.396	1.00	33.11	B	C
ATOM	454	O6	DG	B	22	-2.032	-4.038	19.923	1.00	33.81	B	O
ATOM	455	OP2	DG	B	22	6.238	-0.054	19.526	1.00	50.97	B	O
ATOM	456	P	DC	B	23	2.833	2.874	25.222	1.00	69.16	B	P
ATOM	457	OP1	DC	B	23	3.533	4.135	24.860	1.00	67.95	B	O
ATOM	458	O5'	DC	B	23	1.515	3.305	26.020	1.00	60.67	B	O
ATOM	459	C5'	DC	B	23	0.540	4.225	25.456	1.00	55.51	B	C
ATOM	460	C4'	DC	B	23	-0.848	3.744	25.799	1.00	49.85	B	C
ATOM	461	C3'	DC	B	23	-0.963	3.281	27.254	1.00	46.91	B	C
ATOM	462	O3'	DC	B	23	-1.810	4.080	28.095	1.00	62.85	B	O
ATOM	463	C2'	DC	B	23	-1.531	1.881	27.169	1.00	47.23	B	C
ATOM	464	C1'	DC	B	23	-1.892	1.663	25.711	1.00	44.13	B	C
ATOM	465	O4'	DC	B	23	-1.124	2.582	24.965	1.00	52.32	B	O
ATOM	466	N1	DC	B	23	-1.575	0.295	25.246	1.00	42.25	B	N
ATOM	467	C6	DC	B	23	-0.311	-0.216	25.354	1.00	40.65	B	C
ATOM	468	C5	DC	B	23	-0.039	-1.469	24.976	1.00	36.20	B	C
ATOM	469	C4	DC	B	23	-1.124	-2.255	24.495	1.00	37.38	B	C
ATOM	470	N4	DC	B	23	-0.924	-3.510	24.102	1.00	32.57	B	N
ATOM	471	N3	DC	B	23	-2.367	-1.775	24.423	1.00	36.06	B	N
ATOM	472	C2	DC	B	23	-2.624	-0.511	24.807	1.00	36.95	B	C
ATOM	473	O2	DC	B	23	-3.770	-0.035	24.765	1.00	34.27	B	O
ATOM	474	OP2	DC	B	23	3.475	1.785	26.011	1.00	68.51	B	O
ATOM	475	P	DG	B	24	-1.895	3.756	29.706	1.00	65.98	B	P
ATOM	476	OP1	DG	B	24	-2.219	5.008	30.422	1.00	73.34	B	O
ATOM	477	O5'	DG	B	24	-3.196	2.853	29.878	1.00	50.48	B	O
ATOM	478	C5'	DG	B	24	-4.458	3.294	29.395	1.00	43.21	B	C
ATOM	479	C4'	DG	B	24	-5.321	2.078	29.209	1.00	45.92	B	C
ATOM	480	C3'	DG	B	24	-5.315	1.187	30.445	1.00	43.48	B	C
ATOM	481	O3'	DG	B	24	-6.193	1.662	31.464	1.00	45.23	B	O
ATOM	482	C2'	DG	B	24	-5.735	-0.142	29.874	1.00	41.50	B	C
ATOM	483	C1'	DG	B	24	-5.165	-0.110	28.478	1.00	43.73	B	C
ATOM	484	O4'	DG	B	24	-4.737	1.236	28.188	1.00	42.19	B	O
ATOM	485	N9	DG	B	24	-4.037	-1.019	28.308	1.00	40.63	B	N
ATOM	486	C4	DG	B	24	-4.104	-2.263	27.734	1.00	37.01	B	C
ATOM	487	C5	DG	B	24	-2.807	-2.735	27.730	1.00	38.88	B	C
ATOM	488	N7	DG	B	24	-1.944	-1.810	28.299	1.00	37.54	B	N
ATOM	489	C8	DG	B	24	-2.718	-0.815	28.643	1.00	42.88	B	C
ATOM	490	N3	DG	B	24	-5.223	-2.863	27.267	1.00	32.29	B	N
ATOM	491	C2	DG	B	24	-4.959	-4.057	26.772	1.00	34.86	B	C
ATOM	492	N2	DG	B	24	-5.951	-4.819	26.308	1.00	31.79	B	N
ATOM	493	N1	DG	B	24	-3.700	-4.605	26.728	1.00	33.42	B	N
ATOM	494	C6	DG	B	24	-2.559	-4.030	27.258	1.00	37.91	B	C
ATOM	495	O6	DG	B	24	-1.479	-4.624	27.181	1.00	42.12	B	O
ATOM	496	OP2	DG	B	24	-0.776	2.839	30.100	1.00	58.28	B	O
TER												
HETATM	497	O	HOH	C	1	0.588	-3.711	5.454	1.00	47.03	C	O
HETATM	498	O	HOH	C	2	-4.986	3.972	19.066	1.00	48.74	C	O
HETATM	499	O	HOH	C	3	-0.804	-6.736	21.651	1.00	62.93	O	O
HETATM	500	O	HOH	C	4	3.639	-1.444	-6.010	1.00	43.57	O	O
HETATM	501	O	HOH	C	5	-1.918	-3.898	1.851	1.00	42.95	O	O

HETATM	502	O	HOH	C	6	9.979	-6.519	0.022	1.00	55.15	O
HETATM	503	O	HOH	C	7	1.705	1.415	13.189	0.50	28.51	O
HETATM	504	O	HOH	C	8	-1.325	3.757	16.259	0.50	29.57	O
HETATM	505	O	HOH	C	9	2.674	-3.617	7.388	0.50	33.17	O
HETATM	506	O	HOH	C	10	-7.307	-6.067	6.373	0.50	37.42	O
HETATM	507	O	HOH	C	11	-15.049	-3.165	17.560	0.50	46.43	O
HETATM	508	O	HOH	C	12	1.173	8.787	10.026	0.50	40.69	O

END

File A-4: Dictionary file for 5-carboxyl-2'-deoxycytidine.

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5CC      C4    C   CR6     0.000    13.391    1.297   132.751
5CC      N4    N   N        0.000    13.023    2.590   132.778
5CC      HN4   H   H        0.000    12.105    2.823   132.683
5CC      N3    N   NRD6    0.000    12.462    0.326   132.532
5CC      C2    C   CR6     0.000    12.852   -0.992   132.517
5CC      O2    O   O        0.000    11.979   -1.885   132.293
5CC      C6    C   CR16    0.000    15.218   -0.432   132.852
5CC      H6    H   H        0.000    16.250   -0.723   133.004
5CC      N1    N   NR6     0.000    14.260   -1.376   132.611
5CC      "C1'"  C   CH1     0.000    14.634   -2.814   132.670
5CC      "H1'"  H   H        0.000    13.766   -3.462   132.484
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5CC      "C2'"  C   CH2     0.000    15.902   -3.362   131.989
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5CC	"C5'"	C	CH2	0.000	17.055	-3.800	135.257
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5CC	N4	C4	HN4	.
5CC	HN4	N4	.	.
5CC	N3	C4	C2	.
5CC	C2	N3	O2	.
5CC	O2	C2	.	.
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5CC	"HO3'"	"O3'"	.	.
5CC	"C4'"	"C3'"	"C5'"	.
5CC	"H4'"	"C4'"	.	.
5CC	"C5'"	"C4'"	"O5'"	.
5CC	"H5'1"	"C5'"	.	.
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5CC	"O5'"	"C5'"	P	.
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5CC	N1	C2	.	ADD

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5CC	H6	C6	N1	120.000	3.000
5CC	C6	N1	"C1'"	120.000	3.000
5CC	C6	N1	C2	120.000	3.000
5CC	"C1'"	N1	C2	120.000	3.000
5CC	N1	"C1'"	"H1'"	109.470	3.000
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5CC	"H1'"	"C1'"	"O4'"	109.470	3.000
5CC	"H1'"	"C1'"	"C2'"	108.340	3.000
5CC	"O4'"	"C1'"	"C2'"	109.470	3.000
5CC	"C1'"	"O4'"	"C4'"	111.800	3.000
5CC	"C1'"	"C2'"	"H2'1"	109.470	3.000
5CC	"C1'"	"C2'"	"H2'2"	109.470	3.000
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5CC	"C3'"	"C4'"	"C5'"	111.000	3.000
5CC	"C3'"	"C4'"	"O4'"	109.470	3.000
5CC	"H4'"	"C4'"	"C5'"	108.340	3.000
5CC	"H4'"	"C4'"	"O4'"	109.470	3.000
5CC	"C5'"	"C4'"	"O4'"	109.470	3.000
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5CC	"C4'"	"C5'"	"H5'2"	109.470	3.000
5CC	"C4'"	"C5'"	"O5'"	109.470	3.000
5CC	"H5'1"	"C5'"	"H5'2"	107.900	3.000
5CC	"H5'1"	"C5'"	"O5'"	109.470	3.000
5CC	"H5'2"	"C5'"	"O5'"	109.470	3.000
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5CC	"O5'"	P	OP2	108.200	3.000
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5CC      CONST_7    C6      N1      C2      N3      0.000      0.000      0
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5CC      var_9      "C3'"   "C4'"   "C5'"   "O5'"   180.000     20.000      3
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REFERENCES

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